MORPHOLOGICAL CHANGES IN ORAL MUCOSA OF RABBITS INDUCED BY LIGHT EMITTING DIODE (LED) USED AS DENTAL CURING LIGHT

¹ABDUL KHALIQ ²NADIA NASEEM ³RABIA ANJUM ⁴AH NAGI

ABSTRACT

A dental curing light is a piece of dental tool that is used for the hardening of light cure composites. Many studies have shown these lights induce changes in DNA, mitosis and mitochondria through free radicals production. Light emitting diode (LED) are most commonly used and claim less hazards to the adjacent soft tissues. Current study was therefore designed to observe the morphological changes induced by light emitting diode (LED) as a dental curing light source in oral mucosa of experimental animals.

Fifty rabbits were divided into 5 groups (4 experimental and 5th as a control group). Cervical margin of central incisors of each animal in the experimental groups was exposed 3 times with LED light; duration of each exposure being 40 seconds with a gap of 30 seconds. Punch biopsies were taken after 24 hours, 48 hours, 1week and 2weeks from group 1, 2, 3 and 4 animals respectively.

Results showed ulceration (4%), acanthosis and vascular pathological changes (100%), enlarge (bulbous) rete ridges (97.5%), basal layer vacuolization (85%), acantholysis (27.5%) and atypical mitosis (10%) in all the experimental animals. With passage of time, a significant increase (P=0.000) in frequency of basal cell hyperplasia (90% in group 1 to 100% in group 4) and basal layer atypia (70% in group 1 to 90% in group 4) was observed. While inflammation dropped from 100% in group 1 to 0% in group 4 (P=0.000) due to healing of tissues. Changes were similar to the previous studies except some severe effects like atypical mitosis and basal layer atypia were observed which may be attributed to increase in number of light exposures in our study that is in compliance with the clinical practice in our set up.

These findings may help in creating awareness among the dental practitioners to use dental curing lights with caution keeping appropriate safety measures for the adjacent oral soft tissue in consideration.

Key Words: Morphological changes, oral mucosa, light emitting diode.

INTRODUCTION

A dental curing light is a dental tool used for the hardening of light cure composites.¹ Light-emitting diode (LED) was introduced to overcome the problems associated with previous light sources. It provides blue visible light having a spectrum in the range of 450nm to 490nm.² Curing light is employed in many aspects

Received for Publication:February 23, 2016Approved:March 7, 2016

of modern dentistry like restorative, preventive and orthodontic dentistry.³ Usually, composite material is applied in the form of small increments which are cured each time except in very small restorations. In order to get better result of final restoration, low intensity should be used with extended intervals during which polymerization is allowed to continue.⁴ When using latest generation of LED units having light intensity of 1,500-2,000 mW/cm2 curing time can be reduced to 20 seconds.⁵ In addition to curing the material, this high intensity light can damage nearby tissues i.e. oral mucosa through its biological effects and rise in temperature.⁶

Experimental studies have shown that oral tissues when exposed to curing light, results in a T-cell induced inflammation.⁷ Due to the heat generated by the curing

¹ Abdul Khaliq, BDS, MPhil (Oral Pathology, University of Health Sciences, Lahore) **For Correspondece:** Abdul Khaliq, Dental Surgeon, RHC, Lal Sohanra, District Bahawalpur Cell: 0333-6395192

² Nadia Naseem, MBBS, MPhil, (Histopathology) PhD Scholar, Assistant Professor (University of Health Sciences, Lahore)

³ Rabia Anjum, BDS, MPhil (Oral Pathology, University of Health Sciences, Lahore)

⁴ AH Nagi, MB, PhD, FCPS, FRC Path, Head of Pathology Department, BDS, MPhil, (University of Health Sciences, Lahore

unit and the exothermic nature of the polymerization process itself, the rise in temperature may cause coagulation of protoplasm, expansion and outflow of fluid from the dentinal tubules, changes in blood vessel structure and tissue necrosis.⁸ Angiogenesis is normally a tightly regulated process that rarely occurs in the adult organism. There are certain pathological conditions including cancer, diabetic retinopathy, ischemic cardiovascular and chronic inflammatory diseases associated with aberrant angiogenesis. Angiogenesis is an essential process to supply nutrients, oxygen and growth factors thus supporting cellular function and survival. It involves the complex interaction between endothelial cells, extracellular matrix proteins and soluble factors in plasma.⁹ The present study we used rabbits of New Zealand breed, to study the angiogenic changes in oral mucosa with three exposures of the light each of 40sec with a gap of 30sec because morphological features of rabbits are closely similar to human gingival mucosa.

METHODOLOGY

This was an experimental study, conducted in the Experimental (Animal) Research Laboratory and the Department of Morbid Anatomy and Histopathology/ Oral Pathology, University of Health Sciences, Lahore. The sample size of 10 rabbits/ group was calculated by Gertsman formula.¹⁰ A total of 50 rabbits were divided randomly through balloting/lottery method into 5 groups, each consisted of 10 rabbits. One group was control and other four were experimental groups. The study animals were maintained in The Experimental Research Laboratory of University of Health Sciences, Lahore, under standard environment and diet, two weeks before start of experiment.

The Dental Curing Light, LED. B made by Guilin Woodpecker Medical Instrument Co., Ltd. Patent no. CN 200730092316.9 with light output 850mW/cm2-1000mW/cm2 and wave length 420nm to 480nm was used. Cervical margin of central incisors was exposed for 40 seconds, exposure was done 3 time with 30 sec gap between each exposure because composite filling is done in small increments in most of the cases but in earlier studies only one exposure for 40 seconds. Biopsies were taken after 24 hours, 48 hours, 1week and 2weeks from group 1, 2, 3 and 4 animals respectively. The specimens from the specified area i.e. oral mucosa adjacent to the maxillary central incisor were obtained after anesthetizing the animals. Representative tissue sections were taken and tissue specimens were fixed in 10% formalin and processed. Paraffin embedded blocks were made after taking these through ascending grades of ethanol. The sections 3-4 micro meter were cut using microtome and stained using Haemotoxylin and eosin stain and submitted for microscopic examination.

Statistical Analysis

The data were entered and analyzed using SPSS 18.0. Mean \pm SD was given for quantitative variables like weight and the frequencies along with percentages were given for qualitative variables like gender and morphological changes. Pearson Chi Square and Fisher exact test were applied to observe associations between groups and morphological changes. A P-value ≤ 0.05 was considered as statistically significant.

RESULTS

A total of n=50 rabbits including females = 11(22%)and males = 39(78%), with mean weight 1.765 ± 0.1767 kgs (Range: $1.5\rightarrow2.0$ kgs) were included. As regards pathological characteristics observed in the experimental animals, gross morphological features like consistency and no areas of necrosis were observed in both control and experimental animals while considering the colour of tissue it appeared as pink, reddish pink and reddish. The pink colour was present in 50.0%, 30.0%, 100.0% and 60.0% cases of group 1, 2, 3 and 4 respectively. Reddish pink colour was present in 20%cases of group 1 and 40% cases of both group 2 and 4 while group 3 did not show reddish pink color. Reddish colour was seen only in group 1 and 2 in 30% cases. Overall, the following microscopic changes were noted.

Within two weeks duration, a significant increase (P=0.000) in basal cell hyperplasia (90% in group1 to 100% in group4) was also observed. Acute inflammation showing predominantly neutrophilic infiltrate with macrophages was observed in 100% each of the animals in group1 and group 2 while only 30% and none of the animals in group 3 and 4 respectively. Regarding the vascular changes, angiogenesis and thickened vessel walls were found in 100% (Fig 3) while vascular congestion and dilatation was seen in 82.5% of the experimental animals.

DISCUSSION

This study was done to observe the histological changes in oral mucosa of rabbits induced by light emitting diode (LED) used as dental curing light. We used rabbits of New Zealand breed, as morphological features of rabbits are closely similar to human gingival mucosa as well as ease of their availability.¹¹

Pathological variables were studied and results were compiled. As regards the microscopic changes,

ulceration¹² was reported in the present study while none of other studies in the literature described such a change. It is usually believed that visible light sources (LEDs) are non-mutagenic but they can incite thermal energy in biological tissues which can cause burns and colour changes as observed in experimental animals (from pink to red). In the present study Acanthosis, the hyperplasia of prickle cell layer was observed in all

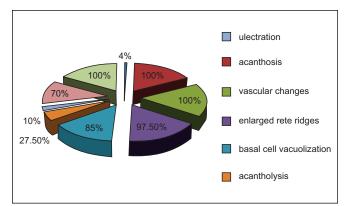


Fig 1: This figure shows distribution of different histological changes among n=40 experimental animals

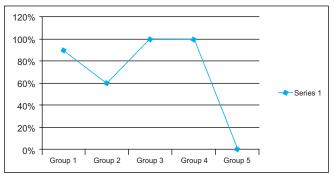


Fig 2: This figure shows frequency of basal cell hyperplasia in relation to time duration among different groups. Note that increase in frequency from group1 (90%) to group4 (100%) of experimental animals

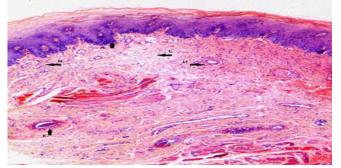


Fig 3: Photomicrograph of gingival mucosa of rabbit in experimental group 2 showing basal layer atypia (arrow A), angiogenesis (arrow B), stromal edema (arrow C) perivascular acute and chronic inflammatory cells (arrow D) and vessel wall thickness (arrow E). (H&E, 10 ×10x)

experimental groups (100%). Nearly all precancerous lesions (leukoplakia, erythroplakia and lichen planus) in their histology shows acanthotic change but it is reactive in nature in most of the cases.¹³ Similar changes were noted in studies performed by Mohamed and Rita in 1997 in Egypt and Swad in 2009 in Iraq, however, rats and rabbits were used as experimental animals respectively.^{14,15} In both studies a single light exposure of 40 seconds was applied. On contrary, a progressive decrease in epithelial thickness was observed in a study carried out by Murdiastuti and associates in 2010 in Indonesia.¹⁶ Acantholysis is the loss of intercellular bridges of cells of spinosum layer which can result in the formation of blisters and ulcers in oral mucosa.¹⁷ It was observed in 27.5% of experimental groups with no such change in control group (P=0.000). Mohamed and Rita documented similar change in oral mucosa in their experimental groups in their study performed in Egypt in 1997.¹⁴

Mittal and co-authors reported that when skin is exposed to UV radiations, infiltrating CDl lb+ inflammatory cells result in oxidative stress which overcome normal anti oxidative mechanism of skin leading to basal cell vacuolization observed in phototherapy.¹⁸ Same mechanism is involved in visible light through LED. In the present study, basal cell vacuolization was observed 80% with 100% each of group 1 and 2 while in 90% and 50% of group 3 and 4 respectively of experimental animals while no such change among control group was noted (P=0.000). Mohamed and Rita also documented such change.¹⁴ In present study basal cell hyperplasia was found but none of other studies related to our work described such a change. Basal layer atypia characterized by change in the basal cells showing pleomorphism, altered N/C ratio, prominent nucleoli, and increase in typical mitotic figures¹⁹ was reported in the present study. No study in the literature related to LED effects on oral mucosa described such a change. Atypical mitosis includes tripolar, multipolar and ring shaped mitotic bodies/ figures within the nucleus as described by Rubio.²⁰ In the present study it was observed predominantly in basal and lower 2/3rd of the keratinocytes of the mucosa of 10% animals of the total 40 experimental animals. While none of the control group animal depicted such change (P=0.783). Although finding is statistically not significant but increase in mitotic rate and presence of these atypical mitosis might be a significant pathological change if it persists even after cessation of stimulus. None of the studies done on Light curing effects on oral mucosa described such a change which might be

related because of a single exposure to light whereas in our study three exposures were done which is in compliance with the clinical practice on human patients in our clinical set up.

A part from epithelial changes, significant stromal alterations were also observed. Stromal oedema which was observed from 90% to 100% experimental animals it may signify a healing process. None of the control group animals showed this change (P=0.000). This finding is consistent with the results of the studies done by Mohamed and Rita as well as Swad.^{14,15}

Visible light from curing unit gives thermal and photo dynamic stimuli resulting in an acute inflammatory response.²¹ In present study acute inflammation with predominantly neutrophils and macrophages was observed in 100% animals in each in group 1 and 2 while in 30% and 0% of experimental animals in group 3 and 4 respectively. Results shows that in early 48 hours (group 1 and 2) there is maximum inflammatory response which decrease to 30% after 1week (group 3) and 0% after two weeks (group 4). None of the animals in the control group depicted such change (P=0.000). Murdiastuti and associates in 2010 at Indonesia reported similar finding in his study.¹⁶ Swad in 2009 at Iraq aslo observed inflammation in his study.¹⁵ Chronic inflammatory infiltrate predominantly lymphoplasmacytic and macrophages with plump fibroblast was observed in 100% of all groups of experimental animals, while none of the animals depicted such change in control group (P=0.000). Visible light from curing unit acts as irritant and result in vessel congestion and dilatation which results in an increase blood flow to that area helping in inflammatory process and to overcome oxidative stress. In present study vessel wall congestion and dilatation was observed 82.5% with 100% animals each in group 1, 2 and 4 while in 30% in group 3 animals. Findings are same in group 1, 2 and 4 (100%) while drop (30%) in group 3 may be due to host response. When compared to control group, none of the animal depicted such a change (P=0.000). Vessel wall thickness increased due to increased protein deposition (hyalinization) and also formation of neo intima. It's a healing response of vessel to any stimuli.²² It was in response to increasing exposure to dental curing light (LED) and reported in 100% animals of all the experimental groups, while none of control animals depicted such a change (P=0.000).

CONCLUSION

To date, much data is available on the efficacy of dental curing light and composite material but little focus has been given to the hazardous effects of curing light on the oral mucosa adjacent to the tooth being irradiated. Results of the present study were similar to the previous studies except some severe effects like atypical mitosis, basal layer atypia and basal cell hyperplasia are exclusively reported in the present study, which may be attributed to increase in number of exposure in the present study, compliant with the usual clinical procedure.

Stromal inflammation and vascular pathological changes including angiogenesis were also significantly observed in all experimental group animals. To our knowledge no such study is done in Pakistan as well as no relevant data in the previous literature describing the actual statistics related to the histological changes in experimental animals exposed to LED has been found. So, the present study not only is an important addition in terms of reporting the statistically significant histological findings in oral mucosa after exposure to LED light source but also may help in creating awareness among dental practitioner to use dental curing light (LED) with caution maintaining protective measures for the adjacent oral soft tissue in consideration.

Acknowledgments

The authors acknowledge the encouragement extended by the Vice Chancellor of UHS, Lahore. Also the laboratory staff of Oral Pathology Dept of UHS, Lahore for their technical and logistic support.

REFERENCES

- 1 Sherwood, and Anand. Essentials of Operative Dentistry. St. Louis, MO. Jaypee Brothers Medical Publishers. 2010.
- 2 D'Alpino PH, Wang L, Rueggeberg FA, et al. Bond strength of resin-based restorations polymerized with different light-curing sources. J Adhes Dent. 2006; 8(5): 293-98.
- 3 Jung H, Friedl KH, Hiller KA, Furch H, Bernhart S, Schmalz G. Polymerization efficiency of different photocuring units through ceramic discs. Oper Dent. 2006; 31: 68-77.
- 4 Apicella A, Simeone M, Aversa R, Lanza A, Apicella D. Light shielding effect of overlaying resin composite on the photopolymerization cure kinetics of a resin composite and a dentin adhesive. Dent Mater. 2005; 10: 954-61.
- 5 Ernst CP, Meyer GR, Muller J, Stender E, Ahlers MO, Willershausern B. Depth of cure of LED vs QTH light-curing devices at a distance of 7 mm. J Adhes Dent. 2004; 6: 141-50.
- 6 Bruzell Roll E, Jacobsen N, Hensten-Pettersen A. Health hazards associated withcuring light in the dental clinic. Clin Oral Investig. 2004; 8: 113-17.
- 7 Ahlfors EE, Roll EB, Dahl JE, Lyberg T and Christensen T. Blue light exposure of the oral mucosa induces T cell-based inflammatory reactions. In: Book of Abstracts of the 13th International Congress on Photobiology and 28th Annual Meeting of the American Society for Photobiology. AIP (Association Internationale de Photobiologie). 2000; 32: 106-07.

- 8 Knežević A, Tarle Z, Meniga A, Šutalo J and Pichler G. Influence of light intensity from different curing units upon composite temperature rise. J. Oral Rehabil., 2005; 32: 362-67.
- 9 Sappayatosok K , Maneerat Y, Swasdison S, Viriyavejakul P, Dhanuthai K, Zwang J, Chaisri U. Expression of pro-inflammatory protein, iNOS, VEGF and COX-2 in Oral Squamous Cell Carcinoma (OSCC), relationship with angiogenesis and their clinico-pathological correlation . Medicina Oral, Patologia Oral y Cirugia Bucal. 2009; Vol 14. Pages E319-E324.
- 10 Gertsman BB. Basic Biostatics, Statistics for public health practice 4th edit. Sudbury, Jones & Bastlett. 2008.
- 11 Nanci, A. Ten Cate's Oral Histology, Development, Structure and Function 7th edition. Chapter 12: Oral Mucosa. 2008; 331-32.
- 12 WHO\IARC classifies electromagnetic fields as possibly carcinogenic to human. Press release Lyon, France. 2011.
- 13 Speight PM, Farthing PM and Bouquot JE. The pathology of oral cancer and precancer. Curr. Diag. Path. 1996; 3: 165-77.
- 14 Mohamed AA and Rita MK. East. Mediterr. Health J. 1997; 3(3): 540-48.
- 15 Swad AA. Effect of visible light emitted from the cure gun used as filling hardener on the oral mucosa of rabbits. Iraqi J. Vet. Sci. 2009; 23: 163-66.

- 16 Murdiastuti K, Suryono, Aini M, Mefi PS and Rani G. The Effect of Visible Light Cure (VLC) Exposure to Gingival Tissue's Sprague dawley Rats. The Indo. J. Dent. Res. 2011. 1(2), 78-83.
- 17 Yeh SW, Ahmed B, Sami N and Ahmed AR. Blistering disorders: diagnosis and treatment. Derma. Ther. 2003; 16(3): 214-23.
- 18 Mittal A, EImets CA and Katiyar SK. CDI Ib+ cells are the major source of oxidative stress in UV radiation-irradiated skin: possible role in photoaging and photocarcinogenesis. Photochem. Photobiol. 2003; 77: 259-64.
- 19 Barnes L, Eveson JW, Reichart P. and Sidransky D. WHO classification of tumours: pathology and genetics of head and neck tumours. Lyon: ARC Press. 2005; 163-208.
- 20 Rubio CA, Yo K and Tomoyuki K. Frequency of Atypical Mitosis in Intestinal Metaplasia of Gastric Mucosa in Japanese Patients. Jpn. J. cancer Res. 1994; 85: 284-89.
- 21 Guyton AC and Hall JE. 9th Ed. EGC, Jakarta. 1997; 543.
- 22 Sata M. Role of circulating vascular progenitors in angiogenesis, vascular healing, and pulmonary hypertension: lessons from animal models. Arterioscler Thromb Vasc Biol. 2006; 26: 1008.

CONTRIBUTION BY AUTHORS	
1 Abdul Khaliq:	Conception, synthesis and planning of the research, active participation in methodology, interpretation, analysis and discussion
2 Nadia Naseem:	Conception, synthesis and planning of the research interpretation, analysis and discussion.
3 Rabia Anjum:	Conception, synthesis and planning of the research, active participation in methodology, interpretation, analysis and discussion.
4 AH Nagi:	Conception, synthesis and planning of the research, interpretation, analysis and discussion.