REVIEW

Rheumatoid arthritis: What have we learned about the causing factors?

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Abstract: Rheumatoid Arthritis (RA) is a common inflammatory autoimmune disease characterized by the synovitis of both small and large joints, which may lead to the destruction of cartilage and bones causing significant disabilities due to erosion of bones surfaces, if left untreated. It is a multifactorial and heterogeneous disease having contribution of both genetic (50-60%) and environmental factors. The unawareness of general public might be a contributing factor in the high prevalence rate of RA world-wide. This review article focuses on the causing factors (genetics and environmental) involved in this devastating disease. We also gave brief overview of the treatment options and animal models of RA. The literature was reviewed using mesh terms in PubMed search "etiology of RA, genetics of RA, environmental factors in RA, Genome Wide Association Studies (GWAS) in RA". The data was thoroughly reviewed and comprehensive information was extracted to help the readers in improving understanding towards the mechanisms, which trigger the outcomes of RA. The more we increase awareness about RA, the better we manage this disease and hence can improve life style and socio-economic status.

Keywords: Rheumatoid Arthritis, etiology, genetic factors, environmental factors.

INTRODUCTION

Rheumatoid Arthritis (RA) is a common, systemic and chronic inflammatory disease characterized hv inflammation of synovium of any joint including small joints of hands and feet and large joints of shoulder and knees. The synovitis of joints leads to the destruction of bones and cartilage resulting in the (radiographic) damages (Imboden, 2009). These damages can cause significant disability and even permanent loss of function, due to erosion of bone surface, if left untreated (Silman and Pearson, 2002; Majithia and Geraci, 2007). The etiology of RA is very complex and is yet to be explored properly. It has a wide spectrum of clinical manifestations, variability in disease severity, progression and differences in therapeutic response. These heterogeneous phenotypes of RA may suggest that variety of factors can contribute in the development of this complex trait, which includes environmental, hormonal and genetic factors.

The concordance rate of RA is about 3 to 4% in di-zygotic twins, 12 to 15% in monozygotic twins, 2 to 4% in non-twin siblings and is less than 1% in general population. Thus, RA has a strong genetic basis with estimated heritability ranging from 50% to 60% (Silman *et al.*,

1993; Seldin et al., 1999; Mac Gregor et al., 2000; Bax et al., 2011). In other words, siblings of the affected individuals are at high risk to RA than general population (Wandstrat and Wakeland, 2001). Since 2000, large number of studies have been conducted to understand the genetic susceptibility to RA among which Genome Wide Association Studies (GWAS) is considered to be a powerful tool to discover novel variants and loci especially in common complex diseases. Recent GWAS and meta-analysis of GWAS have reported more than 40 RA susceptibility loci/genes in different population. About 30% of genetic susceptibility of RA is contributed by Human Leukocyte Antigen (HLA) region while the non-HLA loci/gene account for about 5% susceptibility (Raychaudhuri et al., 2008; Kochi et al., 2010; Stahl et al., 2010; Craddock et al., 2010; deVries, 2011; Okada et al., 2012; Eyre et al., 2012; Jalil et al., 2013), suggesting the role of rare variants and gene-environment interaction in remaining heritability (Asimit and Zeggini 2011; Zuk et al., 2012).

Clinical features and ACR criteria of rheumatoid arthritis

Initially, based on the phenotypic characters and clinical presentations of the patients the American College of Rheumatology (ACR) developed an ACR 1987 criteria for the diagnosis of RA (Arnett *et al.*, 1988). These include morning stiffness (lasting for up to an hour),

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arthritis of three or more joints, arthritis of hand joints, rheumatic nodules, symmetrical arthritis, radiographic damages, degree of erosion and serum rheumatoid factor. A person would have RA if he or she satisfies at least four out of these criteria. These criteria are accepted worldwide and are used by both basic researcher for inclusion/exclusion of patients in the studies and by clinicians for diagnosis of the RA patients. The American College of Rheumatology Subcommittee on Rheumatoid Arthritis (ACR-SRA) recommends a baseline laboratory evaluations and clinical tests which include a complete blood cell, Rheumatoid Factor (RF), Anti-cyclic Citrullinated Peptide (ACCP) antibody, Erythrocyte Sedimentation Rate (ESR) or C-reactive Protein (CRP) and radiographic findings of involved joints (Ruddy et al., 2005). The onset of initial symptoms can be slow insidious (55-65% of cases) i.e. over weeks or months or an explosive sudden (8-15% of cases) which reaches to peak within few days (Fleming et al., 1976). The most commonly involved joints are the wrists, elbow, knee, metacarpophalangeal (MCP) joints, proximal interphalangeal (PIP) joints, metatarsophalangeal (MTP) joints and toe PIPs and cervical spine and the lumbosacral spine.

Later on, some limitations were noticed in 1987 ACR criteria. Actually, these classification criteria were developed from the patients with established and long-standing RA (Silman and Symmons, 1995). So, these criteria may not be helpful in early diagnosis and in-time treatment of the disease, which is necessary to avoid erosive bone damage and functional loss (van der Heide *et al.*, 1996). However, a major goal of the modern therapies is to prevent the erosive bone damages (Emery and Salmon, 1995), which becomes difficult once the bone is deformed.

Therefore, these flaws in 1987 ACR criteria for RA led to the development of new classification criteria. For this purpose a joint collaborative project was held between ACR and European League Against Rheumatism (EULAR) resulted in the development of new purely datadriven classification criteria for RA (Funovits et al., 2010). These 2010 ACR/EULAR criteria do consider some markers for inflammatory arthritis from 1987 ACR criteria like swelling and tenderness of small joints, serology and acute phase reactants. Similarly, 1987 ACR criteria excluded some traditional markers such as symmetry and morning stiffness to be less important during diagnosis phase. The major goal of this joint project was to make an early diagnosis possible and prevent erosion of the bones. It was decided that diagnosis should not be based on the sever outcomes of the definite disease and effective and timely therapy should be provided to all patients. Thus, typical features of established RA such as rheumatic nodules formation and erosion of the bone surface or deformities were excluded from the classification criteria during diagnosis.

The irreversible erosive damages of the bones in the chronic RA are due to the intra-articular inflammation. Although, RA is clinically very heterogeneous disease and severity of RA varies among the patients ranging from mild and self-limiting to an active and severely progressive disease (Lee and Weinblatt, 2001). However, level of inflammatory markers (C-reactive protein, CRP and Erythrocyte Sedimentation Rate, ESR) and serological markers (Rheumatoid Factor, RF and anti citrullinated protein antibodies, ACCP) are widely used both in diagnosis and as indicators of severity of RA (van Gaalen et al., 2004; Lindqvist et al., 2005) but still are not very specific to RA (Smolen et al., 2008). The genetic predisposition to the severity of joint destruction in RA patients was recently investigated and it was reported that 58% of variation in the progression rate of bone erosion is due to genetics of the patients (P=0.003) (Knevel et al., 2012). Thus differences in the genetic makeup of severe disease course than milder disease can predict about the patients with high risk for future bone damages. Several studies have tested correlation of the disease severity with candidate locus/allele. These studies explain the need for genetic predictors of the disease severity, which could be useful biomarkers in future (Marinou et al., 2010; Stahl and Raychaudhuri, 2012).

Epidemiology of rheumatoid arthritis

Rheumatoid arthritis is a multifactorial heterogeneous disease with different incidence rate and prevalence across different populations. The variations in epidemiology of RA can be due to exposure of different population to specific environmental triggers and can also be associated with the study design such as statistical methods used, case-ascertainment criteria, and number of cases enrolled etc.

The incidence of RA varies from population to population. Different studies have shown incidence of RA in certain populations like 9/100,000 in France; 25/100,000 in Norfolk, UK; 31/100,000 in Massachusetts, USA; 33/100,000 in Rochesher, Minnesota USA from 1985-1994; 34/100,000 in Finnish population; 35/100,000 from 1995-2001 in southern part of Denmark and 36/100,000 in Finland (Chan *et al.*, 1993; Guillemin *et al.*, 1994; Symmons *et al.*, 1994; Kaipiainen-Seppanen and Aho, 2000; Doran *et al.*, 2002; Savolainen *et al.*, 2003; Alamanos *et al.*, 2006; Pedersen *et al.*, 2009). It was summarized that incidence of RA (case per 100,000 population) as 29 (24-36) in Northern Europe, 16.5 (9-24) in Sothern Europe and 38 (31-45) in North America (Tobón *et al.*, 2010).

Like incidence, prevalence of RA also vary according to geographical area and population (Costenbader *et al.*, 2008) and is more prevalent in developed countries than developing ones. Several studies conducted on European and European derived populations have reported that prevalence of RA in North America and Northern Europe

is 0.5% to 1.1%; while in Southern Europe it is 0.3% to 0.7%. Furthermore, significantly lower prevalence of 0.1% to 0.5% has been reported in studies conducted in developing countries and 0.2% to 0.3% in Asian population (Saraux *et al.*, 1999; Guillemin *et al.*, 2001; Silman and Pearson, 2002; Symmons *et al.*, 2002; Carmona *et al.*, 2002; Teng *et al.*, 2011). The lower prevalence of RA in developing countries may be due to the fact that limited clinical diagnostic procedures ignore most of the patients from clinical assessment.

Studying the distribution of certain disease or diseases and the factors responsible in causing these conditions across multiple regions and populations of the world is called geo-epidemiology. Thegeo-epidemiologymight uncover population specific or ethnogenetic risk factors; like observing particular HLA types or other genes associated with RA in certain population. These observations can be compared with neighboring population and communities which can help in identifying the environmental triggers involved in pathogenesis of RA in particular region (Shapira *et al.*, 2010).

The trends in incidence and prevalence of RA has not been well investigated. However, some studies have suggested declines in both incidence and prevalence of RA after 1960 (Doranet al., 2002; Kaipiainen-Seppanen and Kautiainen, 2006). This hypothesis was justified by the observation that three factors may be involved in this decline. Firstly, it was suggested to be due to variations in methodologies and case-enrollment criteria. Several studies conducted before 1987 ACR criteria of RA can be biased; because of difficulties in differentiating RA from other polyarthritis. Secondly, ethnic and geographic factors are also equally important; like higher incidence of RA was reported in Pima Indians than other population of American and Europe. Finally, a true decline in incidence of RA has been noticed specifically in women using oral contraceptives (Doran et al., 2002; Savolainen et al., 2003; Doran et al., 2004).

Molecular genetics of rheumatoid arthritis

Rheumatoid arthritis is the most common multifactorial disease, which depends on the contribution of various factors including genetic, environmental and hormonal factors for the onset and development of clinical manifestation. The genetic heritability and familial susceptibility of RA can be evident from familial clustering. The higher concordance rate of RA in siblings than general population suggested a strong genetic basis with estimated heritability ranging from 50% to 60%. (Silman et al., 1993; MacGregor et al., 2000; Seldin et al., 1999; Bax et al., 2011). A wide range of genetic studies including candidate gene approaches, GWAS and their meta analysis have identified more than 35 genetic susceptibility loci/genes in different ethnic groups. We will discuss general function of some important genes below:

Human leukocyte antigen (HLA)

Major histo-compatibility complex (MHC)(HLA region) is responsible for about 30% (One-third) of the genetic susceptibility of RA with most important *HLA-DRB1* gene (MacGregor *et al.*, 2000) having DRB1*04:01 and DRB1*04:04major risk alleles in Caucasians and DRB1*04:05in East Asian populations (Newton *et al.*, 2004). The human MHC genomic region has been divided in to three main classes which are MHC I, II and III. MHC class I consists of three genes, *HLA-A*, *-B*, *-C* and MHC class II consists of *-DR*, *-DQ*, *-DP*. The Class I antigens such as HLA-A, *-B*, *-C* consist of a β 2-micro globulin and a highly polymorphic heavy chain. Similarly, Class II antigens (HLA-DR, *-DQ*, *-DP*) have an alpha chain and a highly polymorphic beta chains that is encoded by the *HLA-DRB1*, *-DQB1*, *-DPB1* genes.

HLA-DR antigen of MHC class II has shared epitope (SE) on beta chain (a five amino acids at positions 70-74). which has significant association with susceptibility and severity of RA (Gregersen et al., 1987; du Montcel et al., 2005; Gorman et al., 2004). These residues forma helical domain and may likely to influence antigen presentation by making an antigen binding site (Newton et al., 2004). The MHC class III is present between MHC class I and II. It has been determined that MHC Class III region also contains AIF1 and NFKBIL1, which are important RAsusceptible genes (Mu et al., 1999; Mattey et al., 1999; Ota et al., 2001; Ando et al., 2003; Tamiya et al., 2005; Lin et al., 2006; Yan et al., 2007; Harney et al., 2008; Yang et al., 2009). Six HLA loci which are in strong linkage disequilibrium (LD) can be ordered as HLA-A, -C, -B, -DRB1, -DQB1 and -DPB1 (from telomere to centromere) (Geraghty et al., 1999). A recent study on Asian population have shown that HLA-DRB1SE alleles (DRB1*04:05) have strong interaction with smoking and increasing the risk of RA in anti citrullinated protein antibodies (ACPA) positive individuals (Too et al., 2012).

Out of all these HLA loci only *HLA-DRB1* and the SE have been well explored with respect to RA. However, other HLA genes because of their highly polymorphic nature needed to be examined for their possible role in progression or protection of this devastating disease.

Peptidylarginine deiminase 4 (PADI4)

PADI gene is a family of gene present on chromosome 1(1p36). This gene coding for enzyme peptidylarginine deiminase 4 (PADI4), which converts arginine to citrulline with in peptides through posttranslational modification mechanism. Involvement of PADIs in the pathophysiology of RA was suggested after confirming that synovial fluid is the site for citrullination of auto antigenic peptides (Kinloch *et al.*, 2008). This idea was further supported by Chang *et al.*, (2009) through measuring the expression level of PADI4 in the synovial of RA patients. It was further investigated that autoantibodies to cyclic citrullinated peptides (ACCP) are

highly specific to RA (patients with ACCP have more swollen joints and radiological destructions as compare to those with no ACCP) and can predict about the onset of the disease even couple of years before the symptoms appear (Rantapaa-Dahlqvist et al., 2003; Vossenaar et al., 2003). Association of PADI4 with RA has been controversial and inconsistent between Asian and European population base studies, whereas some groups have shown positive association of this gene with RA while others have not. Likewise, studies conducted on Japanese population and Korean population from Asia have shown that PADI4 is involved in the outcome of RA, while not in Chinese Han population (Suzuki et al., 2003; Ikari et al., 2005; Kang et al., 2006; Chen et al., 2011). Similarly, North American population have reported positive association of this gene with RA (Plenge et al., 2005). Another large population-based study conducted on Caucasians for the first time reported strong association of PADI4 with RA (Eyre et al., 2011). Similarly, Stahl et al., 2010 showed modest effect of PADI4 through a GWAS-meta analysis in Europeans populations. However, a recent meta analysis of 27 studies suggested that PADI4 is a significant risk factor of RA in Asian Population than Europeans and Europeans derived populations (Hou et al., 2013).

TNF-receptor associated factor 1-complement component (TRAF1-C5)

The TRAF1-C5 consist of two important parts, one TNFreceptor associated factor 1 (TRAF1) and second complement component 5 (C5). These two are important immune system related genes, which are involved in perpetuation of inflammation. Through GWAS of 1522 RA cases and 1850 controls of European descent TNFR1-C5 was mapped on 9q33-34 as a novel genetic risk in ACCP positive RA patients (Plenge et al., 2007). Association of TNFR1-C5 gene with RA was replicated and validated by different studies in Caucasian populations (Kurreeman et al., 2007; Barton et al., 2008; Chang et al., 2008; Zervou et al., 2008; Kurreeman et al., 2008). Nishimoto et al., (2012), through a multicentercase control study reported positive association of TRAF1 in Japanese RA patients. These results from different groups confirmed the susceptibility of TRAF1-C5 to RA.

TRAF1 gene encodes intracellular protein, which mediates signal transduction. During this process TRAF1 binds to TNF receptors (mainly 1, 2) and CD40 and regulate cytokine signaling pathways including TNF α by binding several protein kinase and adaptor protein (Lee and Choi, 2007). In addition to its direct role in TNF α signaling *TRAF1* also mediate activation and proliferation of T cells (Sabbagh *et al.*, 2006). Through mouse model experiments it has been shown that *TRAF1* konockout mice had more T-cell proliferation and activation in response to TNF, suggesting the role of *TRAF1* as a negative regulator of this signaling pathway (Bradley and Pober, 2001). Tumor necrosis factor (TNF) is an

important cytokine with well established role in the pathophysiology and pathogenesis of RA (Firestein, 2003). For this reason. RA is treated with TNF antagonists from decades (Elliott et al., 1994; Weinblatt et al., 1999). The primary function of complement component to protect against microorganisms, while deregulated activity can play role in inflammation as well. Its pivotal role in the pathogenesis of RA has been recorded. Depletion of complement component from synovial fluid occur during inflammation in RA has been noticed, because cleavage of C5 generates pro-inflammatory anaphylatoxin C5a and C5b; generating membrane-attack complex later (Zvaifler, 1973; Cooke et al., 1975; Bao and Quigg, 2007). Furthermore, studies have shown that C5 deficient mice are also resistant to inflammatory arthritis and targeting C5 by antibodies prevent the onset of RA and reduce the clinical severity (Wang et al., 1995; Wang et al., 2000; Ji et al., 2002), which also explains the role of C5 in RA onset and pathogenesis. These observations reveal that two important immune mediator genesC5 and TRAF1 found adjacent to each other on chromosome 9 have combine affects in pathogenesis of RA.

Protein tyrosin phosphatase Non-recptor 22 (PTPN22)

Protein tyrosin phosphatase non-recptor 22 (PTPN22) is one of the strongest risk factors of autoimmunity outside major histo-compatibility complex (MHC), located on chromosome 1p13.3-13.1 and is ranked second in term of single-gene contribution to the etiology of RA in Caucasian population (Todd et al., 2007; Fiorillo et al., 2010). PTPN22 encodes lymphoid tyrosin phosphatase (Lyp), which form a complex with C-terminal Src tyrosine kinase (Lyp-Csk) and acts as a negative regulator of T-cell receptor (TCR) signaling (Gjörloff-Wingren et al., 1999; Clouteir and Veillete, 1999; Hill et al., 2002; Begovich et al., 2004; Vang et al., 2005; Todd et al., 2007). A well established non-synonymous C1858T single nucleotide polymorphism (rs2476601) which results in Arg620Trp (R620W) has been reported in number of autoimmune diseases including RA, among many populations especially in ACCP positive RA (Begovichet al., 2004; Hasegawa et al., 2004; Viken et al., 2005; Gregersen, 2005; Rieck et al., 2007). There are two types of protein tyrosin phophatase; receptor (membrane bounded-RPTP) and non-receptor (cytoplasmic-NRPTP). The Lyp is a ~105-kDa protein with ~300 amino acid Nterminal domain and ~200 amino acid C-terminal domain, includes four putative polyproline motifs from P1-P4. The C and N-terminus are separated by ~300 amino acid domain called interdomain. Interaction between proteintyrosin phosphatase (PTP) Lyp and protein-tyrosin kinase (PTK) Csk is mediated by P1 motif of N-terminal domain (Fiorillo et al., 2010). The Lyp-Csk complex can inhibit T-cell receptor signaling only when they are physically associated with each other (Cloutier and Veillette, 1999). The Lyp physically bound through SH3 domain to Csk (Cohen et al., 1999). The single amino acid change

R620W disrupt this complex formation and hence causes suppression of T-cell activation. It has been shown that Tallele of PTPN22 bind less efficiently to Csk then Callele; making cell with T-allele hyper-responsive and hence individuals with this allele show autoimmunity (Bottini et al., 2004; Begovich et al., 2004). Gregersen, (2005) presented a simple scheme of autoimmunity through R620W resulting in gain of enzymatic function as shown in fig.1. This fig. shows that two mechanisms, either one or both are involved in this reaction. According to first mechanism the shift in signaling threshold could cause positive selection of thymocytes (that normally deleted) and potentially auto reactive T-cells appears in the periphery. While, in second mechanism this shift could cause deficiency in the regulation of auto reactive T-cell, making individual susceptible to autoimmunity.

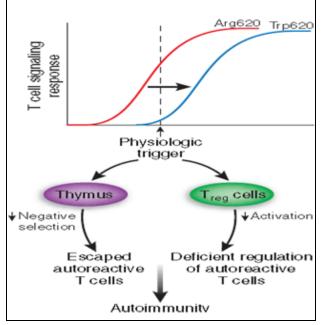


Fig. 1: A general scheme of T-cell signaling and autoimmunity (by Gregersen, 2005).

Signal transducers and activators of transcription 4 (STAT4)

A whole genome-wide SNP linkage scan of 642 Caucasian RA families with Illumina IV SNPs linkage panel containing 5850 SNP markers across the genome, established strong linkage of RA at 2q33 (LOD score 3.52) (Amos *et al.*, 2006). Same group of researcher conducted case-control study on both RA and SLE using candidate genes at this region. They tested SNPs in and around 13 candidate genes within 2q33 region and found association of a SNP rs7574865 at *STAT4*gene with both RA (P=2.82E-07; OR=1.32) and SLE (P=1.87E-09; OR=1.55) (Remmers *et al.*, 2007). Association of *STAT4*gene with RA has been confirmed in different populations after it was first reported with significant role in RA along with the evidence that antibody status (RF positive or negative; ACCP positive or negative) does not

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affect susceptibility to RAby STAT4/rs7574865 (Lee et al., 2007; Martinez et al., 2008; Zervou et al., 2008; Orozco et al., 2008; Kobayashi et al., 2008; Barton et al., 2008; Palomino-Morales et al., 2008). A meta-analysis on T-allele (susceptible allele) of rs7574865 using 15 studies (10 Europeans; 4 Asian; 1 Latin American) containing 16,066 RA patients and 16,509 controls subjects revealed association of RA and STAT4 (Over all OR=1.271, 95% CI=1.197-1.350, P<0.001). Furthermore, STAT4 was found significantly associated with RA in both Europeans (OR=1.300, 95% CI=1.195-1.414, P<0.001) and Asian (OR=1.216, 95% CI=1.135-1.303, P<0.001) (Lee et al., 2010). Another recent meta-analysis of 40 studies (published before September 2011), confirmed the association of STAT4 (rs7574865; T-allele) with multiple autoimmune diseases including RA and systemic lupus erythematosus (SLE) (Liang et al., 2012).

Signal transducers and activators of transcription (STATs) is a family of proteins including STAT1, STAT2, STAT3, STAT4, STAT5a, STAT5b and STAT6. Different cytokines, growth factors and hormones are involved in activation of STATs proteins. During activation process these cytokines, growth factors and hormones binds to STATs receptors and phosphory late STAT proteins on either tyrosine or serine residues (Leonard and O'Shea, 1998; Visconti *et al.*, 2000).

STAT4 is one of the important members of this family of proteins expressed in peripheral blood monocytes, machrophages and dendritic cells at site of inflammation (Frucht et al., 2000) and is involved in regulation of hematopoitic process STAT4 is a latent cytosolic factor which is first phosphorylated and then accumulated in the nucleus after activation by cytokines. SATA4 is also highly expressed in synovium of RA patients as compare to normal tissue (Walker et al., 2006; Walker et al., 2007). STAT4 encodes a transcription factor which transmit signals induced by several key cytokines, including IL-12 and type 1 interferons and IL-23 and also play a crucial role in differentiation and proliferation of helper T-cells (Th1 and Th17) (Murphy and Reiner, 2002; Watford et al., 2004; Mathur et al., 2007). STAT4-dependent signaling byIL-12 receptors results in differentiation of CD4+ T-cells in to interferon- γ producing Th1 cells linage and plays a critical role in the development of Th1-type Tresponse. After activationSTAT4 stimulates cell transcription of interferon- γ , which is akey indicator of Tcell differentiation into type 1 helper T (Th1) cells. While, signaling by IL-23 receptors helps in developments of IL-17- secreting helper T-cell (Th17), which play critical role in autoimmune diseases such as RA (Morinobu et al., 2002; Nishikomori et al., 2002; Watford et al., 2004; Skapenko et al., 2005; Bettell et al., 2007; Steinman, 2007). Furthermore, animal model studies have provided many evidences about the role of STAT4 in pathogenesis of RA and hence suggested it as a possible therapeutic target.

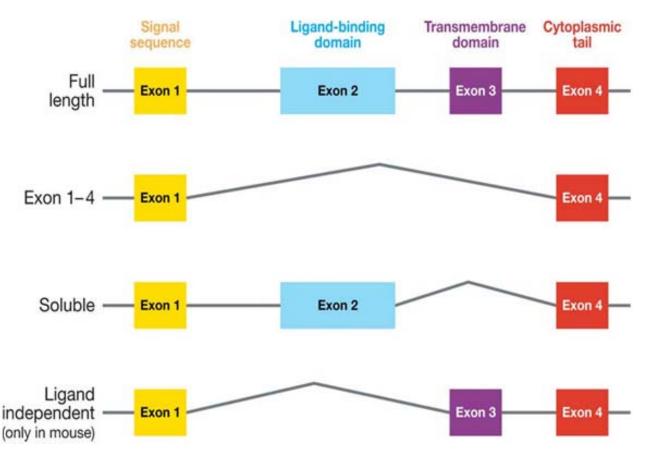


Fig. 2: The CTLA4 gene splice variants in human and mouse (adapted from Wendy et al., 2006)

Kinesin family member 5A (KIF5A)

KIF5A gene encodes a member of kinesin family of protein called kinesin heavy chain is form 5A. This gene was first mapped to 12q13 in Hereditary Spastic Paraplegia (HSP); which is group of inherited disease with progressive stiffness and contraction in the lower limbs because of the dysfunction of the nerves (Hamlin et al., 1998; Depienne et al., 2007). KIF5A is a motor protein, which facilitates intracellular movement of organelles and microtubules (Wang et al., 2007). Some other complications like contracts, ataxia, epilepsy, peripheral neuropathy, and deafness have also been noticed in HSP patients. Studies have shown that HSP patients have defective transport system including protein transport and transportation of other substance in the cell. Recently, it was found that long nerves which transport materials through long distance are the main targets in HSP (DeMatteis and Luini, 2011). Recent genome wide association studies and their meta analysis have confirmed the association of KIF5A gene to RA (Plant et al., 2010; Stahl et al., 2010; Bowes et al., 2012).

Cytotoxic T-lymphocyte antigen-4 (CTLA4)

CTLA4,a 6173 bp (6.17kb) long gene at 2q13, downregulate T-cell activation and hence protects from T-cell autoimmunity. It encodes a transmembrane 223 amino

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acid long glycoprotein belongs to immunoglobulin superfamily, having 35 amino acid signal peptides. The extra cellular part of this protein molecule is encoded by exon 1 and 2, containing the B7-1(CD80) and B7-2 (CD 86) ligand binding sites on exon 2 and leader sequences on exon 1 (Brunet et al., 1987; Dariavach et al., 1988; Lindsten et al., 1993; Linsley et al., 1995; Metzler et al., 1997; Ling et al., 1999;Ostrovet al., 2000). The transmembrane region of CTLA4 molecule is encoded by exon 3 and the 36 amino acid cytoplasmic portion (lacking any enzymatic activity) is encoded by exon 4 (Dariavach et al., 1988; Ling et al., 1999; Baroja et al., 2000). It contains proline rich region at position 169, lysin rich motifs and two tyrosine residues at position 165 and 182, which have shown to be involved in modulating its function by variety of signaling molecules (Baroja et al., 2000; Baroja et al., 2002; Lee et al., 1998; Schneider et al., 1995; Shiratori et al., 1997). Splicing variants identified in human and mice are shown in fig. 2. Human have one full-length mRNA having all four exons, a second transcript without exon 3 and is called soluble CTLA4 (sCTLA4), and a third transcript containing only exon 1 and exon 4 (Brunet et al., 1987; Dariavach et al., 1988: Ling et al., 1999:Ueda et al., 2003:Vijavakrishnan et al., 2004; Magistrelli et al., 1999; Oaks et al., 2000); while mouse have an additional splicing variant which does not contain exon 3 and 4 and is called ligand-independent *CTLA4* (liCTLA4) (Ueda *et al.*, 2003).

Since, the co-stimulatory molecule *CD28* have structural similarities and sequence homology with CTLA4; thus compete for the same ligand (B7-1 and B7-2) (Linsley *et al.*, 1990; Freeman *et al.*, 1993a; 1993b). However, *CTLA4* have greater affinity for ligand then *CD28* (Linsley *et al.*, 1991; Linsley *et al.*, 1994).

Although, the CTLA4 gene is primarily expressed in Tcells; however, expression in other cells like $CD4^+$, CD25, and regulating T-cells have also been determined (Harper et al., 1991; Perkins et al., 1996; Takahashi et al., 2004). Variety of other cells like B-cells, monocytes, granulocytes, CD34+, placental fibroblast and mouse embryonic cell do express this gene for unexplained regulatory function (Pioli et al., 2000; Pistillo et al., 2003; Wang et al., 2002). Several authors have demonstrated the association of CTLA4 gene with number of autoimmune diseases. Some single nucleotide polymorphisms (SNP) like -1722T/C, -1661A/G, -318C/T and +49A/G are well studied variants. The initial three variants (-1722T/C, -1661A/G, -318C/T) are found in regulatory/promoter region and are thought to be associated with higher promoter activity and hence increase CTLA4 expression (Wang et al., 2002). The transition at +49A/G causes threonine to alanine substitution in the leader peptide of exon 1 (Nistico et al., 1996; Deichmann et al., 1996; Donner et al., 1997; Kristiansen et al., 2000; Wang et al., 2002) which may affects the inhibitory function of CTLA4 and may also influence the endocytosis (Wang et al., 2001). A recent ethnicity-specific meta-analysis was performed on Caucasian and Asian populations, including 5,752 RA patients and 5,508 controls from 19 studies (9 Caucasian, 8 Asian, 1 Mexican, and 1 Tunisian population) demonstrated that +49A/G polymorphism confer susceptibility to RA in Asian population but not in Caucasians (Lee et al., 2012).

Other non-HLA RA susceptibility genes/loci include Solute Carrier Family 22A4, Complement component 5-TNF receptor-associated factor 1(C5-TRAF1), Macrophage migration inhibitory factor (*MIF*), Runt-Related Transcription Factor (*RUNX1*), Tumor Necrosis Factor Alpha Receptor 2 (*TNFR2*), Cluster of Differentiation 244 (*CD244*), Corticotropin-releasing hormone (*CRH*) and Angiotensin-Converting Enzyme (*ACE*). These genes either involved in T cell proliferation or cytokines regulatory pathways and hence add to autoreactivity and autoimmunity in the human body.

Environmental risk factors of RA

The differences in prevalence of RA across different regions and populations of the world have focused the scientists on environmental factors and gene-environment interactions in addition to genetic factors in pathogenesis of RA. Several important environmental determinants involved in the development and severity of RA have been extensively studied. We are discussing them in brief below.

Smoking

In addition to association of cigarette smoking with many diseases like several malignancies, cardiovascular diseases and pulmonary diseases; smoking is also suggested to be the strongest environmental risk factor associated with the development of RA. Vessey et al., (1987) reported for the first time that smoking is an important risk factors in RA pathogenesis. After this initial report interaction of smoking and RA was studied and replicated in various populations and higher risk of development of RA was found in heavy smokers as compare to non-smokers or who smoked less (Karlson et al., 1993; Heliovaara et al., 1993; Symmons et al., 1997; Uhlig et al., 1999; Criswell et al., 2002; Padyukov et al., 2004; Karlson et al., 2010). A large study conducted on Caucasian women smokers, the Iowa Women's Health Study (IWHS) reported that risk of RA was 18% which mean one in six of new RA cases can be due to smoking and can be prevented if smoking is eliminated (Criswell et al., 2002). Studies have also shown that risk of RA further increases with increasing duration and amount of cigarette taken (Stoltet al., 2003). A similar linear relation between smoking and risk of RA has been observed in another large prospective, Nurse's Health Study (NHS). According to this study heaviest smokers with more than 40 packsyears have two-fold higher risk of RA as compare to nonsmoker controls (Costenbader et al., 2006).

Furthermore, interaction of smoking and genetic factors was examined and their co-relation with RA risk was observed. It was suggested that *HLA-DRB1*-shared epitope (SE) is strongly associated with increased risk due to smoking and is more evident in seropositive RA. Smokers having two copies of *HLA-DRB1*-shared epitope (SE) have higher risk of RA than those who never smoke and having no SE allele (Klareskog *et al.*, 2006; Karlson *et al.*, 2010). A recent study of Bang *et al.*, (2010) reported that SE-alleles and smoking are associated with both anti-CCP positive and anti-CCP negative RA. Smokers with two copies of the SE allele have higher risk of both ACCP-positive and ACCP-negative RA, 36.11-fold and12.29-fold, respectively, as compared to nonsmokers not carrying SE alleles.

Klareskog *et al.* (2006) obtained bronchoalveolar lavage specimens from both smokers and nonsmokers and reported citrullinated proteins in smokers but not non-smokers. Later, it was found that smoking upregulate expression of peptidylarginine deiminase (PAD) in the lungs and was concluded that long-term smoking and possibly with interaction of other environmental triggers may convert arginine of peptide antigens to citrulline in the lungs (Makrygiannakis *et al.*, 2008).

In addition to cigarette smoking, the role of other environmental pollutants in risk of RA have been explored. The "distance-to-road" was examined in a prospective study of NHS cohort. It was noted that women living less than 50 meters from road had 30% increased risk of RA, than those residing at a distance from the road (Hart *et al.*, 2009). Similarly, exposure to silica and silica dust from stone works, mining, glass or ceramics manufacturing, stone drilling and rock crushing might increase RA risk. In a Swedish case-control study of silica exposed individuals had ACPA-positive RA (OR, 1.7; 95% CI, 1.1-2.5) as compared to unexposed individuals. However, no ACPA-negative RA has been reported in silica-exposed individuals (Stolt *et al.*, 2010).

Alcohol

The association of alcohol consumption and the risk of RA has been studied and protective effect of moderate alcohol intake on development of RA has been suggested. An inverse association between consumption of alcohol and risk of rheumatoid arthritis has been observed (Hazes et al., 1990; Maxwell et al., 2010). A Danish study reported lower risk of developing ACPA-positive RA in those who consume alcohol (Pedersen et al., 2006). Two independent case-control populations; a Danish CACORA (case-control study on Rheumatoid Arthritis) and a Swedish EIRA (epidemiological investigation of rheumatoid arthritis) demonstrated dose-dependent effect of alcohol and reduction of RA risk. They found higher rate of alcohol consumption in control individuals versus patients. Individuals with highest alcohol consumption (\geq drinks or 80g ethanol per week) was found to have 40% to 50% decreased risk of RA than those with lower to no consumption (<0.5g ethanol per week) (Kallberg et al., 2009). A recent study conducted by Di-Giuseppe et al., (2012) observed 37% decrease in risk of RA among heavy drinker women (>4 glasses of alcohol (1 glass = 15g of ethanol) per week compared with women who drank <15g per week or who never drank alcohol (RR, 0.63; 95% CI, 0.42 to 0.96; P=0.04). These observations suggested that moderate to low level consumption of alcohol is associated with reduced risk of RA. Further investigations on the biological mechanisms and pathways have shown that alcohol down regulate immune response inanimals and humans (Mandrekar et al., 2004; Verma et al., 2008; Fan et al., 2011) and decrease the production of proinflammatory cytokines (Waldschmidt et al., 2006). Furthermore, ethanol candelays the onset and may stops the progression of RA in mice by interacting with innate immunity (Jonsson et al., 2007).

Dietary factors

Dietary factors play a vital role in the onset and development of inflammatory processes. In examining the relationship between diet and RA, researchers have found some important factors like antioxidants, micronutrients and some vitamins and proteins. Antioxidant are present in serum where it reduces inflammatory products and cease inflammation. Antioxidant have important protective role against oxygen species, which can cause tissues damage. Beside this antioxidant suppresses the expression of certain cytokines and collagenase induced by TNF- α (Sato *et al.*, 1996; Li and Micheletti, 2011). Lower concentration of serum-circulation antioxidant including vitamin C, vitamin E, β -carotene and zinc have been shown in RA patients when were compared with normal control individuals (Aaseth *et al.*, 1998; Li and Micheletti, 2011).

The Mediterranean diet of southern European part (having lower prevalence and incidence of RA) which is rich in fruits, vegetables, cereals, beans, nuts, seeds, fish, olive oil and low in red meat suggests the importance of plant foods (Rayman and Callaghan, 2006). Two controlled Mediterranean-diet intervention trials were conducted on RA patients and suggested significant results in improvement of morning stiffness and reducing pain (Skoldstam *et al.*, 2003; McKellar *et al.*, 2007). Increased red meat and protein in diet have been associated with increased risk of RA (Pattison *et al.*, 2004). The vegetarian diet rich in fruits and vegetables and lower fats altering amount of antioxidants, arachidonic acid and fatty acid and hence reducing inflammation in response (Adam *et al.*, 2003; Smedslund *et al.*, 2010).

Vitamin D is an important hormone in development of bones and also exerts anti-inflammatory properties by regulating cells in innate and adaptive immune system through vitamin D receptors (VDR) (Mathieu *et al.*, 2001). Insufficiency of vitamin D has been observed in RA patients (Kerr *et al.*, 2010) and furthermore vitamin D can increase disease severity in patients with polyarticular inflammatory arthritis (Patel *et al.*, 2007).

Socio-economic status

The social class and socio-economic position are collectively called socio-economic status (SES). The term SES has prominent impacts in many medical conditions. Low SES is associated with higher psychiatric diseases, depression and higher mortality rate (Lorant et al., 2003; Stringhini et al., 2010). Low SES is more prone to stress exposure and weaker social support. Different criteria are used as measure of SES like occupation is used in Europe (Stansfeld et al., 1998; Mackenbach et al., 1997; Stringhini et al., 2010), education and income is used in USA (Mitchell et al., 1988; Hawley and Wolfe, 1988; Criswell and Katz, 1994). Although, a single measure is insufficient to give complete picture of SES of certain population; however still give valuable data for understanding. Researchers have linked SES with depression and RA (Fitzpatrick et al., 1991; Berkanovic et al., 1996). Bengtsson et al. (2005) also reported inverse association between SES (measured in terms of education and occupation class) and risk of RA. A recent report published by US national survey suggested that SES including low education and low income are associated with poor mental health and arthritis (Furner *et al.*, 2011).

Other environmental risk factors

In addition to the above-mentioned factors, development and progression of RA has been associated with many other factors with minor or major affect. These miscellaneous factors include high birth weight, breast feeding, oral contraceptive, sex hormonal factors, infectious agents (including bacteria, viruses, mycoplasma), complex gene-environment interactions, gender and age and other pollutants. These factors have either independently associated with RA or show combined affects by activation of other triggers, enhance risk of RA.

Treatment options of rheumatoid arthritis

Currently, variety of treatment options and drugs are available for RA which are used to reduce inflammation of affected joints and hence the disease activity can be monitored if properly administered. These drugs are prescribed as a combination therapy according to the patient age, activity and course of the disease, and response of the patient to the drug.

Treatment of RA patient is started with analgesics like acetaminophen and aspirin to reduce inflammation and pain of the affected areas and to give instant relief to the patients (Cuzzocrea, 2011). Another group of drugs called nonsteroidal anti-inflammatory drugs (NSAIDs) is also used an effective therapy for RA. The major NSAIDs currently in use are nabumetone, diclofenac, ibuprofen, piroxicam, naproxen, oxaprozin, phenylbutazone, sulindac. tolmetin.diflunisal, etodolac. fenoprofen, flurbiprofen, indomethacin, ketoprofen, meclofenamate, mefenamic acid and meloxicam. These drugs can reduce inflammation and pain but do nothing with the course and progression of disease (McCabe et al., 1998; Cuzzocrea, 2011). The progression of disease and damage of the affected joints can be reduced by treating patients with another group of drugs called disease-modifying antirheumatic drugs (DMARDs which include methotrexate (MTX), sulfasalazine (SSZ), leflunomide, Auranofin (oral gold), hydroxychloroquine (HCQ) and gold salts (injectable). These drugs beside reducing progression also decrease pain and swelling in the affected joints, thus should be taken in the earlier stages of the disease rather than later (Arndt et al., 2003). Sever inflammation and activity of life threatening disease is also suppressed with corticosteroids/glycocorticosteroids including methylprednisolone, prednisone and inject able corticosteroids. Steroids are mostly prescribed in combination with DMARDs specially MTX and is considered to be the first-line treatment if given in suitable dose (Tłustochowicz, 2006).

A new and effective class of drugs called biologics have significantly improved treatment of RA. Some important frequently used biologics are anti-TNF compounds (adalimumab, etanercept, infliximab, certolizumab, golimumab can suppress inflammation and hence damage of affected joints), IL-1 inhibitor (anakinra is used in cases who do not respond to DMARDs), B-cell-depleting agent (aituximab is given to patients who do not respond to TNF inhibitors), T-cell co-stimulation antagonist (abatacept) and IL-6 antagonist (tocilizumab). The patients who do not respond to DMARDs or have persistent and progressive disease course are treated with a standard combination therapy of biologics and MTX (Lai and Chen, 2008; Cuzzocrea, 2011).

Animal models and rheumatoid arthritis

Large number of rat and mouse models which mimic different characteristics of RA are available. These experimental animal models can be used for evaluation and understanding the pathogenesis and molecular mechanisms implicating in the RA patients. These models are also used for testing new therapeutic options and drugs before going in human trials.

Collagen-induced arthritis (CIA) is a commonly used experimental mouse model for studying pathological mechanisms and for therapeutic testing of newly developanti-inflammatory drugs against RA. In this method college type II (CII), a collagen in the cartilage, is used to induce CIA. A 200 μ l emulsion of CII and Freund's adjuvant is injected intradermally and followed by a booster dose of 100 μ lon the other side of the tail. Development of CIA is started in susceptible strains (H-2q or H-2r) within two to three weeks after booster dose (Trentham *et al.*, 1977; Courtenay *et al.*, 1980; Wooley *et al.*, 1981; Jirholt *et al.*, 2001; Zhang *et al.*, 2008; Seeuws *et al.*, 2010).

Proteoglycan-induced arthritis (PGIA) is induced by proteoglycans isolated form cartilage of osteoarthritis patients. The mice are immunized with 100μ g of proteoglycan. Emulsion of PG and an adjuvant (dimethyldioctadecylammonium bromide) is prepared in phosphate-buffered saline (pH 7.4). The severity of disease can be determined from swelling and redness in the paws both front and hind, in susceptible strains (C57BL/6J, BALB/c). The female mice of susceptible strains are more prone to develop RA than males (Glant *et al.*, 1987; Glant *et al.*, 2001; Glant *et al.*, 2011).

CIA and PGIA are the most commonly used experimental models. Studies have identified antibodies to both CII and PG, which confirm them as the most relevant animal models of human RA. Furthermore, CIA and PGIA depend on B and T cells and are associated with large number of MHC and non-MHC genes/loci making them polygenic-disease models (Glant *et al.*, 1980; Cook *et al.*,

1994; Svensson et al., 1998; Corthay et al., 1999; Adarichev et al., 2003; O'Neill et al., 2005).

Large number of different other experimental animal models of RA are currently available which include Avridine-induced arthritis (AIA), Oil-induced arthritis (OIA), Streptococcal cell wall-induced arthritis, Genetically manipulated mouse strains, Pristane-induced arthritis (PIA) and Adjuvant-induced arthritis (AIA) (Cuzzocrea, 2011; Villa-Forte and Mandell, 2011; Adipue *et al.*, 2011; Hu *et al.*, 2013).

CONCLUSION

This literature survey presents up to-date etiological findings related to RA, which can be helpful in disease cure and management. It would also be beneficial for clinicians, paramedics, RA patients and general public. Increase in public health awareness about RA etiology would ultimately decrease the prevalence rate of RA and thus may improve socio-economic status.

REFERENCES

- Aaseth J, Haugen M and Forre O (1998). Rheumatoid arthritis and metal compounds-perspectives on the role of oxygen radical detoxification. *Analyst.*, **123**(1): 3-6.
- Adam O, Beringer C and Kless T *et al* (2003). Antiinflammatory effects of a low arachidonic acid diet and fish oil in patients with rheumatoid arthritis. *Rheumatol. Int.*, **23**(1): 27-36.
- Alamanos Y, Voulgari PV and Drosos AA (2006). Incidence and prevalence of rheumatoid arthritis, based on the 1987 American College of Rheumatology criteria: a systematic review. *Semin. Arthritis Rheum*, **36**: 182-188.
- Amos CI, Chen WV, Lee A, Li W, Kern M, Lundsten R, Batliwalla F, Wener M, Remmers E, Kastner DA, Criswell LA, Seldin MF and Gregersen PK (2006).
 High-density SNP analysis of 642 Caucasian families with rheumatoid arthritis identifies two new linkage regions on 11p12 and 2q33. *Genes Immun.*, 7: 277-286.
- Arndt U, Rittmeister M and Möller B (2003). Drug therapy of rheumatoid arthritis. Orthopade., 32(12): 1095-1103.
- Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS and Healey LA *et al* (1988). The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum*, **31**: 315-324.
- Asimit J and Zeggini E (2011). Testing for rare variant associations in complex diseases. *Genome. Med.*, **3**: 24-26.
- Bang SY, Lee KH, Cho SK, Lee HS, Lee KW and Bae SC (2010). Smoking increases rheumatoid arthritis susceptibility in individuals carrying the HLA-DRB1 shared epitope, regardless of rheumatoid factor or anti-

cyclic citrullinated peptide antibody status. *Arthritis Rheum*, **62**(2): 369-377.

- Baroja ML, Darlington PJ, Carreno BM and Madrenas J (2000). Inhibition of T cell activation by CTLA-4: truths and red herrings. *Mod. Asp. Immunobiol.*, **1**: 169-173.
- Baroja ML, Vijayakrishnan L, Bettelli E, Darlington PJ and Chau TA *et al* (2002). Inhibition of CTLA-4 function by the regulatory subunit of serine/threonine phosphatase 2A. *J. Immunol.*, **168**: 5070-5078.
- Barton A, Thomson W, Ke X, Eyre S, Hinks A, Bowes J and Gibbons L *et al* (2008). Re-evaluation of putative rheumatoid arthritis susceptibility genes in the postgenome-wide association study era and hypothesis of a key pathway underlying susceptibility. *Hum. Mol. Genet*, **17**: 2274-2279.
- Bax M, Heemst JV, Huizinga TWJ and Toes REM (2011). Genetics of rheumatoid arthritis: What have we learned. *Immunogenetics*, **63**: 459-466.
- Begovich AB, Carlton VE, Honigberg LA, Schrodi SJ, Chokkalingam AP and Alexander HC *et al* (2004). A missense single-nucleotide polymorphism in a gene encoding a protein tyrosine phosphatase (PTPN22) is associated with rheumatoid arthritis. *Am. J. Hum. Genet*, **75**(2): 330-337.
- Bengtsson C, Nordmark B, Klareskog L, Lundberg I and Alfredsson L (2005). Socio-economic status and the risk of developing rheumatoid arthritis: Results from the Swedish EIRA study. *Ann. Rheum. Dis.*, **64**: 1588-1594.
- Berkanovic E, Oster P, Wong WK, Bulpitt K, Clements P, Sterz M and Paulus H (1996). The relationship between socioeconomic status and recently diagnosed rheumatoid arthritis. *Arthritis Care. Res.*, **9**(6): 257-262.
- Bettelli E, Oukka M and Kuchroo VK (2007) T (H)-17 cells in the circle of immunity and autoimmunity. *Nat. Immunol.*, **8**: 345-350.
- Bottini N, Musumeci L and Alonso A *et al* (2004). A functional variant of lymphoid tyrosine phosphatase is associated with type I diabetes. *Nat. Genet*, **36**: 337-338.
- Bowes J, Ho P, Flynn E, Ali F, Marzo-Ortega H, Coates LC and Warren RB *et al* (2012). Comprehensive assessment of rheumatoid arthritis susceptibility loci in a large psoriatic arthritiscohort. *Ann. Rheum. Dis.*, **71**(8): 1350-1354.
- Brunet JF, Denizot F, Luciani MF, Roux-Dosseto M and Suzan M *et al* (1987). A new member of the immunoglobulin superfamily-CTLA-4. *Nature*, **328**: 267-270.
- Carmona L, Villaverde V, Hernandez-Garcia C, Ballina J, Gabriel R and Laffon A (2002). The prevalence of rheumatoid arthritis in the general population of Spain. *Rheumatology (Oxford).*, **41**: 88-95.

- Chan KW, Felson DT, Yood RA and Walker AM (1993). Incidence of rheumatoid arthritis in central Massachusetts. *Arthritis Rheum*, **36**: 1691-1696.
- Cloutier JF and Veillette A (1999). Cooperative inhibition of T-cell antigen receptor signaling by a complex between a kinase and a phosphatase. *J. Exp. Med.*, **189**: 111-121.
- Cloutier JF and Veillette A (1999). Cooperative inhibition of T-cell antigen receptor signaling by a complex between a kinase and a phosphatase. J. Exp. Med., **189**(1): 111-121.
- Cohen S, Dadi H, Shaoul E, Sharfe N and Roifman CM (1999). Cloning and characterization of a lymphoid-specific, inducible human protein tyrosine phosphatase. *Lyp. Blood*, **93**: 2013-2024.
- Costenbader KH, Chang SC, Laden F, Puett R and Karlson EW (2008). Geographic variation in rheumatoid arthritis incidence among women in the United States. *Arch. Intern. Med.*, **168**: 1664-1670.
- Costenbader KH, Feskanich D, Mandl LA and Karlson EW (2006). Smoking intensity, duration and cessation, and the risk of rheumatoid arthritis in women. *Am. J. Med.*, **119**: 5031-5039.
- Craddock N, Hurles ME, Cardin N, Pearson RD, Plagnol V and Robson S *et al* (2010). Wellcome trust case control consortium, Genome-wide association study of CNVs in16,000 cases of eight common diseases and 3,000 shared controls. *Nature*, **464**: 713-720.
- Criswell LA and Katz PP (1994). Relationship of education level to treatment received for rheumatoid arthritis. *J. Rheumatol.*, **21**(11): 2026-2033.
- Criswell LA, Merlino LA, Cerhan JR, Mikuls TR, Mudano AS and Burma M *et al* (2002). Cigarette smoking and the risk of rheumatoid arthritis among postmenopausal women: Results from the Iowa Women's Health Study. *Am. J. Med.*, **15**: 465-471.
- Cuzzocrea S (2011). Characterization of a novel and spontaneous mouse model of infl ammatory arthritis. *Arthritis Research & Therapy*, **13**: 126.
- Dariavach P, Mattei MG, Golstein P and Lefranc MP (1988). Human Ig superfamily CTLA-4 gene: Chromosomal localization and identity of protein sequence between murine and human CTLA-4 cytoplasmic domains. *Eur. J. Immunol.*, **18**: 1901-1905.
- De Matteis MA and Luini A (2011). Mendelian disorders of membrane trafficking. *New England Journal of Medicine*, **365**(10): 927-938.
- Deichmann K, Heinzmann A, Bruggenolte E, Forster J and Kuehr J (1996). An Mse I RFLP in the human CTLA4 promotor. *Biochem. Biophys. Res. Commun.*, 225: 817-818.
- Depienne C, Stevanin G, Brice A and Durr A (2007). Hereditary Spastic Paraplegia: An Update. *Current Opinions in Neurology*, **20**(6): 674-680.
- deVries R (2011). Genetics of rheumatoid arthritis: Time for a change. *Curr. Opin. Rheumatol.*, **23**(3): 227-232.

- Di-Giuseppe D, Alfredsson L, Bottai M, Askling J and Wolk A (2012). Long term alcohol intake and risk of rheumatoid arthritis in women: A population based cohort study. *BMJ.*, **345**: e4230.
- Donner H, Rau H, Walfish PG, Braun J and Siegmund T et al (1997). CTLA4 alanine-17 confers genetic susceptibility to Graves' disease and to type 1 diabetes mellitus. J. Clin. Endocrinol. Metab., **82**: 143-146.
- Doran MF, Crowson CS, O'Fallon WM and Gabriel SE (2004). The effect of oral contraceptives and estrogen replacement therapy on the risk of rheumatoid arthritis: a population based study. *J. Rheumatol.*, **31**(2): 207-213.
- Doran MF, Pond GR and Crowson CS *et al* (2002). Trends in incidence and mortality in rheumatoid arthritis in Rochester, Minnesota, over a forty-year period. *Arthritis Rheum*, **46**: 625-631.
- du Montcel ST, Michou L and Petit-Teixeira E *et al* (2005). New classification of HLA-DRB1 alleles supports the shared epitope hypothesis of rheumatoid arthritis susceptibility. *Arthritis Rheum*, **52**: 1063-1068.
- Emery P, Salmon M (1995) Early rheumatoid arthritis: Time to aim for remission? *Ann. Rheum Dis.*, **54**: 944-947.
- Eyre S, Bowes J, Diogo D, Lee A, Barton A and Martin P *et al* (2012). High-density genetic mapping identifies new susceptibility loci for rheumatoid arthritis. *Nat. Genet*, **44**(12): 1336-1340.
- Fan J, Edsen-Moore MR, Turner LE, Cook RT, Legge KL and Waldschmidt TJ *et al* (2011). Mechanisms by which chronic ethanol feeding limits the ability of dendritic cells to stimulate T-cell proliferation. *Alcohol. Clin. Exp. Res.*, **35**: 47-59.
- Fiorillo E, Orru V, Stanford SM, Liu Y, Salek M and Rapini N *et al* (2010). Autoimmune associated PTPN22 R620W variation reduces phosphorylation of lymphoid phosphatase on an inhibitory tyrosine residue. *J. Biol. Chem.*, **285**(34): 26506-26518.
- Fitzpatrick R, Newman S, Archer R and Shipley M (1991). Social support, disability and depression: A longitudinal study of rheumatoid arthritis. *Soc. Sci. Med.*, **33**(5): 605-611.
- Fleming A, Crown JM and Corbett M (1976). Early rheumatoid disease. *I. Onset. Ann. Rheum Dis.*, **35**(4): 357-360.
- Freeman GJ, Borriello F, Hodes RJ, Reiser H and Gribben JG *et al* (1993a). Murine B7-2, an alternative CTLA4 counter-receptor that costimulates Tcell proliferation and interleukin2 production. *J. Exp. Med.*, **178**: 2185-2192.
- Freeman GJ, Gribben JG, Boussiotis VA, Ng JW and Restivo VA Jr *et al* (1993b). Cloning of B7-2: A CTLA-4 counter-receptor that costimulates human T cell proliferation. *Science*, **262**: 909-911.
- Frucht DM, Aringer M, Galon J, Danning C, Brown M and Fan S *et al* (2000). Stat 4 is expressed in activated peripheral blood monocytes, dendritic cells and

macrophages at sites of Th1-mediated inflammation. J. Immunol., 164: 4659.

- Funovits J, Aletaha D, Bykerk V, Combe B, Dougados M, Emery P and Felson D *et al* (2010). The 2010 American college of rheumatology/european league against rheumatism classification criteria for rheumatoid arthritis: methodological report phase I. *Ann. Rheum. Dis.*, **69**(9): 1589-1595.
- Furner SE, Hootman JM, Helmick CG, Bolen J and Zack MM (2011). Health-related quality of life of US adults with arthritis: Analysis of data from the behavioral risk factor surveillance system, 2003, 2005 and 2007. *Arthritis. Care Res. (Hoboken)*, **63**(6): 788-799.
- Geraghty D, Inoko H, Beck S, Trowsdale J, Campbell D and Rowen L *et al* (1999). Complete sequence and gene map of a human major his to compatibility complex. The MHC Sequencing Consortium. *Nature*, **401**(6756): 921-923.
- Gjörloff-Wingren A, Saxena M, Williams S, Hammi D and Mustelin T (1999). Characterization of TCRinduced receptor-proximal signaling events negatively regulated by the protein tyrosine phosphatase PEP. *Eur. J. Immunol.*, **29**(12): 3845-3854.
- Gorman JD, Lum RF, Chen JJ, Suarez-Almazor ME, Thomson G and Criswell LA (2004). Impact of shared epitope genotype and ethnicity on erosive disease: A metaanalysis of 3,240 rheumatoid arthritis patients. *Arthritis Rheum*, **50**: 400-412.
- Gregersen PK (2005). Gaining insight into PTPN22 and autoimmunity. *Nat. Genet*, **37**(12): 1300-1302.
- Gregersen PK, Silver J and Winchester RJ (1987). The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. *Arthritis Rheum*, **30**: 1205-1213.
- Guillemin F, Briancon S and Klein JM *et al* (1994). Low incidence of rheumatoid arthritis in France. *Scand J. Rheumatol.*, **23**: 264-268.
- Guillemin F, Saraux A, Guggenbuhl P, Roux CH, Fardellone P and LeBihan E *et al* (2005). Prevalence of rheumatoid arthritis in France: 2001. Ann. Rheum. Dis. **64**: 1427-1430.
- Hamlin PJ, Jones PF, Leek JP, Bransfield K, Lench NJ, Aldersley MA, Howdle PD, Markham AF and Robinson PA (1998). Assignment of GALGT encoding beta-1, 4N-acetylgalactosaminyl-transferase (GalNAc-T) and KIF5A encoding neuronal kinesin (D12S1889) to human chromosome band 12q13 by assignment to ICI YAC 26EG10 and in situ hybridization. *Cytogenet. Cell Genet*, **82**: 267-268.
- Harney SM, Vilari^{*}no-G^{*}uell C and Adamopoulos IE *et al* (2008). Fine mapping of the MHC class III region demonstrates association of AIF1 and rheumatoid arthritis. *Rheumatology (Oxford)*, **47**: 1761-1767.
- Harper K, Balzano C, Rouvier E, Mattei MG, Luciani MF and Golstein P (1991). CTLA-4 and CD28 activated lymphocyte molecules are closely related in both

mouse and human as to sequence, message expression, gene structure and chromosomal location. *J. Immunol.*, **147**: 1037-1044.

- Hart JE, Laden F, Puett RC, Costenbader KH and Karlson EW (2009). Exposure to traffic pollution and increased risk of rheumatoid arthritis. *Environ Health Perspect*, **117**(7): 1065-1069.
- Hasegawa K, Martin F, Huang G, Tumas D, Diehl L and Chan AC (2004). PEST domain-enriched tyrosine phosphatase (PEP) regulation of effect or/memory T cells. *Science*, **303**: 685-689.
- Hawley DJ and Wolfe F (1988). Anxiety and depression in patients with rheumatoid arthritis: A prospective study of 400 patients. J. Rheumatol., **15**(6): 932-941.
- Hazes JM, Dijkmans BA, Vandenbroucke JP, de Vries RR and Cats A (1990). Lifestyle and the risk of rheumatoid arthritis: cigarette smoking and alcohol consumption. *Ann. Rheum Dis.*, **49**: 980-982.
- Heliovaara M, Aho K, Aromaa A, Knekt P and Reunanen A (1993). Smoking and risk of rheumatoid arthritis. *J. Rheumatol.*, **20**: 1830-1835.
- Hill RJ, Zozulya S, Lu YL, Ward K, Gishizky M and Jallal B (2002). The lymphoid protein tyrosine phosphatase Lyp interacts with the adaptor molecule Grb2 and functions as a negative regulator of T-cell activation. *Exp. Hematol.*, **30**(3): 237-244.
- Imboden JB (2009). The immunopathogenesis of rheumatoid arthritis. *Annu. Rev. Pathol.*, **4**: 417-434.
- Jalil SF, Bhatti A, Demirci FY, Wang X, Ahmed I, Ahmed M, Barmada MM, Malik JM, John P and Kamboh MI (2013). Replication of european rheumatoid arthritis loci in a Pakistani population. *J. Rheumatol.*, **40**(4): 401-407.
- Jonsson IM, Verdrengh M, Brisslert M, Lindblad S, Bokarewa M and Islander U *et al* (2007). Ethanol prevents development of destructive arthritis. *Proc. Nat. Acad. Sci. USA*, **104**: 258-263.
- Kaipiainen-Seppanen O and Aho K (2000). Incidence of chronic inflammatory joint diseases in Finland in 1995. *J. Rheumatol.*, 27: 94-100.
- Kaipiainen-Seppanen O and Kautiainen H (2006). Declining trend in the incidence of rheumatoid factor positive rheumatoid arthritis in Finland 1980-2000. J. *Rheumatol.*, **33**(11): 2132-2138.
- Kallberg H, Jacobsen S, Bengtsson C, Pedersen M, Padyukov L and Garred P *et al* (2009). Alcohol consumption is associated with decreased risk of rheumatoid arthritis: results from two Scandinavian case-control studies. *Ann. Rheum Dis.*, **68**: 222-227.
- Karlson E, Lee I, Cook N, Manson J, Buring J and Hennekens C (1999). A retrospective cohort study of cigarette smoking and risk of rheumatoid arthritis in female health professionals. Arthritis Rheum **42**: 910-917.
- Karlson EW, Chang SC, Cui J, Chibnik LB, Fraser PA, Devivo I and Costenbader KH (2010). Geneenvironment interaction between *HLA-DRB1* shared

epitope and heavy cigarette smoking in predicting incident RA. Ann. Rheum Dis., **69**(1): 54-60.

- Kerr GS, Sabahi I, Richards JS, Caplan L, Cannon GW and Reimold A *et al* (2010). Prevalence of vitamin D insufficiency/deficiency in rheumatoid arthritis and associations with disease severity and activity. *J. Rheumatol.*, **38**(1): 53-59.
- Klareskog L, Stolt P, Lundberg K, Kallberg H, Bengtsson C and Grunewald J *et al* (2006). A new model for an etiology of rheumatoid arthritis: Smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. *Arthritis Rheum*, **54**: 38-46.
- Knevel R, Knevel R, Gröndal G, Huizinga TW, Visser AW, Jónsson H, Víkingsson A, Geirsson AJ, Steinsson K and vander Helm-van Mil AH (2012). Genetic predisposition of the severity of joint destruction in rheumatoid arthritis: A population-based study. *Ann. Rheum Dis.*, **71**: 707-709.
- Kobayashi S, Ikari K, Kaneko H, Kochi Y, Yamamoto K, Shimane K and Nakamura Y *et al* (2008). Association of STAT4 with susceptibility to rheumatoid arthritis and systemic lupus erythematosus in the Japanese population. *Arthritis Rheum*, **58**: 1940-1946.
- Kochi Y, Okada Y, Suzuki A, Ikari K, Terao C and Takahashi A *et al* (2010). A regulatory variant in CCR6 is associated with rheumatoid arthritis susceptibility. *Nat. Genet*, **42**: 515-519.
- Kristiansen OP, Larsen ZM and Pociot F (2000). CTLA-4 in autoimmune diseases a general susceptibility gene to autoimmunity? *Genes Immun.*, **1**: 170-184.
- Lai HM and Chen CJ (2008). Biologics in the treatment of rheumatoid arthritis: recent advances. *Formosan Journal of Rheumatology*, **22**: 12-24.
- Lee DM and Weinblatt ME (2001). Rheumatoid arthritis. *Lancet*, **358**: 903-911.
- Lee HS, Remmers EF, Le JM, Kastner DL, Bae SC and Gregersen PK (2007). Association of STAT 4 with rheumatoid arthritis in the Korean population. *Mol. Med.*, **13**: 455-460.
- Lee KM, Chuang E, Griffin M, Khattri R and Hong DK *et al* (1998). Molecular basis of Tcell inactivation by CTLA-4. *Science*, **282**: 2263-2266.
- Lee YH, Woo JH, Choi SJ, Ji JD and Song GG (2010). Association between the rs7574865 polymorphism of STAT4 and rheumatoid arthritis: A meta-analysis. *Rheumatol. Int.*, **30**(5): 661-666.
- Lee YH, Bae SC, Choi SJ, Ji JD and Song GG (2012). Association between the *CTLA4* +49A/G polymorphism and susceptibility to rheumatoid arthritis: A meta-analysis. *Mol. Biol. Rep.*, **39**: 5599-5605.
- Leonard WL and O'Shea JJ (1998). JAKS and STATs: Biological implications. *Annu. Rev. Immunol.*, **16**: 293.
- Li S and Micheletti R (2011). Role of diet in rheumatic disease. *Rheum. Dis. Clin. North Am.*, 37(1): 119-133.

- Liang YL, Wu H, Shen X, Li PQ, Yang XQ, Liang L, Tian WH, Zhang LF and Xie XD (2012). Association of *STAT4* rs7574865 polymorphism with autoimmune diseases: A meta-analysis. *Mol. Biol. Rep.*, **39**(9): 8873-8882.
- Lin CH, Cho CL and Tsai WC *et al* (2006). Inhibitors of kB-like gene polymorphisms in rheumatoid arthritis. *Immunol. Lett.*, **105**: 193-197.
- Lindqvist E, Eberhardt K, Bendtzen K, Heinegard D and Saxne T (2005). Prognostic laboratory markers of joint damage in rheumatoid arthritis. Ann. Rheum Dis., **64**: 196-201.
- Lindsten T, Lee KP, Harris ES, Petryniak B and Craighead N *et al* (1993). Characterization of CTLA-4 structure and expression on human T cells. *J. Immunol.*, **151**: 3489-3499.
- Ling V, Wu PW, Finnerty HF, Sharpe AH, Gray GS and Collins M (1999). Complete sequence determination of the mouse and human CTLA4 gene loci: Cross-species DNA sequence similarity beyond exon borders. *Genomics*, **60**: 341-355.
- Linsley PS, Brady W, Urnes M, Grosmaire LS, Damle NK and Ledbetter JA (1991). CTLA-4 is a second receptor for the B cell activation antigen B7. *J. Exp. Med.*, **174**: 561-569.
- Linsley PS, Clark EA and Ledbetter JA (1990). T-cell antigen CD28 mediates adhesion with B cells by interacting with activation antigen B7/BB-1. *Proc. Natl. Acad. Sci.*, USA **87**: 5031-5035.
- Linsley PS, Greene JL, Brady W, Bajorath J, Ledbetter JA and Peach R (1994). Human B7-1 (CD80) and B7-2 (CD86) bind with similar avidities but distinct kinetics to CD28 and CTLA-4 receptors. *Immunity*, **1**: 793-801.
- Linsley PS, Nadler SG, Bajorath J, Peach R and Leung HT *et al* (1995). Binding stoichiometry of the cytotoxic T lymphocyte-associated molecule-4 (CTLA-4). A disulfide-linked homodimer binds two CD86 molecules. *J. Biol. Chem.*, **270**: 15417-15424.
- Lorant V, Deliege D, Eaton W, Robert A, Philippot P and Ansseau M (2003). Socioeconomic inequalities in depression: A meta-analysis. *Am. J. Epidemiol.*, **157**(2): 98-112.
- MacGregor AJ, Snieder H, Rigby AS, Koskenvuo M, Kaprio J, Aho K and Silman AJ (2000). Characterizing the quantitative genetic contribution to rheumatoid arthritis using data from twins. *Arthritis Rheum*, **43**: 30-37.
- Mackenbach JP, Kunst AE, Cavelaars AE, Groenhof F and Geurts JJ (1997). Socio- economic inequalities in morbidity and mortality in western Europe. The EU Working Group on Socioeconomic Inequalities in Health. *Lancet*, **349**(9066): 1655-1659.
- Magistrelli G, Jeannin P, Herbault N, Benoit DC and Gauchat JF *et al* (1999). A soluble form of CTLA-4 generated by alternative splicing is expressed by non stimulated human T cells. *Eur. J. Immunol.*, **29**: 3596-3602.

- Majithia V and Geraci SA (2007). Rheumatoid arthritis: Diagnosis and management. *Am. J. Med.*, **120**: 936-939.
- Makrygiannakis D, Hermansson M, Ulfgren AK, Nicholas AP, Zendman AJ and Eklund A *et al* (2008). Smoking increases peptidylarginine deiminase 2 enzyme expression in human lungs and increases citrullination in BAL cells. *Ann. Rheum Dis.*, **67**(10): 1488-1492.
- Mandrekar P, Catalano D, Dolganiuc A, Kodys K and Szabo G (2004). Inhibition of myeloid dendritic cell accessory cell function and induction of T cell anergy by alcohol correlates with decreased IL-12 production. *J. Immunol.*, **173**: 3398-3407.
- Marinou I, Maxwell JR and Wilson AG (2010). Genetic influences modulating the radiological severity of rheumatoid arthritis. *Ann. Rheum Dis.*, **69**: 476-482.
- Martinez A, Varade J, Marquez A, Cenit MC, Espino L, Perdigones N and Santiago JL *et al* (2008). Association of the STAT 4 gene with increased susceptibility for some immune-mediated diseases. *Arthritis Rheum*, **58**: 2598-2602.
- Mathieu C, Van Etten E and Gysemans C *et al* (2001). *In vito* and *in vivo* analysis of the immune system of vitamin D receptor knockout mice. *J. Bone Miner Res.*, **16**: 2057-2065.
- Mathur AN, Chang HC, Zisoulis DG, Stritesky GL, Yu Q, O'Malley JT, Kapur R, Levy DE, Kansas GS and Kaplan MH (2007). Stat 3 and Stat 4 direct development of IL-17-secreting Th cells. *J. Immunol.*, **178**: 4901-4907.
- Mattey DL, Hassell AB, Dawes PT, Ollier WE and Hajeer A (1999). Interaction between tumor necrosis factor microsatellite polymorphisms and the HLA-DRB1 shared epitope in rheumatoid arthritis: Influence on disease outcome. *Arthritis Rheum*, **42**: 2698-2704.
- Maxwell JR, Gowers IR, Moore DJ and Wilson AG (2010). Alcohol consumption is inversely associated with risk and severity of rheumatoid arthritis. *Rheumatology (Oxford)*, **49**: 2140-2146.
- McCabe CJ, Akehurst RL, Kirsch J, Whitfield M, Backhouse M, Woolf AD, Scott DL, Emery P, Haslock I (1998). Choice of NSAID and management strategy in rheumatoid arthritis and osteoarthritis. The impact on costs and outcomes in the UK. *Pharmacoeconomics*, **14**(2): 191-199.
- McKellar G, Morrison E and McEntegart A *et al* (2007). A pilot study of a Mediterranean-diet intervention in female patients with rheumatoid arthritis living in areas of social deprivation in Glasgow. *Ann. Rheum. Dis.*, **66**(9): 1239-1243.
- Metzler WJ, Bajorath J, Fenderson W, Shaw SY and Constantine KL *et al* (1997). Solution structure of human CTLA-4 and delineation of a CD80/CD86 binding site conserved in CD28. *Nat. Struct. Biol.*, **4**: 527-531.

- Mitchell JM, Burkhauser RV and Pincus T (1988). The importance of age education and co-morbidity in the substantial earnings losses of individuals with symmetric polyarthritis. *Arthritis Rheum*, **31**(3): 348-357.
- Morinobu A, Gadina M and Strober W *et al* (2002). STAT4 serine phosphorylation is critical for IL-12induced IFN-gamma production but not for cell proliferation. *Proc. Natl. Acad. Sci. USA*, **99**: 12281-12286.
- Mu H, Chen JJ, Jiang Y, King MC, Thomson G and Criswell LA (1999). Tumor necrosis factor a micro satellite polymorphism is associated with rheumatoid arthritis severity through an interaction with the HLA-DRB1 shared epitope. *Arthritis Rheum*, **42**: 438-442.
- Murphy KM and Reiner SL (2002). The lineage decisions of helper T cells. *Nat. Rev. Immunol.*, **2**: 933.
- Newton JL, Harney SM, Wordsworth BP and Brown MA (2004). A review of the MHC genetics of rheumatoid arthritis. *Genes Immun.*, **5**: 151-157.
- Nishikomori R, Usui T, Wu CY, Morinobu A, O'Shea JJ and Strober W (2002). Activated STAT 4 has an essential role in Th1 differentiation and proliferation that is independent of its role in the maintenance of IL-12R beta 2 chain expression and signaling. *J. Immunol.*, **169**: 4388-4398.
- Nistico L, Buzzetti R, Pritchard LE, Van der Auwera B and Giovannini C *et al* (1996). The CTLA-4 gene region of chromosome 2q33 is linked to, and associated with, type 1 diabetes. Belgian Diabetes Registry. *Hum. Mol. Genet.*, **5**: 1075-1080.
- Oaks MK, Hallett KM, Penwell RT, Stauber EC, Warren SJ and Tector AJ (2000). A native soluble form of CTLA-4. *Cell Immunol.*, **201**: 144-153.
- Okada Y, Terao C, Ikari K, Kochi Y, Ohmura K and Suzuki A *et al* (2012). Meta analysis identifies nine new loci associated with rheumatoid arthritis in the Japanese population. *Nat. Genetics*, **44**(5): 511-516.
- Orozco G, Alizadeh BZ, Delgado-Vega AM, Gonzalez-Gay MA, Balsa A, Pascual-Salcedo D and Fernandez-Gutierrez B *et al* (2008). Association of STAT 4 with rheumatoid arthritis: A replication study in three European populations. *Arthritis Rheum*, **58**: 1974-1980.
- Ostrov DA, Shi W, Schwartz JC, Almo SC and Nathenson SG (2000). Structure of murine CTLA-4 and its role in modulating T cell responsiveness. *Science*, **290**: 816-819.
- Ota M, Katsuyama Y and Kimura A *et al* (2001). A second susceptibility gene for developing rheumatoid arthritis in the human MHC is localized within a 70-kb interval telomeric of the TNF genes in the HLA class III region. *Genomics*, **71**: 263-270.
- Padyukov L, Silva C, Stolt P, Alfredsson L and Klareskog L (2004). A gene environment interaction between smoking and shared epitope genes in HLA-DR provides a high risk of seropositive rheumatoid arthritis. *Arthritis Rheum*, **50**: 3085-3092.

- Palomino-Morales RJ, Rojas-Villarraga A, Gonzalez CI, Ramirez G, Anaya JM and Martin J (2008). STAT 4 but not TRAF1/C5 variants influence the risk of developing rheumatoid arthritis and systemic lupus erythematosus in Colombians. *Genes Immun.*, **9**: 379-382.
- Patel S, Farragher T, Berry J, Bunn D, Silman A and Symmons D (2007). Association between serum vitamin D metabolite levels and disease activity in patients with early inflammatory polyarthritis. *Arthritis Rheum*, **56**(7): 2143-2149.
- Pattison DJ, Symmons DP and Lunt M *et al* (2004). Dietary risk factors for the development of inflammatory polyarthritis: Evidence for a role of high level of red meat consumption. *Arthritis Rheum*, **50**: 3804-3812.
- Pedersen JK, Kjaer NK, Svendsen AJ and Hørslev-Petersen K (2009). Incidence of rheumatoid arthritis from 1995 to 2001: Impact of ascertainment from multiple sources. *Rheumatol. Int.*, **29**(4): 411-415.
- Pedersen M, Jacobsen S, Klarlund M, Pedersen BV, Wiik A and Wohlfahrt J *et al* (2006). Environmental risk factors differ between rheumatoid arthritis with and without auto-antibodies against cyclic citrullinated peptides. *Arthritis Res. Ther.*, **8**: R133.
- Perkins D, Wang Z, Donovan C, He H and Mark D *et al* (1996). Regulation of CTLA-4 expression during T cell activation. *J. Immunol.*, **156**: 4154-4159.
- Pioli C, Gatta L, Ubaldi V and Doria G (2000). Inhibition of IgG1 and IgE production by stimulation of the B cell CTLA-4 receptor. *J. Immunol.*, **165**: 5530-5536.
- Pistillo MP, Tazzari PL, Palmisano GL, Pierri I and Bolognesi A *et al* (2003). CTLA-4 is not restricted to the lymphoid cell lineage and can function as a target molecule for apoptosis induction of leukemic cells. *Blood*, **101**: 202-209.
- Plant D, Flynn E, Mbarek H, Dieude P, Cornelis F and Arlestig L *et al* (2010). Investigation of potential non-HLA rheumatoid arthritis susceptibility loci in a European cohort increases the evidence for nine markers. *Ann. Rheum Dis.*, **69**(8): 1548-1553.
- Raychaudhuri S, Remmers EF, Lee AT, Hackett R, Guiducci C and Burtt NP *et al* (2008). Common variants at *CD40* and other loci confer risk of rheumatoid arthritis. *Nat. Genet*, **40**(10): 1216-1223.
- Rayman MP and Callaghan A (2006). Nutrition and arthritis: Blackwell Publishing Ltd. Oxford, UK, p. 264.
- Remmers EF, Plenge RM, Lee AT, Graham RR, Hom G, Behrens TW and de Bakker PI *et al* (2007). STAT 4 and the risk of rheumatoid arthritis and systemic lupus erythematosus. *N. Engl. J. Med.*, **357**: 977-986.
- Rieck M, Arechiga A, Onengut-Gumuscu S, Greenbaum C and Concannon P *et al* (2007). Genetic variation in PTPN 22 corresponds to altered function of T and B lymphocytes. *J. Immunol.*, **179**: 4704-4710.

- Ruddy S, Harris ED, Sledge CB and Kelley WN (2005). Clinical features of rheumatoid arthritis. *In*: eds. Kelley's Textbook of rheumatology, 7th edn, WB Saunders, Philadelphial, pp.1043-1078.
- Saraux A, Guedes C, Allain J, Devauchelle V, Valls I and Lamour A *et al* (1999). Prevalence of rheumatoid arthritis and spondyloarthropathy in Brittany, France. *J. Rheumatol.*, **26**: 2622-2627.
- Sato M, Miyazaki T, Nagaya T, Murata Y, Ida N and Maeda K *et al* (1996). Antioxidants inhibit tumor necrosis factor-alpha mediated stimulation of interleukin-8, monocyte chemo attractant protein-1 and collagenase expression in cultured human synovial cells. *J. Rheumatol.*, **23**(3): 432-438.
- Savolainen E, Kaipiainen-Seppanen O and Kroger L *et al* (2003). Total incidence and distribution of inflammatory joint diseases in a defined population: Results from the Kuopio 2000 arthritis survey. *J. Rheumatol.*, **30**: 2460-2468.
- Schneider H, Prasad KV, Shoelson SE and Rudd CE (1995). CTLA-4 binding to the lipid kinase phosphatidylinositol 3-kinase in T cells. *J. Exp. Med.*, **181**: 351-355.
- Seldin MF, Amos CI, Ward R and Gregersen PK (1999). The genetics revolution and the assault on rheumatoid arthritis. *Arthritis Rheum*, **42**: 1071-1079.
- Shapira Y, Agmon-Levin N and Shoenfeld Y (2010). Geoepidemiology of autoimmune rheumatic diseases. *Nat. Rev. Rheumatol.*, **6**(8): 468-476.
- Shiratori T, Miyatake S, Ohno H, Nakaseko C and Isono K *et al* (1997). Tyrosine phosphorylation controls internalization of CTLA-4 by regulating its interaction with clathrinassociated adaptor complex AP-2. *Immunity.*, **6**: 583-589.
- Silman AJ and Pearson JE (2002). Epidemiology and genetics of rheumatoid arthritis. *Arthritis Res.*, **4**: 265-272.
- Silman AJ and Symmons DP (1995). Selection of study population in the development of rheumatic disease criteria: Comment on the article by the American college of rheumatology diagnostic and therapeutic criteria committee. *Arthritis Rheum*, **38**: 722-723.
- Silman AJ, MacGregor AJ, Thomson W, Holligan S, Carthy D, Farhan A and Ollier WE (1993). Twin concordance rates for rheumatoid arthritis: Results from a nationwide study. *Br. J. Rheumatol.*, **32**: 903-907.
- Skapenko A, Leipe J, Lipsky PE and Schulze-Koops H (2005). The role of the T cell in autoimmune inflammation. *Arthritis Res. Ther.*, **7**(2): S4-14.
- Skoldstam L, Hagfors L and Johansson G (2003). An experimental study of a Mediterranean diet intervention for patiets with rheumatoid arthritis. *Ann. Rheum Dis.*, **62**(3): 208-214.
- Smedslund G, Byfuglien M and Olsen S *et al* (2010). Effectiveness and safety of dietary interventions for rheumatoid arthritis: A systematic review of

randomized controlled trials. J. Am. Diet. Assoc., 110(5): 727-735.

- Smolen JS, Aletaha D, Grisar J, Redlich K, Steiner G and Wagner O (2008). The need for prognosticators in rheumatoid arthritis. Biological and clinical markers: Where are we now? *Arthritis Res. Ther.*, **10**(3): 208.
- Stahl EA and Raychaudhuri S (2012). Evidence for a genetic component to disease severity in RA. *Nat. Rev. Rheumatol.*, **8**: 312-313.
- Stahl EA, Raychaudhuri S, Remmers EF, Xie G, Eyre S and Thomson BP *et al* (2010). Genome-wide association study meta-analysis identifies seven new rheumatoid arthritis risk loci. *Nat. Genet*, **42**: 508-514.
- Stansfeld SA, Head J and Marmot MG (1998). Explaining social class differences in depression and wellbeing. *Soc. Psychiatry Psychiatr. Epidemiol.*, **33**(1): 1-9.
- Steinman L (2007). A brief history of T (H)17, the first major revision in the T(H)1/T(H)2 hypothesis of T cellmediated tissue damage. *Nat. Med.*, 13: 139-145.
- Stolt P, Yahya A, Bengtsson C, Kallberg H, Ronnelid J and Lundberg I *et al* (2010). Silica exposure among male current smokers is associated with a high risk of developing ACPA-positive rheumatoid arthritis. *Ann. Rheum Dis.*, **69**(6): 1072-1076.
- Stolt P, Bengtsson C, Nordmark B, Lindblad S, Lundberg I, Klareskog L and Alfredsson L (2003). Quantification of the influence of cigarette smoking on rheumatoid arthritis: Results from a population based case-control study, using incident cases. *Ann. Rheum Dis.*, **62**: 835-841.
- Stringhini S, Sabia S, Shipley M, Brunner E, Nabi H, Kivimaki M and Singh-Manoux A (2010). Association of socioeconomic position with health behaviors and mortality. *JAMA*, **303**(12): 1159-1166.
- Symmons D, Turner G, Webb R, Asten P, Barrett E and Lunt M *et al* (2002). The prevalence of rheumatoid arthritis in the United Kingdom: New estimates for a new century. *Rheumatology (Oxford)*, **41**: 793-800.
- Symmons DP, Bankhead C, Harrison BJ, Brennan P, Barrett E and Scott DG *et al* (1997). Blood transfusion, smoking and obesity as risk factors for the development of rheumatoid arthritis: Results from a primary care-based incident case-control study in Norfolk, England. *Arthritis Rheum*, **40**: 1955-1961.
- Symmons DP, Barrett EM and Bankhead CR *et al* (1994). The incidence of rheumatoid arthritis in the United Kingdom: Results from the Norfolk Arthritis Register. *Br. J. Rheumatol.*, **33**: 735-739.
- Takahashi T, Tagami T, Yamazaki S, Uede T and Shimizu J *et al* (2000). Immunologic self tolerance maintained by CD25+CD4+ regulatory T cells constitutively expressing cytotoxic T lymphocyte-associated antigen 4. *J. Exp. Med.*, **192**: 303-310.
- Tamiya G, Shinya M and Imanishi T *et al* (2005). Whole genome association study of rheumatoid arthritis using 27 039 microsatellites. *Hum. Mol. Genet*, **14**: 2305-2321.

- Tłustochowicz W (2006). Rational therapeutic approach in rheumatoid arthritis. *Ann. Acad. Med. Stetin.*, **52**(2): 5-10.
- Tobón GJ, Youinou P and Saraux A (2010) The environment, geo-epidemiology and autoimmune disease: Rheumatoid arthritis. *J. Autoimmun.*, **35**(1): 10-14.
- Todd JA, Walker NM, Cooper JD, Smyth DJ, Downes K, Plagnol V and Bailey R *et al* (2007). Robust associations of four new chromosome regions from genome-wide analyses of type 1 diabetes. *Nat. Genet*, **39**: 857-864.
- Too CL, Yahya A, Murad S, Dhaliwal JS, Larsson PT, Muhamad NA and Abdullah NA *et al* (2012). Smoking interacts with *HLA-DRB1* shared epitope in the development of anti-citrullinated protein antibodypositive rheumatoid arthritis: Results from the Malaysian epidemiological investigation of rheumatoid arthritis (MyEIRA). *Arthritis Res. Ther.*, **14**(2): R89.
- Ueda H, Howson JM, Esposito L, Heward J, Snook H, *et al* (2003). Association of the T-cell regulatory gene *CTLA4* with susceptibility to autoimmune disease. *Nature*, **423**: 506-511.
- Uhlig T, Hagen K and Kvien T (1999). Current tobacco smoking, formal education and the risk of rheumatoid arthritis. *J. Rheumatol.*, **26**: 47-54.
- van der Heide A, Jacobs JW, Bijlsma JW, Heurkens AH, van Booma-Frankfort C, van der Veen MJ and Haanen HC *et al* (1996). The effectiveness of early treatment with second-line antirheumatic drugs. A randomized, controlled trial. *Ann. Intern. Med.*, **124**: 699-707.
- van Gaalen FA, van Aken J, Huizinga TW, Schreuder GM, Breedveld FC, Zanelli E, van Venrooij WJ, Verweij CL, Toes RE and deVries RR (2004). Association between HLA class II genes and autoantibodies to cyclic citrullinated peptides (CCPs) influences the severity of rheumatoid arthritis. *Arthritis Rheum*, **50**: 2113-2121.
- Vang T, Congia M, Macis MD, Musumeci L, Orrú V, Zavattari P, Nika K, Tautz L, Taskén K, Cucca F, Mustelin T and Bottini N (2005). Autoimmuneassociated lymphoid tyrosine phosphatase is a gain-offunction variant. *Nat. Genet*, **37**(12): 1317-1319.
- Verma S, Alexander CM, Carlson MJ, Tygrett LT and Waldschmidt TJ (2008). B-cell studies in chronic ethanol mice. *Methods. Mol. Biol.*, **447**: 295-323.
- Vessey MP, Villard-Mackintosh L and Yeates D (1987). Oral contraceptives, cigarette smoking and other factors in relation to arthritis. *Contraception*, **35**: 457-464.
- Vijayakrishnan L, Slavik JM, Illes Z, Greenwald RJ and Rainbow D *et al* (2004). An autoimmune diseaseassociated CTLA-4 splice variant lacking the B7 binding domain signals negatively in T cells. *Immunity*, **20**: 563-575.

- Viken MK, Amundsen SS, Kvien TK, Boberg KM and Gilboe IM *et al* (2005). Association analysis of the 1858C. T polymorphism in the PTPN 22 gene in juvenile idiopathic arthritis and other autoimmune diseases. *Genes Immun.*, **6**: 271-273.
- Visconti R, Gadina M, Chiariello M, Chen EH, Stancato LF, Gutkind JS and O'Shea JJ (2000). Importance of the MKK6/p38 pathways for interleukin-12-induced STAT4 serine phosphorylation and transcriptional activity. *Blood*, **96**: 1844.
- Waldschmidt TJ, Cook RT and Kovacs EJ (2006). Alcohol and inflammation and immune responses: Summary of the 2005 alcohol and immunology research interest Group (AIRIG) meeting. *Alcohol*, **38**: 121-125.
- Walker JG, Ahern MJ, Coleman M, Weedon H, Papangelis V and Beroukas D *et al* (2006). Expression of Jak3, STAT 1, STAT 4 and STAT 6 in inflammatory arthritis: Unique Jak 3 and STAT 4 expression in dendritic cells in seropositive rheumatoid arthritis. *Ann. Rheum Dis.*, **65**: 149-156.
- Walker JG, Ahern MJ, Coleman M, Weedon H, Papangelis V and Beroukas D *et al* (2007). Characterisation of a dendritic cell subset in synovial tissue, which strongly expresses Jak/STAT transcription factors from patients with rheumatoid arthritis. *Ann. Rheum Dis.*, **66**: 992-999.
- Wandstrat A and Wakeland E (2001). The genetics of complex autoimmune diseases: Non-MHC susceptibility genes. *Nat. Immunol.*, 2: 802-809.
- Wang LL, Worley K, Gannavarapu A, Chintagumpala MM, Levy ML and Plon SE (2002). Intron-Size constraint as a mutational mechanism in rothmund-Thomson Syndrome. *The American Journal of Human Genetics*, **71**: 165.
- Wang XB, Giscombe R, Yan Z, Heiden T, Xu D and Lefvert AK (2002). Expression of CTLA-4 by human monocytes. *Scand J. Immunol.*, **55**: 53-60.
- Wang XB, Zheng CY, Giscombe R and Lefvert AK (2001). Regulation of surface and intracellular expression of CTLA-4 on human peripheral T cells. *Scand J Immunol.*, **54**: 453-458.
- Watford WT, Hissong BD, Bream JH, Kamnno Y, Muul L and O'Shea JJ (2004). Signaling by IL-12 and IL-23 and the immunoregulatory roles of STAT 4. *Immunol. Rev.*, **202**: 139-156.
- Yan Z, Ferucci ED and Geraghty DE *et al* (2007). Resequencing of the human major his to compatibility complex in patients with rheumatoid arthritis and healthy controls in Alaska Natives of Southeast Alaska. *Tissue Antigens*, **70**: 487-494.
- Yang HC, Liang YJ, Chung CM, Chen JW and Pan WH (2009). Genome-wide gene-based association study. *BMC. Proc.*, **3**(7): S135.

- Zervou MI, Sidiropoulos P, Petraki E, Vazgiourakis V, Krasoudaki E, Raptopoulou A, Kritikos H, Choustoulaki E, Boumpas DT and Goulielmos GN (2008). Association of a TRAF 1 and a STAT 4 gene polymorphism with increased risk for rheumatoid arthritis in a genetically homogeneous population. *Hum. Immunol.*, **69**: 567-571.
- Zuk O, Hechter E, Sunyaev SR and Lander ES (2012). The mystery of missing heritability: Genetic interactions create phantom heritability. *Proc. Natl. Acad. Sci. USA*, **109**(4): 1193-1198.