Did in

Herbal Medicine and Parasitic Diseases

Samia E. Etewa¹, Sherif M. Abaza²

Parasitology Department, Faculty of Medicine, Zagazig¹ and Suez Canal² Universities, Ismailia, Egypt

Received: August, 2010

ecepted: October, 2010

INTRODUCTION

Herbal medicine (also known as traditional, folk and alternative medicine) comprises medical knowledge systems that developed over generations within various societies before the era of modern medicine. WHO defines traditional medicine as the health practices, approaches, knowledge and beliefs incorporating plant, spiritual therapies and manual techniques, applied singularly or in combination to treat, diagnose and prevent illnesses or maintain well-being(1). Ancient Egyptian medicine, 1000 Before Christ (BC), was known to use garlic, opium, castor oil, coriander, mint and other herbs for medicinal purposes and Indian medicine used turmeric (Curcumin) possibly as early as 1900 BC(2). N. sativa seeds have been used as a traditional medicine for the treatment of a variety of sicknesses including parasitic diseases(3). The scope of herbal medicine sometimes extended to include fungal and bee products, as well as minerals, shells and certain animal parts⁽⁴⁾. It is estimated that 20,000 species of higher plants are used medicinally throughout the world. In some Asian and African countries, up to 80% of the population rely on traditional medicine for their primary health care needs(1).

The emergence of parasites resistant to current chemotherapies highlights the importance of plant essential oils as novel anti-parasitic agents; e.g., in schistosomiasis(5), malaria(6) and visceral leishmaniasis(7). Some plant oils have immunomodulatory effects that could modify hostparasite immunobiology and the lipid solubility of plant oils might offer alternative, transcutaneous delivery routes(8). In addition, the safety and use of plant essential oils in drug resistant cases are considered the most advantages. However, lack of proper understanding of plant and drug interactions have led to adverse reactions that are sometimes life threatening or lethal⁽⁹⁾. Furthermore, adulteration or counterfeit herbs can also be a health hazard; as WHO confirmed that inappropriate use of traditional medicines or practices can have negative or dangerous effects(1).

The present review is an analysis to through light on different herbs with their active compunds that could be used as drug targets in parasitic diseases. It also discusses some herbs with repellent activity and molluscicidal effects.

Keywords: Herbal Medicine, Parasitic Diseases, Treatment, Repellents, Molluscicides.

Corresponding author: Sherif M. Abaza, smabaza@hotmail.com

Examples of Known Herbs with Antiparasitic Effects

Allium cepa; Onion (Al-Basal)

Two sets of compounds make up the majority of onion's known active constituents: Sulphur compounds such as allicin and allyl propyl disulphide and flavonoids such as quercetin. Allicin exhibits its antimicrobial action mainly by immediate and total inhibition of RNA, DNA and protein synthesis⁽¹⁰⁾.

In a study conducted in 2004, all tested strains of *Leishmania* (*major*, *donovani* and *mexicana*) were found to be sensitive *in vitro* to the onion juice in the promastigote stage⁽¹¹⁾. Aqueous onion extract is similar to metronidazole in inhibiting both multiplication and motility of *T. vaginalis*⁽¹²⁾. *A. cepa* oil has an antihelminthic effect in rats

experimentally infected with *T. spiralis* and increases the production of antibodies generated during life cycle of the parasite⁽¹³⁾. Recently, effectiveness of *A. Cepa* extract in eyelids inflammation caused by *Demodex folliculorum* was tested with promising results⁽¹⁴⁾. Onion reportedly exerted different physiological changes in snails of *B. alexandrina* leading to decrease in glucose and glycogen which are the snail energy fuel and inhibited the phenol oxidase enzyme which led to disturbance in egg-shell formation⁽¹⁵⁾. Some adverse effects have been reported where higher intakes may worsen existing heartburn and there are also isolated reports of allergy manifesting as skin rash and red, itchy eyes⁽¹⁶⁾.

Allium sativum; Garlic (Al-Tom)

The active constituents of garlic exceed 200 chemicals. It contains sulphur compounds (allicin, alliin and agoene), volatile oils, enzymes (allinase, peroxidase and miracynase), carbohydrates (sucrose and glucose), minerals (selenium), amino acids such as cysteine, glutamine, isoleucine and methionine, bioflavonoids such as quercetin and cyanidin, Allistatin I and Allistatin II and vitamins A, B1, B2, C, E and niacin⁽¹⁷⁾. The main antimicrobial effect of allicin is due to its chemical reaction with thiol groups of various enzymes e.g. alcohol dehydrogenase and DNA polymerase which affect the essential metabolism of cysteine proteinase activity involved in the virulence of parasites⁽¹⁸⁾. Garlic appears to be safe during pregnancy and breastfeeding. Some sensitive people may experience heartburn and flatulence(19).

Antiprotozoal effects: Several studies were conducted to investigate the efficacy of garlic in giardiasis. There were reports of complete relief of clinical symptoms within 36 hours⁽²⁰⁾, loss of flagellar movement and motility with trophozoites swelling leading to decreased ability to adhere to host cells⁽²¹⁾, inhibition of *Giardia*'s cysteine proteases activity and excretory/secretory products resulting in reduction of GIT symptoms⁽²²⁾. The therapeutic effect of allicin was evaluated against *C. cayetanensis* and the results showed that Tomex (10 mg/ Kg/day of garlic extract) caused significant reduction in oocyst shedding and improvement of intestinal pathological changes (100% in immunocompetent and 80% in immunosuppressed mice)⁽²³⁾.

The circumsporozoite protein (CSP), which is the major surface protein of *Plasmodia* sporozoites, is proteolytically processed by a parasite-derived cysteine protease and this event is temporally associated with sporozoite invasion of host cells. At low concentrations, allicin inhibited CSP processing and prevented sporozoite invasion of host cells *in vitro*. *In vivo*, mice injected with allicin had decreased *Plasmodia* infections compared to controls. Allicin was also tested on erythrocytic stages where a 4-day regimen of allicin administered either orally or intravenously significantly increased the survival of infected mice by 10 days⁽²⁴⁾.

In mice infected with *L. mexicana*, intraperitoneal injection of garlic extract or its protein fraction augmented parasite engulfiment and destruction of intracellular amastigotes by macrophages. Modulation of the immune response through macrophages activations was postulated as a mechanism of action of garlic⁽²⁵⁾. Another explanation claimed that garlic extract reduced macrophage infection through induction of nitric oxide (NO) production *in vitro*. Therefore, it may act on both T cells and macrophages to stimulate IFN-γ production and NO synthesis for parasite killing⁽²⁶⁾.

In a study conducted in 2007, *Eimeria stiedae* induced marked hepatic histopathological alterations in rabbits

that were not treated with garlic, versus those treated with garlic. The effect of methanol garlic extract in a concentration of 3.90 mg/ml, on the growth of *Acanthamoeba castellanii* and its cytotoxicity on corneal cells was studied *in vitro*. These findings indicated its amoebicidal, as well as cysticidal effects on trophozoites and cysts, with no cytotoxicity to corneal cells⁽²⁸⁾. In addition, aqueous garlic extract proved similar to metronidazole in inhibiting both multiplication and motility of *T. vaginalis*⁽¹³⁾.

Antihelminthic effects: Aqueous garlic extract, with its potent free radical scavenging and antioxidant properties, seemed to be a highly promising agent in protecting hepatic tissue against oxidative damage due to *S. mansoni* infection⁽²⁹⁾. Garlic efficacy was highest in the group treated with garlic before and after bilharzial infection and resulted in various ultrastructural alterations in the surviving adult worm's tegument⁽³⁰⁾. Results of a study conducted in 2008 showed that chloroform garlic extract exerted the highest effect on the viability of hydatid cyst protoscoleces *in vitro*, in comparison with garlic aqueous extract and hydroalcoholic garlic extract⁽³¹⁾.

Garlic as repellent and its acaricidal effect: The repellent effect of garlic oil was evaluated against female *Phlebotomus* bite. A dose dependent anti-feeding effect, which was 100% effective at 1% concentration, was obtained⁽³²⁾. Topical application of 10% garlic juice by spraying, effectively decreased Northern fowl mite (NFM, *Ornithonyssus sylviarum*) infestation in laying hens. Humans could be bitten by these mites and suffer irritation and allergic reactions⁽³³⁾.

Garlic as molluscicide: The effect of the plant-derived molluscicides *Annona squamosa* and *Lawsonia inermis* combined with *A. sativum* on the reproduction of the snail *Lymnaea* was studied. A significant reduction in the fecundity, hatchability and survival of young snails was observed⁽³⁴⁾.

Artemisia (Al-Sheeh)

There are about 300 species in the *Artemisia* genus. Two of them, sweet annie (*Artemisia annua*) and wormwood (*Artemisia absinthium*), are of medical importance in treatment of some parasitic diseases. The active constituents in the first one are artesunate, dihydroartemisinin, artemether and arteether, while the second contains aromatic oils (thujone and isothujone) and strong bitter agents (absinthin and anabsinthin). Adverse effects associated with artemether treatment were abdominal pain, anorexia, nausea, vomiting, diarrhoea and CNS involvement (headache and dizziness)⁽³⁵⁾. Longer-term use of wormwood (over 4 weeks) can cause nausea, vomiting, insomnia, restlessness, vertigo, tremors and seizures⁽³⁶⁾. In addition, it is not recommended during pregnancy and breast-feeding⁽³⁷⁾.

derivatives: Artemisinin (ART) administration schemes in malaria control should ideally use more than one drug, preferably combinations including a rapid-acting schizonticidal drug, such as ART, as well as primaquine that can kill sexual and liver stages to prevent transmission. This could be considered as an alternative strategy for malaria control, and in combination with other anti-malarial measures may provide a tool for malaria elimination and eradication(38). ART combination therapy is the first-line treatment for uncomplicated P. falciparum malaria in most malaria-endemic countries(39). All ART compounds induce very rapid reduction of parasitaemia, starting almost immediately after administration. The proposed mechanism of action involves cleavage of endoperoxide bridges by iron producing free radicals which damage biological macromolecules causing oxidative stress in parasitized cells⁽⁴⁰⁾. Pure artemisinin has a low solubility in water, oil and therefore it could be administered orally, rectally and intramuscularly. Intramuscular artesunate is easier to administer but is associated with hypoglycaemia⁽⁴¹⁾. Unfortunately, oral administration is often not possible in patients with severe malaria, due to extreme vomiting; hence the development of several semi-synthetic ART derivatives(42).

Artemisinin also showed anti-leishmanial activity for both promastigotes and amastigotes and was accompanied by a high safety index. The leishmanicidal activity of artemisinin was mediated *via* apoptosis as evidenced by externalization of phosphatidylserine, loss of mitochondrial membrane potential and cell-cycle arrest. These data indicated promising anti-leishmanial activity mediated by programmed cell death⁽⁴³⁾.

Moreover, several ART derivatives were shown to be effective against *T. gondii in vitro*⁽⁴⁴⁾ as well as *in vivo* in a murine model of reactivated toxoplasmosis⁽⁴⁵⁾. The mechanism of action is uncertain. However, it was suggested that they may inhibit sarcoplasmic-endoplasmic reticulum (ATPase), thus disrupting calcium homeostasis by increasing the periodicity of calcium oscillations and inducing recurrent, strong calcium spikes⁽⁴⁴⁾. In addition, these new ART derivatives have the ability to inhibit multiple steps of *T. gondii* lytic cycle; they effectively inhibited *T. gondii* growth, tachyzoite replication, attachment to and invasion of host cells⁽⁴⁶⁾.

The ART derivatives were shown to possess a broad spectrum of activities against several helminthic infections. Combination between artemether and praziquantel showed highest efficacies against juvenile and adult worms of *S. haematobium*⁽⁴⁷⁾. Scanning and transmission microscopic observations indicated that artemether induces extensive damage to juveniles and adults of different *Schistosoma spp.*⁽⁴⁸⁾. In addition, significant progress has been made with artemether use for chemoprophylaxis in schistosomiasis, as immature stages of *S. mansoni* are more prone to oxidative

killing than mature worms which probably participates in the mechanism of anti-schistosomal action of ART⁽⁴⁹⁾. The efficacy and safety of artemether was studied in sheep infected with *Fasciola spp.* A single intramuscular dose of artemether significantly reduced both egg and worm burdens. There were no adverse events, however, two abortions were observed 7 days post treatment⁽⁵⁰⁾. Recently, a study was carried out to determine the morphological changes to *F. hepatica* after *in vivo* treatment with artemether and the results showed increased disruption of the tegumental system, with isolated patches of surface blebbing and reduced production of secretory bodies by the tegumental cells⁽⁵¹⁾.

Wormwood: *A. absinthium* extracts were found to be effective against the enteral (adult) and parenteral (larva) phases of trichinellosis⁽⁵²⁾. In addition, oil extracts were tested in the laboratory against host-seeking nymphs of *Ixodes ricinus Linnaeus* (*Acari: Ixodidae*). The results showed 62-70% repellent ability and hence could be used in control of arthropods of medical, veterinary or agricultural importance⁽⁵³⁾.

Commiphora molmol; Myrrh (Al-Mor)

All the three main constituents of myrrh; resin, gum and volatile oil are important in myrrh's activity as herbal medicine(54). The mechanism of the anti-schistosomal action of myrrh on S. mansoni is not fully understood. However, it has been attributed to permanent muscle paralysis of the worms leading to their shift to the liver where destruction takes place(55). Meanwhile, results of studies published on the efficacy of myrrh in the treatment of S. mansoni-infected mice are greatly conflicting. Several experimental and clinical studies reported significant parasite reductions and marked ultrastructural changes after treatment of S. mansoni-infected mice with Mirazid®(56,57) or under field conditions(58). However, other studies negated myrrh efficacy in the treatment of schistosomiasis in either experimental⁽⁵⁹⁾, or clinical studies⁽⁶⁰⁾.

There is also such debate over the efficacy of Mirazid® for the treatment of fascioliasis. Complete cure rate was achieved in experimentally infected rabbits with oral dose of 20 mg/day for 6 consecutive days⁽⁶¹⁾. On the contrary, results obtained for experimentally infected sheep treated with Mirazid® appeared similar to those for untreated infected animals⁽⁶²⁾. On the other hand, Mirazid® was reported to be effective against *D. dendriticum*⁽⁶³⁾, *H. heterophyes*⁽⁶⁴⁾ and *H. nana*⁽⁶⁵⁾. Regarding its antiprotozoal efficacy, Mirazid® was effective against cryptosporidiosis⁽⁶⁶⁾ and trichomoniasis in metronidazole-resistant infected females⁽⁶⁷⁾.

Myrrh proved to have insecticidal activity against mosquito larvae of *Culex pipiens* and *Aedes caspius*⁽⁶⁸⁾. In addition, the molluscicidal activity of myrrh was evaluated in several studies on *B. alexandrina*, *E. truncatus* and *L. cailliaudi* and the results revealed significant lethal and ovicidal effects^(69,70).

Curcuma longa; Turmeric (Al-Korkom or Curcumin)

As turmeric is a very cheap, easily available, effective and acceptable mode of treatment in developing countries, with no toxic or adverse reaction, it was used as paste for the treatment of scabies, with 97% cure rate within 3-15 days of treatment⁽⁷¹⁾. The active constituent known as curcumin showed a wide range of therapeutic actions. When used in the recommended amounts, turmeric is generally safe. Some herbal books recommend not taking high amounts of turmeric during pregnancy as it may cause uterine contractions and patients with gallstones or bile passages obstruction are advised to consult their healthcare practitioner before using turmeric⁽⁷²⁾.

Curcumin had a significant effect on the progression of experimental cerebral malaria. Survival of treated mice was significantly increased and development of cerebral malaria was either delayed or prevented⁽⁷³⁾. Possessing antiprotozoal effects, turmeric as well proved to (1) show cytotoxicity against *T. brucei*⁽⁷⁴⁾, (2) have leishmanicidal activity *in vitro*⁽⁷⁵⁾, (3) inhibit *G. lamblia* trophozoite growth and adhesion, by more than 50%, in dose and time dependent manner⁽⁷⁶⁾ and (4) be an attenuating agent against *T. gondii* tachyzoites in the peritoneal fluid of mice⁽⁷⁷⁾.

Studies were designed to evaluate the schistosomicidal activity of curcumin in vivo as well as immunomodulation of granulomatous inflammation in acute schistosomiasis mansoni. It was found that praziquantel was more effective in lowering worm burden while curcumin was more potent in reducing egg count⁽⁷⁸⁾. The mechanism by which curcumin might function as a schistosomicidal agent is unclear and needs further investigation. In another study, curcumin was effective in reducing both male and female worms and tissue-egg burdens, hepatic granuloma volume and liver collagen content. In addition, curcumin treatment modulates cellular and humoral immune responses of infected mice(79). On the other hand, curcumin is effective in the treatment of many inflammatory diseases. Its efficacy on reducing the histopathological changes of opisthorchiasis in hamsters was evaluated. Results showed reduced inflammatory cells aggregation surrounding the hepatic bile ducts, leading to reduced risk factors of cholangiocarcinoma development(80). For molluscicidal activity, turmeric was evaluated against B. alexandrina and the results showed that its 30 ppm extract stopped snail egg laying and caused deformity of all embryonic stages⁽⁸¹⁾.

Nigella sativa; Nigella (Black Seed or Habbat Al-Barakah)

Nigella oil contains saponin, nigellidine and nigellone and its seeds contain thymoquinone, monoterpenes such as p-cymene and α -pinene⁽⁸²⁾. Several mechanisms were described for the activity of black seeds such as

its protection against hepatotoxicity⁽⁸³⁾, its antioxidant role⁽⁸⁴⁾ and stimulation of immune system⁽⁸⁵⁾. The results of the studies conducted in 2007 proved that its aqueous extract could be useful in the treatment of *B. hominis*⁽⁸⁶⁾ and as an anti-malarial drug⁽³⁾. In the following year, its inhibitory effect against *T. vaginalis* was investigated *in vitro* and the results showed remarkable inhibition of trophozoites growth and motility⁽⁸⁷⁾.

The antioxidant and anti-schistosomal activities of *N. sativa* oil was studied in control and *S. mansoni*-infected mice. Remarkable reduction in worms, tissue eggs and alteration in oogram pattern were recorded in all the treated groups. The data pointed to the importance of these seeds as a promising agent to complement schistosomiasis specific treatment⁽²⁹⁾. Its antiparasitic effect was studied also in *H. nana*-infected mice, where it was found to reduce the infection starting from 2nd day of the treatment⁽⁸⁵⁾. Moreover, its prophylactic treatment prior to *T. spiralis* infection was effective against both adult worms and muscle larval count in infected rats⁽¹⁴⁾.

Zingiber officinale; Ginger (Ganzabil)

The main constituents of ginger are the gingerols, shogaols, zingerone and paradol(88). The anti-amoebic effect of a crude drug formulation against E. histolytica was studied in comparison to metronidazole. Results showed that the product had a minimal inhibitory concentration of 1000 mg/ml as compared with 10 mg/ ml for metronidazole⁽⁸⁹⁾. In vitro, a concentration of 200 mg of ginger extract killed almost all Schistosoma worms within 24 hrs⁽⁹⁰⁾. The anti-helminthic activity of ginger was reported also in sheep naturally infected with mixed species of GIT nematodes e.g. T. colubriformis, Strongyloides papillosus and Trichuris ovis⁽⁹¹⁾. Another study conducted in 2006 showed that there was significant reduction in intestinal adult worms and muscle larval count in mice infected with T. Spiralis⁽⁹²⁾. In addition, there was microfilaricidal activity of aqueous extracts of ginger when administered to dogs infected with D. immitis filarids(93).

Ginger essential oils proved to have repellent results against host-seeking chiggers (scrub typhus) of Leptotrombidium imphalum⁽⁹⁴⁾ and ovicidal and repellent effects against different mosquitoes as Anopheles stephensi, Aedes aegypti and Culex quinquefasciatus⁽⁹⁵⁾. In addition, its petroleum ether extract has larvicidal activity against A. aegypti and C. Quinquefasciatus⁽⁹⁶⁾.

As a molluscicidal agent, gingerol and shogaol exhibited potent effects on *Biomphalaria glabrata*, indicating their ability to interrupt *Schistosoma* transmission⁽⁹⁷⁾. Results of *in vivo* exposure of *Lymnaea acuminata* to gingerol proved its effect on snail neurotransmission mechanisms, either separately or through a complex interaction between the different neurotransmitters⁽⁹⁸⁾.

Berberis vulgaris, Berberis Aristata; Barberry

The active constituent in barberry is berberine which is an alkaloid having antibacterial, anti-amoebic, antifungal, anti-helminthic and leishmanicidal properties. Its reported main side effect was interference with normal liver function in infants. Strong standardized extracts may cause stomach upset and should be used for no more than two weeks continuously. Other symptoms of excessive berberine intake include lethargy, nose bleed, skin and eye irritation and kidney irritation⁽⁷¹⁾.

Berberine is a useful drug for the treatment of visceral as well as cutaneous leishmaniasis. In L. donovani infection, it inhibited in vitro multiplication of amastigotes in macrophage culture and their transformation to promastigotes in cell free culture (99), while 1% berberine sulphate inoculated intra-lesionally was found to be highly effective against cutaneous leishmaniasis in domestic dogs(100). In addition, crude extract formulation including berberine had a maximum cure rate of 73% at a dose of 800 mg/kg/day in hepatic amoebiasis(101). The results of a study conducted in patients with chloroquine-resistant malaria indicated that berberine was more effective in clearing the parasite than both tetracycline and cotrimoxazole and that the combination of pyrimethamine and berberine gave the best results for chloroquine resistant malaria(102). Moreover, berberine sulphate was comparable to metronidazole as regards potency on the growth of T. vaginalis in vitro $^{(103)}$.

Azadirachta indica, Melia Azadirachta; Neem

The major active constituents in neem are fatty acids and terpenoids such as azadirachtin, which is considered to possess anti-microbial and insect repellent effects, among many other actions(104). Neem seed oil is problematic and should be kept out of reach of children⁽¹⁰⁵⁾. In this form, it is mainly used in control of vector borne parasitic diseases. The larvicidal and emergence inhibitory activities of neem were tested successfully against the vectors of malaria, filariasis and dengue fever(106). Besides, water-free neem seed extract shampoo was effective against S. scabiei-infesting dogs in Egypt(107). As an acaricidal agent, neem extracts showed high level of efficacy (80%) for adult ticks after 5 hours of treatment(108). As a pediculicide, its extract shampoo proved to be highly effective against all stages of head lice, with no side effects(109). The significant repellent activities of neem against Phlebotomus papatasi(110) and *Ixodes ricinus*(111) nymphs were reported.

Cinchona officinalis; Cinchona (Al-Quina tree)

The medicinally active constituents of cinchona are a variety of alkaloids as quinine and quinidine. Oral quinine has marked side effects, including tinnitus, dizziness and nausea⁽¹¹²⁾. Quinine has been used as

malaria treatment for more than 350 years in Africa and remains the first-line anti-malarial drug for the treatment of complicated malaria in Europe and Africa. Children with uncomplicated malaria are generally treated with oral medication, except those unable to take oral drugs(113). It has substantial disadvantages mainly its poor tolerability, the long treatment course and the unpleasant bitter taste of its tablets(114). Therefore, it is no longer recommended by the WHO as first line treatment for malaria and should be used only when artemesinins are not available(115). Treatment of S. mansoni-infected female mice with daily intraperitoneal injections of quinine and quinidine caused significant decrease in worm burden and egg production. Their schistosomicidal effects are due to their to interfere with hemozoin formation which is the main heme detoxification pathway in S. mansoni⁽¹¹⁶⁾. In addition, quinine and chloroquine are considered potential drugs against L. loa infection(117).

Triticum vulgare, Triticum aestivum; Wheat germ (Ganin Habit Al-Kamh)

Lectin (wheat germ agglutinin) is the active constituent, however, it is a concentrated source of vitamin E, folic acid, phosphorus, thiamin, zinc and magnesium, as well as essential fatty acids and fatty alcohols(71). In a clinical trial, wheat germ helped in quick resolution of symptoms in Giardia-infected patients. It did not kill the parasites, but prevented their growth, replication and attachment(118). In addition, the effectiveness of wheat germ as a useful dietary supplement administered to E. histolytica-infected patients was reported, both in acute and chronic stages of infection(119). When its therapeutic effect was experimentally evaluated in immunosuppressed mice infected with Cryptosporidium oocysts, it showed significant reduction of excreted oocysts on day 7 post-infection(120). It was also shown to have remarkable inhibitory effect on multiplication and motility of T. Vaginalis⁽⁸⁷⁾.

Cinnamomum zeylanicum; Cinnamon (Al-Kerfa)

Various terpenoids (eugenol and cinnamaldehyde) are believed to account for cinnamon's medicinal effects⁽¹²¹⁾. Some people develop bronchial constriction or skin rash after exposure to cinnamon and chronic use of the concentrated oil may cause mouth inflammation. Cinnamon is not recommended for use by pregnant women⁽⁷¹⁾. Cinnamon essential oil had potent insecticidal and ovicidal activities against human head louse, *P. humanus capitis*⁽¹²²⁾, acaricidal activity against adult *Dermanyssus gallinae*⁽¹²³⁾ and strong larvicidal activity against 4th instar *Aedes aegypti* larvae⁽¹²⁴⁾. Recently, its inhibitory action against other mosquito species as *Aedes albopictus*, *Culex quinquefasciatus* and *Armigeres subalbatus* larvae was reported⁽¹²⁵⁾.

Melaleuca alternifolia; Tea Tree:

Oil of tea tree containing terpenoids such as terpinen-4ol and cineole has the ability to kill resistant fungus and bacteria⁽¹²⁶⁾. The reported side effects included burning and allergic reactions(127). Its oil was reported to have similar repellent effects, as clove oil used against scrub typhus⁽⁹⁴⁾. Tea tree oil (5%) showed similar effects as those of benzyl benzoate, lindane and ivermectin against all stages of S. scabiei(128). Acaricidal and pediculocidal activity against house dust mites and lice was evaluated using three essential oils: Tea tree, lavender and lemon and the results revealed that tea tree oil was the most effective(129). Moreover, improvement of symptoms and dramatic resolution of ocular irritation and conjunctival inflammation were reported in patients with *Demodex* blepharitis who were treated with 50% tea tree oil shampoo for a minimum of 6 weeks⁽¹³⁰⁾.

Cucurbita pepo, Cucurbita maxima; Pumpkin (Karh El-Asal, or Al-Yakten):

Pumpkin seeds contain several active constituents: Essential fatty acids, amino acids, phytosterols (e.g. β-sitosterol), minerals, vitamins and curcurbitin which shows anti-parasitic activity⁽¹³¹⁾. Oral administration of crude ethanolic extracts from *C. maxima* reduced by 50% the levels of parasitaemia in *Plasmodium berghei*-infected mice⁽¹³²⁾. In heterophyiasis, a combined extract of areca nut and pumpkin seeds gave an excellent result than when either extract was given alone⁽¹³³⁾. In addition, it gave effective results against canine tapeworms⁽¹³⁴⁾. Its leaf extract with different solvents showed larvicidal, ovicidal and repellent activities against *Culex quinquefasciatus*⁽¹³⁵⁾.

Less Commonly Used Herbal Medicine with Antiparasitic Effects

Cuminum cyminum; Cumin (Al-Kammoun): Cuminaldehyde, cymene and terpenoids are the major constituents of volatile oils of cumin⁽¹³⁶⁾. It is usually used in traditional medicine as a stimulant, a carminative and an astringent⁽¹³⁷⁾. Significant larvicidal activity (100% mortality) of essential oils derived from cumin against early 4th stage larvae of *Aedes aegypti* and *Culex pipiens* was reported⁽¹³⁸⁾.

Eugenia caryophyllata; Syzygium aromaticum Clove (Al-Kornfel): The main constituents of the clove essential oil are phenylpropanoids such as carvacrol, thymol, eugenol and cinnamaldehyde⁽¹³⁹⁾. Under field conditions, Gel B (20% clove oil) provided significant repellent effects against Aedes aegypti, Culex quinquefasciatus and Mansonia uniformis. These promising results raised the possibility for its use by low-income rural communities against various mosquito species⁽¹⁴⁰⁾. Moreover, essential oils of clove showed repellent effects against host-seeking chiggers (scrub typhus) of Leptotrombidium imphalum⁽⁹⁴⁾.

Pimpinella anisum; Anise (Al-Yanson): The active constituents in anise, particularly the terpenoid anethole, are in its volatile oil⁽¹⁴¹⁾. Its uses for medication in parasitic diseases are due to its insecticidal and repellent effects. The acaricidal activity of anise seed oil against house dust mites Dermatophagoides farinae and D. pteronyssinus was about 8.4 and 6.7 times more toxic than benzyl benzoate, respectively⁽¹⁴²⁾. Besides its acaricidal activity, it was highly effective as both larvicidal and ovicidal against different mosquito spp. As Anopheles, Aedes and Culex⁽⁹⁵⁾. Moreover, its spray preparation had significant cure rates against head louse infestations⁽¹⁴³⁾.

Punica granatum; Pomegranate (Al-Roman): Ellagic acid is likely to be the main constituent responsible for the antiparasitic effect of pomegranate⁽¹⁴⁴⁾. It acts through stimulation of the cell-mediated and humoral components of the immune system⁽¹⁴⁵⁾. The *in vitro* studies conducted in 2009 proved that ellagic acid may contribute as a promising anti-malarial drug⁽¹⁴⁶⁾, it had the ability to inhibit the growth of *P. falciparum* asexual blood stages *in vitro*⁽¹⁴⁷⁾. In addition, a natural plant extract purified from pomegranate was effective against *T. vaginalis in vitro*⁽⁶⁷⁾.

Ricinus communis; Castor Oil Plant (El-Khirwa): Ricinoleic acid is the main component of castor oil, and it constitutes approximately 90% of its fatty acid content (148). Leaves of castor oil plant are toxic to sand flies and offer protection against their bites and decrease the risk of leishmaniasis (149). The methanol extract of castor oil exhibited acaricidal and insecticidal activities against Haemaphysalis bispinosa and Hippobosca maculata (158).

Rosmarinus officinalis; Rosemary (Ekleel Al-Gabal): The active constituents include volatile oil (eucalyptol or cineole) as a potent antibacterial agent, rosmarinic acid with antioxidant activity and carnosol as anti-cancer formation in animal studies. Internal intake of its oil should be avoided during pregnancy because it may cause abortion⁽¹⁵¹⁾. Rosemary oil produced decreased oviposition, had high larvicidal activity against all larval stages of A. aegypti and proved to be a promising agent in mosquito control programs⁽¹⁵²⁾

Salix alba; Willow (Al-Sefsaf): The main active constituent is glycoside salicin which has anti-inflammatory and pain-relieving actions. As with aspirin, some people may experience stomach upset from taking willow⁽¹⁵³⁾. During the American Civil War, the unreliable supply and high cost of quinine forced the Confederate Army to use alternative treatments for malaria which were made from indigenous plants such as dogwood, willow and tulip tree. The quinine substitutes were generally considered useful but not as effective as quinine⁽¹⁵⁴⁾.

Thymus vulgaris; Thyme (Al-Zaatar): The primary constituents are the volatile oils, including phenols, thymoland carvacol. Thyme oil should be reserved for topical

use, as internally it may lead to dizziness, vomiting and breathing difficulties⁽¹⁵¹⁾. So, its hydroalcoholic extract was significantly effective in reduction of ulcer size in cutaneous leishmaniasis⁽¹⁵⁵⁾. Its volatile oil was effective against *Lucilia sericata* 3rd stage larvae⁽¹⁵⁶⁾ and *Pediculus humanus*⁽¹⁵⁷⁾.

Herbal Medicine with Limited Molluscicidal Effects *Ambrosia maritima* (Damsissa): In a field trial in Egypt, the possible use of single annual application of damsissa plant, in snail control program for schistosomiasis transmission season in Lower Egypt, was suggested⁽¹⁵⁸⁾. Longer prepatent period and remarkable decrease in cercarial production was also recorded in snails treated with the sublethal concentrations of this molluscicide⁽¹⁵⁹⁾. In addition, it has lethal effects on *Lymnaea* snails⁽¹⁶⁰⁾.

Azolla pinnata: Molluscicidal activity of A. pinnata was reported against B. alexandrina snails. Its ethanol extract, at 6600 ppm for three hours exposure, killed 100% and 19.4% of S. mansoni miracidia and cercariae, respectively⁽¹⁶¹⁾.

Euphorbia spp. (Sabbaar): The physiological and lethal effects of the latex components of *E. splendens*⁽¹⁶²⁾ and *E. milii*⁽¹⁶³⁾, on *Biomphalaria spp.* were reported. It was found that unfiltered crude latex extract is a more potent molluscicide than filtered latex⁽¹⁶²⁾ and milin (the active component) is likely to be responsible for alteration of normal physiological functions and lethality of snails⁽¹⁶³⁾.

Acknowledgment: The authors would like to thank Prof. Dr. Raefa Darwish and Prof. Dr. Soad Nada, Parasitology Department, Faculty of Medicine, Zagazig University, for revising the review article and Eman Magdy for references collection.

REFERNCES

- WHO. http://www.who.int/mediacentre/factsheets/fs134/en/; December, 2008.
- Aggarwal BB, Sundaram C, Malani N, Ichikawa H. Curcumin: The Indian solid gold. Adv Exp Med Biol; 2007, 595: 1-75.
- Abdulelah HAA, Zainal-Abidin BAH. In vivo anti-malarial tests of Nigella sativa (Black seed) different extracts. Am J Pharmacol Toxicol; 2007, 2: 46-50.
- Acharya D, Shrivastava A. Indigenous Herbal Medicines: Tribal formulations and traditional herbal practices. Aavishkar Publishers Distributor, Jaipur, India; 2008, 440: 252-7.
- WHO. Prevention and control of schistosomiasis and soil-transmitted helminthiasis. Report of a WHO Expert Committee; 2002, WHO Tech Rep Ser No. 912.
- 6. Greenwood BMK, Bojang CJ, Whitty M, Targett GA. Malaria. Lancet; 2005, 365: 1487-98.

- 7. Croft SL, Sundar S, Fairlamb AH. Drug resistance in leishmaniasis. Clin Microbiol Rev; 2006, 19: 111–26.
- Anthony JP, Fyfe L, Smith H. Plant active components: A resource for antiparasitic agents? Trends Parasitol; 2005, 21 (10): 462-8.
- 9. Elvin LM. Should we be concerned about herbal remedies? J Ethnopharmacol; 2001, 75: 141-164.
- Feldberg RS, Chang SC, Kottik AN. *In vitro* mechanism of inhibition of bacterial growth by Allicin. Antimicrob Agents Chemother; 1988, 32: 1763-1768.
- 11. Saleheen D, Ali SA, Yasinzai MM. Antileishmanial activity of aqueous onion extract *in vitro*. Fitoterapia; 2004, 75(1):9-13
- 12. Ahmed SA. *In vitro* effects of aqueous garlic (*Allium sativum*) and onion (*Allium cepa*) extracts on trichomonas vaginalis. PUJ; 2010, 3 (1&2): 45-54.
- 13. Abu El-Ezz NM. Effect of Nigella sativa and Allium cepa oils on Trichinella spiralis in experimentally infected rats. J Egypt Soc Parasitol; 2005, 35 (2): 511-23.
- Stozkowska W, Raczyńska K. Efficiency and application safety of Cepan cream: Observation of a new indication. Przegl Lek; 2008, 65 (5): 241-3.
- 15. Mantawy MM, Mahmoud AH. Effect of *Allium cepa* and *Allium sativum* feeding on glucose, glycogen, protein bands profile and phenol oxidase activity in *Biomphalaria alexandrina*. J Egypt Soc Parasitol; 2002, 32 (1): 271-83.
- Valdivieso R, Subiza J, Varela-Losada S. Bronchial asthma. rhinoconjunctivitis and contact dermatitis caused by onion. J Allergy Clin Immunol; 1994, 94: 928-30.
- 17. Ayaz E, Alpsoy HC. Garlic (*Allium sativum*) and traditional medicine. Turkiye Parazitol Derg; 2007, 31 (2): 145-9.
- Ankri S, Mirelman D. An antimalarial extract from neem leaves is antiretroviral: Antimicrobial properties of allicin from garlic. Microbe Infect; 1999, 1: 125-9.
- 19. Davis SR. An overview of the antifungal properties of allicin and its breakdown products: The possibility of a safe and effective antifungal prophylactic. Mycoses; 2005, 48 (2): 95-100.
- 20. Soffar SA, Mokhtar GM. Evaluation of the antiparasitic effect of aqueous garlic (*Allium sativum*) extract in *Hymenolepiasis nana* and giardiasis. J Egypt Soc Parasitol; 1991, 21: 497-502.
- 21. Harris JC, Plummer S, Turner MP, Lloyd D. The microaerophilic flagellate *Giardia intestinalis:* Allium sativum (garlic) is an effective antigiardial. Microbiology; 2000, 146 (12): 3119-27.
- Jimenez JC, Uzcanga G, Zambrano A. Identification and partial characterization of excretory/secretory products with proteolytic activity in *Giardia intestinalis*. J Parasitol; 2000, 86: 859-62.
- 23. El-Nahas N, El-Melegy M, Abdel-Wahed M. Effect of garlic on experimental cyclosporiasis. Egypt J Med Sci; 2003, 24 (1): 53-64.

- Coppi A, Cabinian M, Mirelman D, Sinnis P. Antimalarial activity of allicin, a biologically active compound from garlic cloves. Antimicrob Agents Chemother; 2006, 50 (5): 1731-7.
- Ghazanfari T, Hassan ZM, Khamesipour A. Enhancement of peritoneal macrophage phagocytic activity against Leishmania major by garlic (Allium sativum) treatment. J Ethnopharmacol; 2006, 103 (3): 333-7.
- Gamboa LMR, Aranda GI, Mut MM, García MMR, Dumonteil E. *In vivo* and *in vitro* control of *Leishmania mexicana* due to garlic-induced NO production. Scand J Immunol; 2007, 66 (5): 508-14.
- Toulah FH, Al-Rawi MM. Efficacy of garlic extract on hepatic coccidiosis in infected rabbits (*Oryctolagus* cuniculus): Histological and biochemical studies. J Egypt Soc Parasitol; 2007, 37 (3): 957-68.
- Polat ZA, Vural A, Ozan F, Tepe B, Ozcelik S, Cetin A. In vitro evaluation of the amoebicidal activity of garlic (Allium sativum) extract on Acanthamoeba castellanii and its cytotoxic potential on corneal cells. J Ocul Pharmacol Ther; 2008, 24 (1): 8-14.
- 29. El-Shenawy NS, Soliman MF, Reyad SI. The effect of antioxidant properties of aqueous garlic extract and *Nigella sativa* as anti-schistosomiasis agents in mice. Rev Inst Med Trop Sao Paulo; 2008, 50 (1): 29-36.
- Riad NHA, Taha HAT, Mahmoud YI. Effects of garlic on albino mice experimentally infected with *Schistosoma mansoni*: A parasitological and ultrastructural study. Trop Biomed; 2009, 26 (1): 40-50.
- 31. Sadjjadi SM, Zoharizadeh MR, Panjeshahin MR. *In vitro* screening of different *Allium sativum* extracts on hydatid cysts protoscoleces. J Invest Surg; 2008, 21 (6): 318-22.
- 32. Valerio L, Maroli M. Evaluation of repellent and antifeeding effect of garlic oil (*Allium sativum*) against the bite of *Phlebotomine* sandflies (*Diptera: Psychodidae*). Ann Ist Super Sanita; 2005, 41 (2): 253-6.
- 33. Birrenkott GP, Brockenfelt GE, Greer JA, Owens MD. Topical application of garlic reduces northern fowl mite infestation in laying hens. Poult Sci; 2000, 79(11): 1575-7.
- 34. Singh A, Singh DK. Effect of herbal molluscicides and their combinations on the reproduction of the snail *Lymnaea acuminata*. Arch Environ Contam Toxicol; 2004, 46 (4): 470-7.
- 35. AlKadi HO. Antimalarial drug toxicity: A review. Chemotherapy; 2007, 53:385-91.
- Leung AY, Foster S. Encyclopedia of Common Natural Ingredients Used in Food, Drugs, and Cosmetics. 2nd Edition, John Wiley and Sons, New York; 1996, 3: 382.
- 37. McGuffun M, Hobbs C, Upton R, Goldberg A. American Herbal Products Association's Botanical Safety Handbook. CRC Press, Boca Raton, FL; 1997, 15.
- 38. Song J, Socheat D, Tan B *et al.* Rapid and effective malaria control in Cambodia through mass administration of artemisinin-piperaquine. Malar J; 2010, 23: 9-57.

- 39. Dondorp AM, Yeung S, White L *et al.* Artemisinin resistance: Current status and scenarios for containment. Nat Rev Microbiol; 2010, 8 (4): 272-80.
- 40. Posner GH, O'Neill PM. Knowledge of the proposed chemical mechanism of action and cytochrome p450 metabolism of antimalarial trioxanes like artemisinin allows rational design of new antimalarial peroxides. Acc Chem Res; 2004, 37 (6): 397-404.
- 41. Woodrow CJ, Planche T, Krishna S. Artesunate versus quinine for severe *falciparum* malaria. Lancet; 2006, 14 (9505): 110-1.
- 42. Golenser J, Waknine JH, Krugliak M, Hunt NH, Grau GE. Current perspectives on the mechanism of action of artemisinins. Int J Parasitol; 2006, 36: 1427–41.
- 43. Sen R, Bandyopadhyay S, Dutta A *et al.* Artemisinin triggers induction of cell-cycle arrest and apoptosis in *Leishmania donovani* promastigotes. J Med Microbiol; 2007, 56 (9): 1213-8.
- 44. Nagamune K, Moreno SN, Sibley LD. Artemisininresistant mutants of *Toxoplasma gondii* have altered calcium homeostasis. Antimicrob Agents Chemother; 2007, 51 (11): 3816-23.
- 45. Dunay IR, Chan WC, Haynes RK, Sibley LD. Artemisone and artemiside control acute and reactivated toxoplasmosis in a murine model. Antimicrob Agents Chemother; 2009, 53 (10): 4450-6.
- 46. D'Angelo JG, Bordón C, Posner GH, Yolken R, Jones-Brando L. Artemisinin derivatives inhibit *Toxoplasma gondii in vitro* at multiple steps in the lytic cycle. J Antimicrob Chemother; 2009, 63 (1): 146-50.
- 47. Yang YQ, Xiao SH, Tanner M *et al.* Histopathological changes in juvenile *Schistosoma haematobium* harboured in hamsters treated with artemether. Acta Trop; 2001, 79: 135-41.
- Xiao SH, Shen BG, Tzinger JU, Chollet J, Tanner M. Ultrastructural alterations in adult *Schistosoma* mansoni caused by artemether. Mem Inst Oswaldo Cruz; 2002, 97: 717–24.
- 49. El-Bassiouni EA, Helmy MH, Saad EI, Abdel-El-Nabi KM, Abdel-Meguid E, Hussein HS. Modulation of the antioxidant defence in different developmental stages of Schistosoma mansoni by praziquantel and artemether. Br J Biomed Sci; 2007, 64 (4): 168-74.
- 50. Keiser J, Rinaldi L, Veneziano V *et al.* Efficacy and safety of artemether against a natural *Fasciola hepatica* infection in sheep. Parasitol Res; 2008, 103 (3): 517-22.
- O'Neill JF, Johnston RC. Halferty L, Brennan GP, Keiser J, Fairweather I. Adult triclabendazole-resistant *Fasciola hepatica*: Morphological changes in the tegument and gut following *in vivo* treatment with artemether in the rat model. J Helminthol: 2009, 83 (2): 151-63.

- 52. Caner A, Doskaya M, Degirmenci A, Can H, Baykan S. Comparison of the effects of *Artemisia vulgaris* and *Artemisia absinthium* growing in western Anatolia against trichinellosis (*Trichinella spiralis*) in rats. Exp Parasitol; 2008, 119 (1): 173-9.
- 53. Jaenson TG, Palsson K, Borg-Karlson AK. Evaluation of extracts and oils of tick-repellent plants from Sweden. MedVet Entomol; 2005, 9 (4): 345-52.
- 54. Mills SY. Out of the Earth: The Essential Book of Herbal Medicine. Middlesex, UK: Viking Arkana, 1991: 500-2.
- Badria F, Abou-Mohamad G, El-Mowafy A, Masoud A, Salama T. Mirazid: A new schistosomicidal drug. Pharmaceut Biol; 2001, 39: 127-131.
- Massoud AM, El-Ebiary FH, Abou-Gamra MM, Mohamed GF, Shaker SM. Evaluation of schistosomicidal activity of myrrh extract: Parasitological and histological study. J Egypt Soc Parasitol; 2004, 34 (3): 1051-76.
- 57. Hamed MA, Hetta MH. Efficacy of *Citrus reticulata* and Mirazid in treatment of *Schistosoma mansoni*. Mem Inst Oswaldo Cruz; 2005, 100: 771–8.
- 58. Abo-Madyan AA, Morsy TA, Motawea SM Efficacy of Myrrh in the treatment of schistosomiasis (haematobium and mansoni) in Ezbet El-Bakly, Tamyia Center, El-Fayoum Governorate. Egypt. J Egypt Soc Parasitol, 34: 423-46.
- Botros S, William S, Ebeid FD, Cioli N, Day TA. Lack of evidence for an antischistosomal activity of myrrh in experimental animals, Am J Trop Med Hyg: 2004, 71: 206–10.
- 60. Barakat R, El-Morshedy H, Fenwick A. Efficacy of myrrh in the treatment of human schistosomiasis *mansoni*. Am J Trop Med Hyg; 2005, 73: 365-7.
- 61. Mahmoud MS, Dobal SA, Soliman K. Immune response in *Fasciola gigantica* experimentally infected rabbits treated with either carnosine or Mirazid®. Res J Parasitol; 2008, 3: 40-9.
- 62. Botros SS, El-Lakkany NM, Badawy AA, Mahmoud SS, Ebeid FA, Fenwick A. Mirazid shows insignificant activity against ovine fascioliasis. Ann Trop Med Parasitol; 2009, 103 (7): 605-16.
- 63. Al-Mathal EM, Fouad MA. Myrrh (Commiphora molmol) in treatment of human and sheep dicrocoeliasis dendriticum in Saudi Arabia. J Egypt Soc Parasitol; 2004, 34 (2): 713-20.
- 64. Massoud AM, El-Shazly AM, Morsy TA. Mirazid (*Commiphora molmol*) in treatment of human heterophyiasis. J Egypt Soc Parasitol, 37 (2): 395-410.
- Massoud, AM, Shazly AM, Shahat SA, Morsy TA. Mirazid in treatment of human hymenolepiasis. J Egypt Soc Parasitol; 2007, 37 (3): 863-76.
- 66. Massoud AM, Hafez AO, Abdel-Gawad AG, El-Shazly AM, Morsy TA. Mirazid alone or combined with Paromomycin in treating cryptosporidiosis parvum in immunocom-petent hospitalized patients. J Egypt Soc Parasitol; 38 (2): 399-418.

- 67. El-Sherbini GT, El-Gozamy BR, Abdel-Hady NM. Morsy TA. Efficacy of two plant extracts against vaginal trichomoniasis. J Egypt Soc Parasitol; 2009. 39(1): 47-58.
- 68. Massoud AM, Labib IM. Larvicidal activity of Commiphora molmol against Culex pipiens and Aedes caspius larvae. J Egypt Soc Parasitol; 2000, 30(1): 101-15.
- 69. Allam AF, El-Sayad MH, Khalil SS. Laboratory assessment of the molluscicidal activity of *Commiphora molmol* (Myrrh) on *Biomphalaria alexandrina*. *Bulinus truncatus* and *Lymnaea cailliaudi*. J Egypt Soc Parasitol; 31: 683-90.
- Massoud AM, Metwally DM, Khalifa KE, Habib FS. Compatibility of *Biomphalaria alexandrina* snails to infection with *Schistosoma mansoni* after exposure to sublethal concentrations of Myrrh. J Egypt Soc Parasitol; 34: 995–1008.
- 71. Blumenthal M, Busse WR, Goldberg A. The Complete Commission E Monographs: Therapeutic guide to herbal medicines. Boston, MA, Integrative Medicine Communications; 1998; 309.
- Charles V, Charles SX. The use and efficacy of Azadirachta indica ADR ('Neem') and Curcuma longa ('Turmeric') in scabies: A pilot study. Trop Geogr Med; 1992, 44(1-2):178-81
- Waknine-Grinberg JH, McQuillan JA, Hunt N, Ginsburg H, Golenser J. Modulation of cerebral malaria by fasudil and other immune-modifying compounds. Exp Parasitol; 2010, 125 (2): 141-6
- 74. Nose M, Koide T, Ogihara Y, Yabu Y, Ohta N. Trypanocidal effects of curcumin *in vitro*. Biol Pharm Bull; 1998, 21 (6): 643-5.
- 75. Koide T, Nose M, Ogihara Y. Leishmanicidal Effect of Curcumin *in Vitro*. Biol Pharm Bull; 2002, 25 (1): 131-3.
- Pérez-Arriaga L, Mendoza-Magaña ML, Cortés-Zárate R et al. Cytotoxic effect of curcumin on Giardia lamblia trophozoites. Acta Trop; 2006, 98 (2): 152-161.
- Al-Zanbagi NA. Zelai NT. Two methods for attenuating Toxoplasma gondii tachyzoites RH strain by using ethanol extract of Curcuma longa. J Egypt Soc Parasitol; 2008, 38 (3): 965-76.
- El-Ansary AK, Ahmed SA, Aly SA. Antischistosomal and liver protective effects of *Curcuma longa* extract in *Schistosoma mansoni* infected mice. Ind J Exp Biol; 2007, 45 (9): 791-801.
- 79. Allam G. Immunomodulatory effects of curcumin treatment on murine schistosomiasis *mansoni*. Immunobiology; 2009, 214 (8): 712-27.
- 80. Boonjaraspinyo S, Boonmars T, Aromdee C *et al.* Turmeric reduces inflammatory cells in hamster opisthorchiasis. Parasitol Res; 2009, 105 (5): 1459-63.
- 81. Omran NE, El-Nouby KA, Shoheib ZS, Kabbash AM. Molluscicidal effects of some plant extracts on *Biomophlaria alexandrina*, the intermediate host snail of *Schistosoma mansoni* in Egypt. J Egypt Ger Zooi; 2007, 53: 1-29.

- 82. Boskabady MH. Effect of *Nigella Sativa* on isolated guinea pig trachea. Arch Iran Med; 2002, 5 (2): 103-7.
- 83. EL-Dakhakhny M, Mady NI. Halim MA. *Nigella* sativa oil protects against induced hepatotoxicity and improves serum lipid profile in rats. Arzneimit Forsch; 2000, 50: 832-6.
- 84. Mady NI, Abdel-Aziem T, Matta M. Effect of combination of *Nigella sativa* oil and glibenclamide on some metabolic parameters and oxidative changes in streptozocin-induced diabetic rats (*in vivo* and *in vitro* study). J Egypt Pharmacol Exp Ther; 2001, 20: 359-83.
- 85. Ayaz E, Yilmaz H, Ozbek H, Tas Z, Orunc O. The effect of *Nigella sativa* oil against *Aspiculuris tetraptera* and *Hymenolepis nana* in naturally infected mice. Saudi Med J; 2007, 28 (11): 1654-7.
- 86. El-Wakil HS. Evaluation of the *in vitro* effect of *Nigella sativa* aqueous extract on *Blastocystis hominis* isolates. J Egypt Soc Parasitol: 2007, 37 (3): 801-13.
- 87. Ahmed MA. *In vitro* inhibition of *Trichomonas vaginalis* growth by WGA and *Nigella sativa*. New Egypt J Med; 2008, 39 (1): 156-68.
- 88. Ghayur MN, Gilani AH. Pharmacological basis for the medicinal use of ginger in gastrointestinal disorders. Dig Dis Sci; 2005, 50: 1889-97.
- 89. Sohni YR, Kaimal P, Bhatt RM. The antiamoebic effect of a crude drug formulation of herbal extracts against *Entamoeba histolytica in vitro* and *in vivo*. J. Ethnopharmacol; 1995, 45 (1): 43-52.
- 90. Sanderson L, Bartlett A, Whitfield PJ. *In vitro* and *in vivo* studies on the bioactivity of a ginger (Zingiber officinale) extract towards adult schistosomes and their egg production. J Helminthol; 2002, 76(3):241-7.
- 91. Iqbal Z, Lateef M, Akhtar MS, Ghayur MN, Gilani AH. *In vivo* anthelmintic activity of ginger against gastrointestinal nematodes of sheep. J Ethnopharmacol; 2006, 106 (2): 285-7.
- 92. El-Melegy MA, El-Saify GH, Hassab-El-Nabi SE. Evaluation of the therapeutic effect of ginger compared to flubendazole on experimental trichinellosis in mice. Egypt J Med Sci; 2006, 27 (2): 25-48.
- 93. Merawin LT, Arifah AK, Sani RA *et al.* Screening of microfilaricidal effects of plant extracts against *Dirofilaria immitis*. Res Vet Sci.; 2010, 88 (1): 142-7.
- 94. Eamsobhana P, Yoolek A, Kongkaew W et al. Laboratory evaluation of aromatic essential oils from thirteen plant species as candidate repellents against *Leptotrombidium* chiggers (*Acari: Trombiculidae*), the vector of scrub typhus. Exp Appl Acarol; 2009, 47 (3): 257-62.
- 95. Prajapati V, Tripathi AK, Aggarwal KK, Khanuja SP. Insecticidal, repellent and oviposition-deterrent activity of selected essential oils against *Anopheles stephensi*, *Aedes aegypti* and *Culex quinquefasciatus*. Bioresour Technol; 2005, 96 (16): 1749-57.

- 96. Rahuman AA, Gopalakrishnan G, Venkatesan P, Geetha K, Bagavan A. Mosquito larvicidal activity of isolated compounds from the rhizome of *Zingiber officinale*. Phytother Res; 22 (8): 1035-9.
- 97. Adewunmi CO, Oguntimein BO, Furu P. Molluscicidal and antischistosomal activities of *Zingiber officinale*. Planta Med; 1990, 56 (4): 374-6.
- 98. Singh VK, Singh S, Singh DK. Effect of active molluscicidal component of spices on different enzyme activities and biogenic amine levels in the nervous tissue of *Lymnaea acuminata*. Phytother Res; 1999, 13 (8): 649-54.
- 99. Ghosh AK, Bhattacharyya FK, Ghosh DK. *Leishmania donovani*: Amastigote inhibition and mode of action of berberine. Exp Parasitol; 1985, 60 (3): 404-13.
- 100. Ahuja A, Purohit SK, Yadav JS, Netra PR. Cutaneous leishmaniasis in domestic dogs. Ind J Public Health; 1993, 37 (1): 29-31.
- 101. Sohni YR, Bhatt RM. Activity of a crude extract formulation in experimental hepatic amoebiasis and in immunomodulation studies. J Ethnopharmacol; 1996, 54 (2-3): 119-24.
- 102. Sheng WD, Jiddawi MS, Hong XQ, Abdulla SM. Treatment of chloroquine-resistant malaria using pyrimethamine in combination with berberine, tetracycline or cotrimoxazole. East Afr Med J; 1997, 74 (5): 283-4.
- 103. Soffar SA, Metwali DM, Abdel-Aziz SS, El-Wakil HS, Saad GA. Evaluation of the effect of a plant alkaloid (berberine derived from *Berberis aristata*) on *Trichomonas vaginalis in vitro*. J Egypt Soc Parasitol; 2001, 31 (3): 893-904.
- 104. Rembold H. The azadirachtins-their potential for insect control. Econ Med Plant Res; 1989, 3:57-72.
- 105. Sinniah D, Baskara G, Looi LM, Leong KL. Reyelike syndrome due to margosa oil poisoning: Report of a case with postmortem findings. Am J Gastroenterol; 1982, 77: 158-161.
- 106. Gunasekaran K, Vijayakumar T, Kalyanasundaram M. Larvicidal and emergence inhibitory activities of Neem Azal T/S 1.2% EC against vectors of malaria, filariasis and dengue. Indian J Med Res; 2009, 130 (2): 138-145.
- 107. Abdel-Ghaffar F, Al-Quraishy S, Sobhy H, Semmler M. Neem seed extract shampoo (Wash Away Louse): An effective plant agent against Sarcoptes scabiei mites infesting dogs in Egypt. Parasitol Res; 2008, 104 (1): 145-8.
- 108. Srivastava R, Ghosh S, Mandal DB *et al*. Efficacy of *Azadirachta indica* extracts against *Boophilus microplus*. Parasitol Res; 2008, 104 (1): 149-153.
- 109. Abdel-Ghaffar F, Semmler M. Efficacy of neem seed extract shampoo on head lice of naturally infected humans in Egypt. Parasitol Res; 2007, 100 (2): 329-32.
- 110. Srinivasan R, Kalyanasundaram M. Relative efficacy of DEPA and neem oil for repellent activity against

- *Phlebotomus papatasi*, the vector of leishmaniasis. J Commun Dis; 2001, 33 (3): 180-4.
- 111. Garboui SS, Jaenson TG, Pålsson K. Repellency of MyggA Natural spray (para-menthane-3, 8-diol) and RB86 (neem oil) against the tick Ixodes *ricinus* (*Acari: Ixodidae*) in the field in east-central Sweden. Exp Appl Acarol; 2006, 40 (3-4): 271-7.
- 112. Taylor WR, White NJ. Antimalarial drug toxicity: A review. Drug Saf; 2004, 27: 25–61.
- 113. Kayitare E, Vervaet C, Mehuys E *et al.* Taste-masked quinine pamoate tablets for treatment of children with uncomplicated *Plasmodium falciparum* malaria. Int J Pharmacol; 2010, 392 (1-2): 29-34.
- 114. Kremsner PG, Winkler S, Brandts M, Neifer CS. Clindamycin in combination with chloroquine or quinine is an effective therapy for uncomplicated *Plasmodium falciparum* malaria in children from Gabon. J Infect Dis; 1994, 169:467-70.
- 115. Achan J, Tibenderana JK, Kyabayinze D. Effectiveness of quinine versus artemether-lumefantrine for treating uncomplicated *falciparum* malaria in Ugandan children. Brit Med J; 2009, 338: 2763.
- 116. Corrêa SJB, Menezes D, Vannier-Santos MA, Ferreira-Pereira A, Almeida GT. Interference with hemozoin formation represents an important mechanism of schistosomicidal action of antimalarial quinoline methanols. PLos Negl Trop Dis; 2009, 3 (7): 477.
- 117. Kamgno J, Djomo PN, Pion SD, Thylefors B, Boussinesq M. A controlled trial to assess the effect of quinine, chloroquine, amodiaquine and artesunate on *Loa loa* microfilaremia. Am J Trop Med Hyg; 2010, 82(3): 379-85.
- 118. Grant J, Mahanty S, Khadir A. Wheat germ supplement reduces cyst and trophozoite passage in people with giardiasis. Am J Trop Med Hyg; 2001, 65: 705-10.
- 119. Eassa AHA, Ismail MAM, Younis AIH, El-Wakil SS. Effect of wheat germ supplement in patients with intestinal amoebiasis. Presented in the 22nd. Conference of the Egyptian, Society Of Basic Medical Sciences, 17th, April 2003, Cairo University, Cairo, Egypt.
- 120. Moustafa MA. Role of wheat germ agglutinin (WGA) in treatment of experimental cryptosporidiosis. J Egypt Soc Parasitol; 2003, 33 (2): 443-56.
- 121. Singh HB, Srivastava M, Singh AB, Srivastava AK. Cinnamon bark oil, a potent fungitoxicant against fungi causing respiratory tract mycoses. Allergy, 1995, 50: 995-9.
- 122. Yang YC, Lee HS, Lee SH, Clark JM, Ahn YJ. Ovicidal and adulticidal activities of *Cinnamomum zeylanicum* bark essential oil compounds and related compounds against *Pediculus humanus capitis* (*Anoplura: Pediculicidae*). Int J Parasitol; 2005, 35 (14): 1595-600.
- 123. Kim SI, Yi JH, Tak JH, Ahn YJ. Acaricidal activity of plant essential oils against *Dermanyssus gallinae* (*Acari: Dermanyssidae*). Vet Parasitol; 2004, 120 (4): 297-304.

- 124. Cheng SS, Liu JY, Tsai KH, Chen WJ, Chang ST. Chemical composition and mosquito larvicidal activity of essential oils from leaves of different *Cinnamomum osmophloeum* provenances. J Agricul Food Chem; 2004, 52: 4395-400.
- 125. Cheng SS, Liu JY, Huang CG, Hsui YR, Chen WJ, Chang ST. Insecticidal activities of leaf essential oils from *Cinnamomum osmophloeum* against three mosquito species. Bioresour Technol; 2009, 100 (1): 457-64.
- 126. Carson CF, Cookson BD, Farrelly HD, Riley T. Susceptibility of methicillin-resistant Staphylococcus aureus to the essential oil of Melaleuca alternifolia. J Antimicrob Chemother; 1995, 35:421-4.
- 127. Knight TE, Hansen BM. Melaleuca oil (tea tree oil) dermatitis. Med J Australia; 1994, 30: 423-27.
- 128. Walton SF, Myerscough MR, Currie BJ. Studies in vitro on the relative efficacy of current acaricides for Sarcoptes scabiei var. hominis. Trans R Soc Trop Med Hyg; 2000, 94 (1): 92-6.
- 129. Canyon DV, Speare R. A comparison of botanical and synthetic substances commonly used to prevent head lice (*Pediculus humanus var. capitis*) infestation. Int J Dermatol; 2007, 46 (4): 422-6.
- 130. Kheirkhah A, Casas V, Li W, Raju VK, Tseng SC. Corneal manifestations of ocular *Demodex* infestation. Am J Ophthalmol; 2007, 143 (5): 743-9.
- 131. Rybaltovskii OV. On the discovery of cucurbitin-a component of pumpkin seed with anthelmintic action. Med. Parazitol; 1966, 35: 487-8.
- 132. Amorim CZ, Marques AD, Cordeiro RS. Screening of the antimalarial activity of plants of the *Cucurbitaceae* family. Mem Inst Oswaldo Cruz; 1991, 86 (Suppl 2): 177-80.
- 133. Mahmoud LH, Basiouny SO, Dawoud HA. Treatment of experimental heterophyiasis with two plant extracts, areca nut and pumpkin seed. J Egypt Soc Parasitol; 2002, 32 (2): 501-6.
- 134. Díaz OD, Lloja LL, Carbajal ZV. Preclinical studies of *Cucurbita maxima* (pumpkin seeds): A traditional intestinal antiparasitic in rural urban areas. Rev Gastroenterol Peru; 2004, 24 (4): 323-327.
- 135. Mullai K, Jebanesan A. Larvicidal, ovicidal and repellent activities of the leaf extract of two cucurbitacious plants against filarial vector *Culex quinquefasciatus* (Say) (*Diptera: Culicidae*). Trop Biomed; 2007, 24 (1): 1-6.
- 136. El-Hamidi A, Ahmed SS. The content and composition of some umbelliferous essential oils. Die.Pharmazie; 1966, 7: 438-9.
- 137. Allahghadri T, Rasooli I, Owlia P *et al.* Antimicrobial property, antioxidant capacity and cytotoxicity of essential oil from cumin produced in Iran. J Food Sci; 2010, 75 (2): 54-61.
- 138. Lee HS. Mosquito larvicidal activity of aromatic medicinal plant oils against *Aedes aegypti* and *Culex pipiens pallens*. J Am Mosq Control Assoc; 2006, 22 (2): 292-5.

- 139. Chaieb K, Hajlaoui H, Zmantar T et al. The chemical composition and biological activity of clove essential oil, Eugenia caryophyllata (Syzigium aromaticum L. Myrtaceae): A short review. Phytother Res; 2007, 21 (6): 501-6.
- 140. Trongtokit Y, Rongsriyam Y, Komalamisra N, Krisadaphong P, Apiwathnasorn C. Laboratory and field trial of developing medicinal local Thai plant products against four species of mosquito vectors. Southeast Asian J Trop Med Public Health; 2004, 35 (2): 325-33.
- 141. Weiss RF. Herbal Medicine. Beaconsfield Publishers Ltd; Gothenberg, Sweden, 1985, 203.
- 142. Lee HS. P-Anisaldehyde: Acaricidal component of *Pimpinella anisum* seed oil against the house dust mites *Dermatophagoides farinae* and *Dermatophagoides pteronyssinus*. Planta Med; 2004, 70 (3): 279-81.
- 143. Burgess IF, Brunton ER, Burgess NA. Clinical trial showing superiority of a coconut and anise spray over permethrin 0.43% lotion for head louse infestation. Eur J Pediatr; 2010, 169 (1): 55-62.
- 144. Soh PN, Witkowski B, Olagnier D, Nicolau MC. *In vitro* and *in vivo* properties of ellagic acid in malaria treatment. Antimicrob Agents Chemoth; 2009, 53: 1100–6.
- 145. Gracious R, Selvasubramanian RS, Jayasundar S. Immunomodulatory activity of *Punica granatum* in rabbits: A preliminary study. J Ethnopharmacol; 2001, 78:85-7.
- 146. Dell'Agli M, Galli GV, Corbett Y *et al.* Antiplasmodial activity of *Punica granatum* L fruit rind. J Ethnopharmacol; 2009, 125 (2): 279-85.
- 147. Sturm N, Zimmermann Y, Hu HK, Wolf S. Compounds structurally related to ellagic acid show improved antiplasmodial activity. Antimicrob Agents Chemoth; 2009, 53: 622-30.
- 148. Burdock GA, Carabin IG, Griffiths JC. Toxicology and pharmacology of sodium ricinoleate. Food Chem Toxicol; 2006, 44: 1689-98.
- 149. Schlein Y, Jacobson RL, Müller GC. Sand fly feeding on noxious plants: A potential method for the control of leishmaniasis. Am J Trop Med Hyg; 2001, 65 (4): 300-3.
- 150. Zahir AA, Rahuman AA, Bagavan A, Santhoshkumar Mohamed RR, Marimuthu S. Evaluation of botanical extracts against Haemaphysalis bispinosa Leach. and Hippobosca maculata Parasitol Res; 2010, 107 (3): 585-92.
- 151. Newall CA, Anderson LA, Phillipson JD. Herbal Medicines: A guide for health-care professionals. The Pharmaceutical Press, London; 1996, 256-7.
- 152. Waliwitiya R, Kennedy CJ, Lowenberger CA. Larvicidal and oviposition-altering activity of monoterpenoids, trans-anithole and rosemary oil to the yellow fever

- mosquito Aedes aegypti (Diptera: Culicidae). Pest Manag Sci; 2009, 65 (3): 241-8.
- 153. Bradley PR. British Herbal Compendium. Vol 1. British Herbal Medicine Association, Bournemouth, Dorset, UK; 1992, 224-6.
- 154. Hasegawa GR. Quinine substitutes in the confederate army. Mil Med; 2007, 172 (6): 650-5.
- 155. Nilforoushzadeh MA, Shirani BL, Zolfaghari-Baghbaderani A. Comparison of *Thymus vulgaris* (Thyme), *Achillea millefolium* (Yarrow) and propolis hydroalcoholic extracts versus systemic glucantime in the treatment of cutaneous leishmaniasis in balb/c mice. J Vector Borne Dis; 2008, 45 (4): 301-6.
- 156. Morsy TA, Shoukry A, Mazyad SA, Makled KM. The effect of the volatile oils of *Chenopodium ambrosioides* and *Thymus vulgaris* against the larvae of *Lucilia sericata* (Meigen). J Egypt Soc Parasitol; 1198, 28 (2): 503-10.
- 157. Veal L. The potential effectiveness of essential oils as a treatment for headlice, *Pediculus humanus capitis*. Complement Ther Nurs Midwifery; 1996, 2 (4): 97-101.
- 158. El-Sawy MF, Duncan J, Marshall TF, Shehata MA, Brown N. The molluscicidal properties of *Ambrosia maritima* L. (*Compositae*). 2. Results from a field trial using dry plant material. Trop Parasitol; 1984, 35 (2): 100-4.
- 159. El-Ansary A, El-Bardicy SS, Zayed N. Sublethal concentration of *Ambrosia maritima* (Damsissa) affecting compatibility of *Biomphalaria alexandrina* snails to infection with *Schistosoma mansoni th*rough disturbing the glycolytic pathway. J Egypt Soc Parasitol; 2000, 30 (3): 809-19.
- 160. Abou Basha LM, El-Sayad MH, Allam AF, Osman MM. The effect of *Ambrosia maritima* (Damsissa) on the viability of *Lymnaea cailliaudi*; an experimental study. J Egypt Soc Parasitol; 1994, 24 (3): 513-7.
- 161. Zidan ZH, Hafez AM, Abdel-Megeed MI, El-Emam MA, Ragab FM, El-Deeb FA. Susceptibility of *Biomphalaria alexandrina* to the plant *Azolla pinnata* and some herbicides in relation to infection with *Schistosoma mansoni* miracidia. J Egypt Soc Parasitol; 1998, 28 (1): 89-100.
- 162. Schall VT, Vasconcellos MC, Rocha RS, Souza CP, Mendes NM. The control of the schistosometransmitting snail *Biomphalaria glabrata* by the plant Molluscicide *Euphorbia splendens* var. hislopii (Syn *milli* Des. Moul): A longitudinal field study in an endemic area in Brazil. Acta Trop; 2001, 79 (2): 165-70.
- 163. Yadav SC, Jagannadham MV. Physiological molluscicidal and effects changes of crude Milin on latex Biomphalaria glabrata. Chemosphere; 2008, 71 (7): 1295-300.