

# ***In vitro* antibacterial susceptibility of different brands of oral levofloxacin 250 mg tablet against *Staphylococcus aureus* and *Escherichia coli***

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**Abstract:** Antibiotics are not only used in morbidity but also help in prevention of infection. The irrational use of broad spectrum antibiotics is now increasing the resistance against pathogens. This present study has been carried out to evaluate the *in-vitro* antibacterial effect of levofloxacin against clinical isolates. According to Clinical and Laboratory Standards Institute (CLSI) guidelines, minimum inhibitory concentrations 90% (MIC<sub>90</sub>) of the levofloxacin tested were evaluated by an agar dilution method. Total 63 clinical isolates *Staphylococcus aureus* (n=34) and *Escherichia coli* (n=29) were collected from different hospitals at Karachi and were evaluated MIC<sub>90</sub> of eleven different brands of levofloxacin tablet (250 mg). Levofloxacin (Reference) was tested against *E.coli* standard (American Type Culture Collection) (ATCC=25922) with (MIC<sub>90</sub>; 0.03µg/ml) and compared with different eleven brands of levofloxacin tablets 250mg (MIC<sub>90</sub>; 0.5µg/ml -16.0µg/ml). Levofloxacin (Reference) sensitivity against *S. aureus* standard (ATCC=25923) is (MIC<sub>90</sub>; 0.12µg/ml) and similarly when it was compared with same levofloxacin tablets (MIC<sub>90</sub>; 0.5-16.0µg/ml). It has been concluded by the present study, a large number of strains of bacteria have shown better bactericidal action of different brands of levofloxacin and nearly all commercialized drugs were appropriate for therapeutic use.

**Keywords:** Levofloxacin, generics, *in vitro*, antibacterial activity, susceptibility.

## **INTRODUCTION**

Microorganisms are ubiquitous found from shallow of seas to air. These may be pathogenic or beneficial for human being (Arsalan *et al.*, 2013a, 2013b, 2013c, 2013d). Antibiotics are not only used in morbidity but also help in prevention of infection (Arsalan *et al.*, 2014a, 2014b). The irrational use of broad spectrum antibiotics has now increased the resistance against pathogens (Arsalan *et al.*, 2010, 2015, 2017). Antibiotics are the most often given drug in modern medicine to treat and/or prevent disease due to its bactericidal or bacteriostatic effect (Arsalan *et al.*, 2016). Quinolones have been probably the fastest growing group of antibiotics with broader and wider spectrum of anti-bacterial activity (Bhanot *et al.*, 2001; Ahmad *et al.*, 2016). Among quinolones, levofloxacin third generation has been possessed wide range of bactericidal effect against Gram-negative and positive and atypical pathogens. Levofloxacin has shown its lethal activity by inhibiting enzymes topoisomerase IV and DNA gyrase against virulent and resistant organisms like *Pseudomonas aeruginosa*, methicillin-resistant *Staphylococcus aureus*, *Mycobacterium tuberculosis*, penicillin-resistant *Streptococcus pneumoniae*, *Escherichia coli* (Zhanel *et al.*, 2002; Stein and Goldstein, 2006). Therapeutically, it is used for urinary tract infections (UTIs); respiratory tract infections (RTIs); biliary tract infections (BTIs); sinusitis;

chronic bronchitis; pneumonia and uncomplicated mild to moderate infection of skin and skin structure (Blondeau 1999; File 2004; Prajapati *et al.*, 2008). Orally levofloxacin has been well absorbed, because tissue and fluid concentrations often exceed the serum drug concentration; peak plasma concentration has been usually attained within one to two hours (Zhanel and Noreddin, 2001).

Among Gram-positive *S. aureus* infections have been a major cause of morbidity and mortality worldwide (Bishop and Howden, 2007). *S. aureus* may cause severe infections like endocarditis, pneumonia, osteomyelitis, septicemia (Gagliardi *et al.*, 1998, Mader *et al.*, 2000). Since last two decades, the resistance of *S. aureus* to various antibiotics has been raised. The incidence has been accompanied by a rise in antibiotic-resistant strains particularly, methicillin-resistant *Staphylococcus aureus* (MRSA) and more recently, vancomycin-resistant *Staphylococcus aureus* (VRSA) (Stevenson *et al.*, 2005). *Escherichia coli* is a Gram-negative bacilli and one of the most frequent clinical isolated pathogen has been involved in infections like cholangitis, pneumonia, meningitis, urinary tract infections (UTI), gastro-intestinal infections (GII) and diarrhea (Bano *et al.*, 2011). The resistance of *E. coli* against levofloxacin has been observed by several workers (Tenover 2006; Reinert *et al.*, 2007; Jang *et al.*, 2011).

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## MATERIALS AND METHODS

Standard (STD) levofloxacin drug was kindly gifted from M/s *Sanofi-Aventis (Pakistan) Limited*. Twelve different marketed Levofloxacin were purchased from local pharmacies of Karachi, Pakistan. STD Gram-negative (American Type Culture Collection; ATCC) (*E. coli*, ATCC® 25922) and Gram positive (*S. aureus*, ATCC® 25923) have been kindly gifted by M/s. Brookes Pharmaceutical (Private) Limited. The clinical isolates were obtained from January 2013 to August 2013 from different hospitals at Karachi, Pakistan. Mueller-Hilton broth (Merck Germany) and freshly boiled distilled water was always used to prepare different dilution (Wiegand *et al.*, 2008).

### **Instrumentation**

The determination of MICs of levofloxacin drug and their dosage form a Shimadzu UV-1601 spectrophotometer was used for quantitative calculation.

### **Broth dilution method**

A complete protocol has been found in the clinical and laboratory standards institute (CLSI), for broth dilution methods by reducing the quantity of the anti-bacterial agent to be evaluated, generally prepared in sequential two fold dilutions in broth tubes (Andrews, 2001).

### **Preparation of MacFarland standard**

Sulphuric acid 1% is prepared and 1.175% aqueous solution of barium chloride is added in it. Now with constant slow agitation, add the designated amounts of the two solutions to the tubes make a total of 10 ml per tube. The suspended barium sulfate precipitate corresponds approximately to homogenous *E. coli* cell densities per ml throughout the range of standard (Wiegand *et al.*, 2008).

### **Preparation of inoculum**

These suspensions were prepared by using the top of the colonies of the standard and isolated microorganisms. STD *E. coli* (ATCC® 25922) and STD *S. aureus* (ATCC® 25923) and clinical isolates of *E. coli* and *S. aureus* were incubated in test tubes at 37°C for 2-8 hours until the turbidity exceeds that of 0.5 McFarland standards (Wiegand *et al.*, 2008).

### **Preparation of antibiotic stock solutions**

Weighed accurately for a required amount of standard antibiotic powder (standard powder of levofloxacin and its different brands). Prepare stock solution using the formula (Wiegand *et al.*, 2008).

$$W = \frac{V \times C \times 1000}{P}$$

Where V is volume in ml required,  
W is weight of the antimicrobial to be dissolved in V,  
C is final concentration of solution and  
P is potency of the antibiotic base.

### **Preparation of antibiotic dilution range**

Generally prepared a series of varying concentrations two fold serial dilutions (0.03, 0.06, 0.12, 0.25, 0.5, 1, 2, 4, 8, 16 µg/ml). Equal volume of inoculum has been added in test tubes (Wiegand *et al.*, 2008).

### **Procedure**

Prepared Mueller-Hilton broth, arranged sufficient sterile test tubes for levofloxacin to cover the range of antibiotic dilutions. 9ml of broth and 1ml of each antimicrobial agent's dilution was transferred in separate broth tubes. Now inoculum was added to each anti-microbial containing tube in the dilution series. The tubes were incubated at 37°C for 12 to 18 hours. Examined the tubes with visually and with spectrophotometer (546 nm) for the existence or absence of microbial growth and compared the result with the growth in the control tube.

## STATISTICAL ANALYSIS

The data were analyzed by one way ANOVA (by Graphpad software, Quick calcs online calculator for scientists).

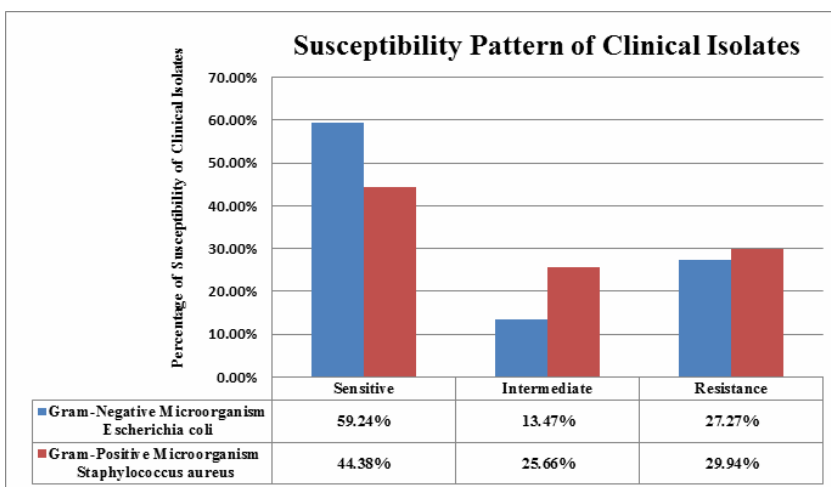
## RESULTS

In our study, total 63 most common clinical isolates *Staphylococcus aureus* (n=34) and *Escherichia coli* (n=29) were collected from different hospitals at Karachi, Pakistan as shown in table 1 and MIC<sub>90</sub> were evaluated against eleven different brands of levofloxacin tablet (250 mg) has been shown in table 2 and table 3. The overall resistance pattern of Gram-negative *E. coli* and Gram-positive *S. aureus* clinical isolates has been illustrated in table 4 and fig. 1.

Levofloxacin (Reference) was tested against *E.coli* standard (ATCC = 25922) with (MIC<sub>90</sub>; 0.03µg/ml) and compared with different eleven brands of levofloxacin tablets 250mg (MIC<sub>90</sub>; 0.5µg/ml -16.0µg/ml). Levofloxacin (Reference) sensitivity against *S. aureus* standard (ATCC=25923) is (MIC<sub>90</sub>; 0.12µg/ml) and similarly when it was compared with same levofloxacin tablets (MIC<sub>90</sub>; 0.5- 16.0µg/ml).

## DISCUSSION

*In-vitro* analysis of antibacterial activity has been conducted commonly, as consequences of the antimicrobial susceptibility have been done to observe antibiotic efficacy inside the body (Hannan *et al.*, 2008). For a long period of time, resistance of bacteria to antibiotics has been appeared as one of the major troubles encountered by health associated professionals (Arsalan *et al.*, 2014). Levofloxacin is a third generation quinolone, exhibits fine bactericidal activity against Gram-positive and Gram-negative. Because of its wide spectra even



**Fig. 1:** Susceptibility Pattern of Clinical Isolates against Marketed Levofloxacin 250 mg tablets

**Table 1:** Summary of Clinical Isolates

Clinical Isolates	Source of Isolates	Number of Isolates
<i>Staphylococcus aureus</i>	Surgical, burn and accidental wound pus, pneumonia, blood sample	41
<i>Escherichia coli</i>	Stool and urine, blood sample	37

**Table 2:** Comparison of standard levofloxacin and different brands of levofloxacin against standard (STD) *Escherichia coli* & different clinical isolates of *Escherichia coli*

MICROBES n=29	LEVOFLOXACIN (µg/ml)											
	STD LF	LF 01	LF 02	LF 03	LF 04	LF 05	LF 06	LF 07	LF 08	LF 09	LF 10	LF 11
STD <i>E. coli</i> ATCC 25922	0.03	0.06	0.06	0.12	1.0	0.5	0.5	0.03	0.06	0.25	0.25	0.12
Ec 01	0.25	0.5	0.5	1	0.5	1	1	4	1	1	16	1
Ec 02	0.25	4	1	1	16	16	0.5	1	1	16	16	1
Ec 03	0.5	1	4	1	1	16	16	1	1	1	16	1
Ec 04	0.25	0.5	0.5	1	16	1	1	1	0.5	0.5	1	16
Ec 05	0.06	4	1	1	0.5	16	16	1	1	1	1	16
Ec 06	0.5	0.5	1	16	1	8	16	16	1	16	1	2
Ec 07	0.5	0.5	4	1	16	16	1	2	16	4	1	1
Ec 08	0.25	1	1	4	16	1	4	1	0.5	16	1	1
Ec 09	0.12	1	1	16	1	1	1	1	8	1	16	1
Ec 10	0.12	16	0.5	0.5	8	16	4	1	1	1	1	1
Ec 11	0.25	1	1	0.5	1	16	1	4	16	16	4	16
Ec 12	0.25	0.5	1	16	16	4	1	16	4	1	1	1
Ec 13	0.12	1	0.5	1	1	16	1	1	1	16	8	1
Ec 14	0.06	16	4	1	0.5	8	0.5	1	16	4	0.5	1
Ec 15	1	16	16	1	16	16	16	16	2	16	1	16
Ec 16	0.5	16	0.5	1	8	16	16	1	16	0.5	16	1
Ec 17	0.06	1	1	4	16	1	1	16	0.5	1	4	1
Ec 18	0.06	4	16	1	1	0.5	16	0.5	0.5	4	0.5	1
Ec 19	0.12	0.5	1	16	1	1	4	16	1	1	16	8
Ec 20	0.5	1	4	16	16	16	1	0.5	16	1	1	1
Ec 21	0.12	16	1	4	16	1	1	4	1	1	1	16
Ec 22	0.25	4	1	1	0.5	1	16	16	1	16	1	1
Ec 23	0.12	1	1	4	16	4	1	16	16	1	4	0.5
Ec 24	0.06	1	16	1	1	1	16	4	16	1	1	16
Ec 25	0.5	1	1	16	16	16	16	0.5	1	1	0.5	1
Ec 26	0.5	1	1	16	16	1	16	1	0.5	16	4	4
Ec 27	0.06	0.5	1	1	16	1	1	1	1	16	1	1
Ec 28	0.12	1	4	1	1	4	1	16	1	0.5	1	4
Ec 29	0.5	4	1	0.5	1	16	16	1	1	1	16	1

**Table 3:** Comparison of standard levofloxacin and different brands of levofloxacin against standard (STD) *Staphylococcus aureus* and different clinical isolates of *Staphylococcus aureus*

MICROBES n=34	LEVOFLOXACIN (µg/ml)											
	STD LF	LF 01	LF 02	LF 03	LF 04	LF 05	LF 06	LF 07	LF 08	LF 09	LF 10	LF 11
STD. <i>S. aureus</i> ATCC 25923	0.12	0.12	0.5	0.12	0.12	0.5	0.25	0.25	0.12	0.5	0.12	0.25
Sa 01	0.12	0.5	8	1	4	8	1	8	8	2	1	0.5
Sa 02	0.25	0.5	4	0.5	4	8	1	0.5	2	1	8	8
Sa 03	0.25	4	1	2	0.5	8	4	1	0.5	2	1	4
Sa 04	0.12	2	0.5	0.5	1	0.5	8	4	0.5	0.5	2	0.5
Sa 05	0.5	8	8	0.5	0.5	8	0.5	0.5	0.5	8	8	8
Sa 06	0.12	0.5	0.5	1	0.5	2	8	0.5	2	1	1	8
Sa 07	0.12	0.5	0.5	4	0.5	8	2	1	0.5	2	0.5	0.5
Sa 08	0.25	1	4	8	1	0.5	0.5	0.5	0.5	0.5	8	0.5
Sa 09	0.5	1	0.2	0.5	0.5	8	8	0.5	8	2	0.5	0.5
Sa 10	0.5	0.5	8	1	0.5	8	8	0.5	0.5	8	0.5	8
Sa 11	0.25	0.5	0.5	8	0.5	8	0.5	8	0.5	1	0.5	0.5
Sa 12	0.12	0.5	1	8	0.5	8	0.5	8	1	8	0.5	0.5
Sa 13	0.5	8	0.2	0.5	0.5	1	0.5	1	2	0.2	0.5	8
Sa 14	0.25	0.5	8	1	8	8	1	0.5	1	4	8	0.5
Sa 15	0.12	0.5	8	0.5	1	8	0.5	2	0.5	0.5	8	1
Sa 16	0.25	4	0.5	8	0.5	8	8	8	0.5	0.5	0.5	0.5
Sa 17	0.12	1	8	1	8	2	0.5	0.5	2	0.5	8	0.5
Sa 18	0.12	0.5	2	8	0.5	0.5	0.5	8	0.5	8	0.5	0.5
Sa 19	0.5	8	8	0.5	8	4	8	8	2	8	0.5	2
Sa 20	0.25	0.5	0.5	8	0.5	0.5	0.5	1	0.5	0.5	0.5	4
Sa 21	0.12	0.5	8	0.5	0.5	1	0.5	0.5	0.5	8	0.5	8
Sa 21	0.25	8	0.5	8	0.5	0.5	2	0.5	1	0.5	1	0.5
Sa 23	0.5	0.5	1	0.5	0.5	8	8	8	8	0.5	8	0.5
Sa 24	0.12	0.5	0.2	0.5	2	8	0.5	1	8	0.5	0.5	2
Sa 25	0.25	4	0.5	0.5	1	2	0.5	0.5	0.5	8	8	0.5
Sa 26	0.25	8	0.5	8	0.5	8	0.5	2	0.5	8	0.5	8
Sa 27	0.12	0.5	0.5	1	8	8	0.5	0.5	8	0.5	8	2
Sa 28	0.12	0.5	8	0.25	0.5	0.5	8	8	1	0.5	0.5	0.5
Sa 29	0.12	0.5	8	0.5	1	8	0.5	0.5	0.5	8	0.5	8
Sa 30	0.25	1	2	0.5	8	0.5	8	0.5	0.5	0.5	8	0.5
Sa 31	0.12	8	8	0.5	2	0.5	0.5	1	0.5	0.5	1	8
Sa 32	0.25	0.5	0.5	8	0.5	0.5	2	8	0.5	2	8	8
Sa 33	0.12	0.5	8	0.5	0.5	1	8	8	0.5	8	0.5	8
Sa 34	0.25	8	8	0.5	0.5	8	0.5	8	8	8	0.5	2

**Table 4:** Resistance Pattern of *Escherichia coli* and *Staphylococcus aureus* against Eleven Brands of Marketed Levofloxacin

Resistance Pattern	Gram-Negative Microorganism <i>Escherichia coli</i>	Gram-Positive Microorganism <i>Staphylococcus aureus</i>
Sensitive	59.24%	44.38%
Intermediate	13.47%	25.66%
Resistance	27.27%	29.94%

against resistant pathogens, it has been recommended in severe infections like community acquired pneumonia (CAP) (Mandell *et al.*, 2007), nosocomial pneumonia

(Welte and Pletz, 2010), inhalation anthrax (Li *et al.*, 2010), treatment of pulmonary infections due to *Pseudomonas aeruginosa* and other bacteria in patients

with cystic fibrosis (Mesaros *et al.*, 2007). Due to irrational use of Levofloxacin and change in serotype and source of microbes, there is a change in resistance pattern of clinical isolates as shown in various studies (Bucaneve *et al.*, 2005; MacDougall *et al.*, 2005; Lockhart *et al.*, 2007). According to CLSI (2011), for *E. coli* MIC<sub>90</sub> ≤ 2µg/ml possessed susceptible, 4 to 8µg/ml and ≥8µg/ml has intermediate and resistant concentration, respectively, in broth dilution method of levofloxacin, while ≤1µg/ml has susceptibility, 2 to 4µg/ml and ≥4µg/ml have intermediate and resistant concentration of levofloxacin against *S. aureus*.

Antimicrobial assay of different marketed levofloxacin by broth dilution method was evaluated against standard *S. aureus* (ATCC 25923) and *E. coli* (ATCC 25922) and their clinical isolates. The broth dilution tests have been allowed the determination of the MIC<sub>90</sub> of an antimicrobial agent (Basset *et al.*, 2011). The broth dilution test has been specified that the MIC<sub>90</sub> of standard levofloxacin against standard *E. coli* (MIC<sub>90</sub> = 0.03µg/ml) indicated by Drago *et al.* (2002). The present studies has shown that resistance against levofloxacin has acquired by *E. coli* and its clinical isolates, significant variations have been found in MIC<sub>90</sub> of different marketed levofloxacin (MIC<sub>90</sub> = 0.5µg/ml-16.0µg/ml). Soriano and co-workers (2005) have been found MICs of standard levofloxacin (MIC<sub>90</sub> = 0.12µg/ml) against standard *S. aureus*. The present study has prevailed that different brands of 250mg levofloxacin tablets has shown resistance acquired by *S. aureus* and significant variations has been found with (MIC<sub>90</sub> = 0.5µg/ml-8.0µg/ml) against different clinical isolates of *S. aureus*.

The resistance depend upon the source of infections; serotype; genes of microbes; environmental factors; immunity due to socioeconomic conditions (Goossens *et al.*, 2005; Erb *et al.*, 2007; Shakya *et al.*, 2013). In our study, clinical isolates of *E. coli* and *S. aureus* have possessed 27.5% supported by Noviello *et al.* (2006) and Nasiri *et al.* (2013), in contrast to present study Woo *et al.* (2011) found 37.94% and Zemkova *et al.*, 2007 reported 11%.

Marangon and co-workers (2004) reported 25.5% resistance of *S. aureus* against levofloxacin, similarly 23% resistance has been reported by Shafiq *et al.* (2012). These studies have been supported our study in which 29.94% has been a positive sign of resistance while a fire alarming situation for health associated professionals is that only 44.38% sensitivity of *S. aureus* against levofloxacin has been noticed. In the past half a century, there has been a constant increase in the use of antibiotics for the treatment of bacterial infections. The continuous use and often abuse of the drug have led to an increased resistance to levofloxacin.

## CONCLUSION

The prime object of all pharmaceutical industries is to provide quality medicines with efficacy and safety profile. It has been concluded from the present study that there were no significant variations found in MIC<sub>90</sub> values of different brands of levofloxacin which has been indicated that the levofloxacin tablets manufactured either by multinational or local pharmaceutical industries has been produced approximate similar results. The approximate similarity in results of MIC<sub>90</sub> of levofloxacin 250 mg tablets of pharmaceutical industries in Pakistan is mainly due to fulfill the requirements of drug regulatory authorities.

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## REFERENCES

- Ahmad I, Arsalan A, Ali SA, Bano R, Minir I, Arif Sabah A (2016). Formulation and stabilization of norfloxacin in liposomal preparations. *Eur. J. Pharm. Sci.*, **91**: 208-215.
- Andrews JM (2001). Determination of minimum inhibitory concentrations. *J. Antimicrob. Chemother.*, **48**(Suppl. SI): 5-6.
- Arsalan A, Sabah A, Ahmad I, Naqvi SBS and Ali SI (2010). The role of antiseptic in surgery. *J. Baqai Med. Uni.*, **13**(2): 47-54.
- Arsalan A, Naqvi SBS, Ali SI and Anwar Z (2013a). Contamination of microorganisms in pediatric infant formula marketed in Karachi. *Annals Food Sci. Technol.*, **14**(2): 318-326.
- Arsalan A, Anwar Z, Ahmad I, Saba A and Naqvi SBS (2013b). Microbes in pediatric infant formula. *Annals Food Sci. Technol.*, **14**(1): 90-99.
- Arsalan A, Anwar Z, Ahmad I, Shad Z and Ahmed S (2013c). *Cronobacter sakazakii*: An emerging contaminant in Pediatric infant milk formula. *Int. J. Pharm. Res.*, **4**(4): 17-22.
- Arsalan A, Alam M, Naqvi SBS, Ahmad I and Anwar Z (2013d). Oxygen as a facilitator in the reduction of surgical site infections. *J. Uni. Med. Dent. Coll.*, **4**(2): 1-8.
- Arsalan A, Naqvi SBS, Sabah A, Bano R and Ali SI (2014a). Resistance pattern of clinical isolates involved in surgical site infections. *Pak. J. Pharm. Sci.*, **27**(1): 97-102.
- Arsalan A, Naqvi SBS, Iqbal A and Shakeel O (2014b). Temperature monitoring of vaccines' storage compartments in different health centres and pharmacies at Karachi, Pakistan. *Int. J. Pharm. Teach. Practic.*, **5**(3): 984-988.

- Arsalan A, Naqvi SBS, Ali SA, Ahmed S and Shakeel O (2015). *In vitro* bactericidal activity of cefepime and ceftiofloxacin against clinical isolates at Karachi. *Pak. J. Pharm. Sci.*, **28**(3): 841-847.
- Arsalan A, Shayam SB, Pasha R and Shakeel O (2016). First generation and fourth generation cephalosporin susceptibility against clinical isolates. *J. Uni. Med. Dent. Coll.*, **7**(2): 52 – 58.
- Arsalan A, Ahmad I and Ali SA (2017). Cefaclor: clinical, biochemical, analytical and stability aspects. In: *Advances in Medicine and Biology*. Berhardt LV (Ed.) Nova Science Publishers, Inc., New York, USA. Vol. 123, pp. 1-52.
- Bano R, Arsalan A, Shad Z and Ahmad I (2011). Levofloxacin: A broad spectrum potent antibiotic. *J. Baqai Med. Univ.*, **14**(1): 29-37.
- Basset P, Feil EJ, Zanetti G and Blanc DS (2011). In: *Genetics and Evaluation of Infectious Disease*. Tibayreng M (Ed.) Elsevier London, 1<sup>st</sup> ed., pp.669-675.
- Bhanot SK, Singh M and Chatterjee NR (2001). The Chemical and Biological Aspects of Fluoroquinolones: Reality and Dreams. *Curr. Pharm. Des.*, **7**: 313-337.
- Bishop EJ and Howden BP (2007). Treatment of *Staphylococcus aureus* infections: New issues, emerging therapies and future directions. *Expert Opin. Emerg. Drugs*, **12**(1): 1-22.
- Blondeau JM (1999). Expanded activity and utility of the new fluoroquinolones: A review. *Clin. Ther.*, **21**(1): 3-40.
- Bucaneve G, Micozzi A, Menichetti F, Martino P, Dionisi MS, Martinelli G, Allione B, D'Antonio D, Buelli M, Nosari AM, Cilloni D, Zuffa E, Cantaffa R, Specchia G, Amadori S, Fabbiano F, Delilieri GL, Lauria F, Foà R, Del Favero A and Gruppo Italiano Malattie Ematologiche dell'Adulto (GIMEMA) Infection Program (2005). Levofloxacin to prevent bacterial infection in patients with cancer and neutropenia. *N. Engl. J. Med.*, **353**(10): 977-987.
- Clinical and Laboratory Standards Institute (CLSI) (2011). Performance Standards for Antimicrobial Susceptibility Testing; Twenty-First Informational Supplement. **31**(1) Approved standard Approved standard M100-S21, Wayne, PA, USA.
- Drago L, De Vecchi E, Lombardi A, Nicola L, Valli M and Gismondo MR (2002). Bactericidal activity of levofloxacin, gatifloxacin, penicillin, meropenem and rokitamycin against *Bacillus anthracis* clinical isolates. *J. Antimicrob. Chemother.*, **50**(6): 1059-1063.
- Erb A, Sturmer T, Marre R and Brenner H (2007). Prevalence of antibiotic resistance in *Escherichia coli*: overview of geographical, temporal and methodological variations. *Eur. J. Clin. Microbiol. Infect. Dis.*, **26**(2): 83-90.
- File Jr TM (2004). New Insights in the Treatment by Levofloxacin. *Chemotherapy*, **50**(1): 22-28.
- Fu KP, Lafredo SC, Foleno B Isaacson DM, Barrett JF, Tobia AJ and Rosenthale ME (1992). *In vitro* and *in vivo* antibacterial activities of levofloxacin (1-ofloxacin), an optically active ofloxacin. *Antimicrob. Ag. Chemother.*, **36**: 860-866.
- Gagliardi JP, Nettles RE, McCarty DE, Sanders LL, Corey GR and Sexton DJ (1998). Native valve infective endocarditis in elderly and younger adult patients: comparison of clinical features and outcomes with use of the Duke Criteria and the Duke Endocarditis Database. *Clin. Infect. Dis.*, **26**: 1165-1168.
- Gesu GP, Marchetti F, Piccoli L and Cavallero A (2003). Levofloxacin and ciprofloxacin *in vitro* activities against 4,003 clinical bacterial isolates collected in 24 Italian laboratories. *Antimicrob. Ag. Chemother.*, **47**(2): 816-819.
- Goossens H, Ferech M, Stichele RV and Elseviers M (2005). Outpatient antibiotic use in Europe and association with resistance: A cross-national database study. *Lancet*, **365**(9459): pp.579-87.
- Gums JG (2002). Assessing the impact of antimicrobial resistance. *Am. J. Healthm Syst. Pharm.*, **59**(3): 4-6.
- Hannan A, Saleem S, Chaudhary S, Barkaat M and Arshad MU (2008). Anti Bacterial Activity of Nigella Sativa against Clinical Isolates Of Methicillin Resistant *Staphylococcus aureus*. *J. Ayub Med. Coll. Abbottabad.*, **20**(3): 72-74.
- Jang WH, Yoo DH and Park SW (2011). Prevalence of and Risk Factors for Levofloxacin-Resistant *E. coli* Isolated from Outpatients with Urinary Tract Infection. *Korean J. Urol.*, **52**(8): 554-559.
- Li F, Nandy P, Chien S, Noel GJ and Tornoe CW (2010). Pharmacometrics-based dose selection of levofloxacin as a treatment for post exposure inhalational anthrax in children. *Antimicrob. Agents Chemother.*, **54**(1): 375-379.
- Lockhart SR, Abramson MA, Beekmann SE, Gallagher G, Riedel S, Diekema DJ, Quinn JP and Doern GV (2007). Antimicrobial resistance among Gram-negative bacilli causing infections in intensive care unit patients in the United States between 1993 and 2004. *J. Clin. Microbiol.*, **45**(10): 3352-3359.
- MacDougall C, Powell JP, Johnson CK, Edmond MB and Polk RE (2005). Hospital and community fluoroquinolone use and resistance in *Staphylococcus aureus* and *Escherichia coli* in 17 US hospitals. *Clin. Infect. Dis.*, **41**(4): 435-440.
- Mader JT, Shirliff ME, Bergquist S and Calhoun JH (2000). Bone and joint infections in the elderly: Practical treatment guidelines. *Drugs Aging*, **16**: 67-80.
- Mandell LA, Wunderink RG, Anzueto A, Bartlett JG, Campbell GD, Dean NC, Dowell SF, File TM Jr, Musher DM, Niederman MS, Torres A and Whitney CG (2007). Infectious Diseases Society of America and American Thoracic Society consensus guidelines on

- the management of community-acquired pneumonia in adults. *Clin. Infect. Dis.*, **44**(2): S27-72.
- Marangon FB, Miller D, Muallem MS, Romano AC and Alfonso EC (2004). Ciprofloxacin and levofloxacin resistance among methicillin-sensitive *Staphylococcus aureus* isolates from keratitis and conjunctivitis. *Am. J. Ophthalmol.*, **137**(3): 453-458.
- Mesaros N, Nordmann P, Plesiat P, Roussel-Delvallez M, Van Eldere J, Glupczynski Y, Van Laethem Y, Jacobs F, Lebecque P, Malfroot A, Tulkens PM and Van Bambeke F (2007). *Pseudomonas aeruginosa*: Resistance and therapeutic options at the turn of the new millennium. *Clin. Microbiol. Infect.*, **13**(6): 560-578.
- Nasiri MI, Naqvi SB, Zaidi AA, Saeed R and Raza G (2013). Report - Comparative study on resistant pattern of clinical isolates against Levofloxacin and Cefepime. *Pak. J. Pharm. Sci.*, **26**(2): 415-419.
- Noviello S, Ianniello F, Leone S and Esposito S (2006). *In vitro* activity of prulifloxacin, levofloxacin and ciprofloxacin against urinary pathogens. *Infez. Med.*, **14**(1): 24-28.
- Prajapati BS, Prajapati RB and Patel PS (2008). Advances in management of urinary tract infections. *Indian J. Pediatr.*, **75**(8): 809-814.
- Reinert RR, Low DE, Rossi F, Zhang X, Wattal C and Dowzicky MJ (2007). Antimicrobial susceptibility among organisms from the Asia/Pacific Rim, Europe and Latin and North America collected as part of TEST and the *in vitro* activity of tigecycline. *J. Antimicrob. Chemother.*, **60**(5): 1018-1029.
- Shafiq Y, Tasleem S and Naqvi SBS (2012). Comparison of *in-vitro* antibacterial activity of levofloxacin and gatifloxacin. *Pak. J. Pharmacol.*, **27**(2): 21-2.
- Shakya P, Barrett P, Diwan V, Marothi Y, Shah H, Chhari N, Tamhankar AJ, Ashish Pathak A and Lundborg CS (2013). Antibiotic resistance among *Escherichia coli* isolates from stool samples of children aged 3 to 14 years from Ujjain, India. *BMC Infect. Dis.*, **13**: 477.
- Soriano A, Jurado A, Marco F, Almela M, Ortega M and Mensa J (2005). *In vitro* activity of linezolid, moxifloxacin, levofloxacin, clindamycin and rifampin, alone and in combination, against *Staphylococcus aureus* and *Staphylococcus epidermidis*. *Rev. ESP. Quimioter.*, **18**(2): 168-172.
- Stein GE and Goldstein EJC (2006). Fluoroquinolones and Anaerobes. *Clin. Infect. Dis.*, **42**(11): 1598-1607.
- Stevenson KB, Searle K, Stoddard GJ and Samore M (2005). Methicillin-resistant *Staphylococcus aureus* and vancomycin-resistant *Enterococci* in rural communities. *Emerg. Infect. Dis.*, **11**(6): 895-903.
- Tenover FC (2006). Mechanisms of antimicrobial resistance in bacteria. *Am. J. Med.*, **119**(6A): S3-10.
- Welte T and Pletz MW (2010). Antimicrobial treatment of nosocomial methicillin resistant *Staphylococcus aureus* (MRSA) pneumonia: Current and future options. *Int. J. Antimicrob. Ag.*, **36**(5): 391-400.
- Wiegand I, Hilpert K and Hancock RE (2008). Agar and broth dilution methods to determine the minimal inhibitory concentration (MIC) of antimicrobial substances. *Nat. Protoc.*, **3**(2): 163-175.
- Woo Hyuk Jang, Dong Hoon Yoo, Seong Woon Park (2011). Prevalence of and Risk Factors for Levofloxacin-Resistant *E. coli* isolated from outpatients with urinary tract infection. *Korean J. Urol.*, **52**: 554-559.
- Zemkova M, Kotlarova J, Merka V, Cermak P, Vlcek J and Jebavy L (2007). Emergence of fluoroquinolone resistance in *Escherichia coli* isolates at the Department of Clinical Hematology. *New Microbiologica*, **30**: 423-430.
- Zhanel GG and Noreddin AM (2001). Pharmacokinetics and pharmacodynamics of the new fluoroquinolones: Focus on respiratory infections. *Curr. Opin. Pharmacol.*, **1**(5): 459-463.
- Zhanel GG, Enis K and Vercaigne L (2002). A critical review of the fluoroquinolones: Focus on respiratory tract infections. *Drugs*, **62**: 13-59.