

REVIEW ARTICLE

POTENTIALLY SIGNIFICANT BIOMARKERS IN ORAL SUBMUCOUS FIBROSIS

Haya Mohammad, Naila Irum Hadi, Shumaila Younus, Farah Ahmed , Naila Younus

ABSTRACT

Oral submucous fibrosis (OSMF) is a chronic, irreversibly progressive and precancerous disease that effects oral, oropharyngeal and oesophageal mucosa. OSMF is characterised by the inflammation and progressive fibrosis of lamina propria that leads to difficulty in mastication, speech, swallowing and causes limited mouth opening. Arecoline (major alkaloid) in areca nut is the main aetiologi cal factor in causing the disease. Prevalance of OSMF ranges from 0.2 % to 0.5% in South India. The malignant rate of transformation over 17-year period was 7.6%. Trace elements (part of metalloenzymes) are recognised as versatile biomarkers which may be helpful in early detection, prognosis and can reduce the incidence of cancer. Copper, Iron, Zinc, Selenium, Cadmium, antioxidants (Superoxide Dimutase, Vitamin A, Vitamin C, Vitamin E), immunoglobulins and alternation in oncosuppressor genes and other genes have been emphasized as biochemical parameters that play an important role in its pathogenesis. These parameters can also serve as important biomarkers in early detection of a premalignant condition and cancer progression.

Key Words : Oral submucous fibrosis, trace elements, biomarkers, antioxidants, oncosuppressor genes

INTRODUCTION

Oral submucous fibrosis is a precancerous disease with immuno-inflamatory processes and if persistent, will result in activation oncogenes and loss of tumor suppressor genes that will promote abnormal cell growth and risk of cancer ¹. It was first described as ' Atrophica idiopathic mucosa oris' by Schwartz (1952) in five Indian women in Kenya as a fibrosing condition. Later it was characterised as insidious, chronic disease by Pindborg (1966) in any part of oral cavity and pharynx ². Oral submucous fibrosis is now recognised as an Indian disease all over the world and has the highest potential of malignancy among all other premalignant lesions ⁴. The malignant rate of transformation over 17-year period was recorded at 7.6 Prevalence of OSMF was approximately 0.2% to 0.5% (mostly among 20 and 40 years of age) with higher percentage in South India ⁴.

Tobacco (smoked/chewable), pan masala, chilli, malnutrition, autoimmunity and genetic predisposition are multiple factors but areca nut has the main contribution in causation of disease⁴. Studies show that fourth most addictive substance in the world is areca nut ⁶. The

commercially prepared forms of areca nut are scented supari, gutka, mawa, mainpuri tobacco, pan masala and betel quid (may be either with or without tobba co) in the presence of slaked lime, catechu, flavouring agent, etc. These factors play the key role in phenotypical alteration of fibroblasts which lead to fibrosis of oral mucosa ⁵. Clinically, patients with such fibrotic disease suffers from sensitivity to spicy food, vesiculations, ulceration, blanch-ing, stiffness of oral mucosa resulting in the rigidity of tongue and trismus. Histological findings show focal parakeratosis or hyperkeratosis and atrophy of oral epithe-lium ⁷.

Trace elements (part of metalloenzymes) are recognised as versatile biomarkers that may be helpful in early detec-tion, prognosis and can reduce the incidence of oral cancer ⁸. Biochemical investigations of blood, serum and tissues are the best indicators for disease progress and may serve in broad spectrum analysis of causation of potentially malignant condition ⁹.

The molecular basis for the changes caused by Arecanut Insert the flow chart along with explanation

MATERIALS AND METHOD

This review included all articles that were used for the advancement of information about potential biomarkers in oral submucous fibrosis. Appropriate articles were determined according to a reconsideration of abstracts. Search of academic and published literature was carried utilizing the electronic databases of Pub Med, Google scholar, Elsevier from 2000 to 2014 for English-language articles. The research terms applied were: "biochemical markers and Oral Submucous Fibrosis", "Biomarkers of OSMF"and "OSMF". The subjects, titles and abstracts of articles were appraised. Entire and complete text matter and reviews of the studies and researches were analyzed when the abstract corresponding to the inclusion frame-work. Evaluation of selected data include a serious and detailed review of abstracts or full text papers.

Trace elements

Copper

Copper is the nutrient essential to carry out enzymatic functions important for human metabolism, including cytochrome-c oxidase, superoxide dimutase, metal-lothene and lysyl oxidase. In vitro, raised copper concentrations show increase proliferation of fibroblasts. High copper and ceruloplasmin levels were observed in patients with pre-malignant and malignant oral lesions⁴. Areca nut has a high copper content (302 nmol/g) , the substantial amount of which is released into saliva¹² after 15-30 minutes of chewing areca nut ¹³.

Some studies show that high serum copper is responsible for the severity of OSMF ¹². Margalith et al investigated that damage by copper ions is due to superoxide radicals. These complexes react with hydrogen peroxide to form hydroxyl radicals that causes destruction of RNA, DNA and protein ultimately resulting in the malignancy¹⁴. The reason behind increase serum copper might be the realease of copper containing the ceruloplasmin due to inflammatory response by liver or reduced degradation of serum ceruloplasmin. Cytological study confirmed the important role of copper in pathogenesis of OSMF showing intense staining in smears of OSMF as compared to smears from non chewers. Thus, it can prove an efficient marker of early diagnosis of malignant transformation ¹⁵.

Iron

Epidemiologic studies have established the role of diets rich in vegetables and fruits in oral carcinogenesis, with important contribution of vitamins and iron in maintain-ance of oral mucosa (¹⁶). Iron plays an important role in development, maintenance and defense abilities of oral mucosa. It effects the ability of iron containing enzymes which require heme, biological oxidations, transport and is necessary for DNA, RNA, collagen and antibody synthesis (⁹, ¹⁷). Anemia can be treated by increase iron intake, either by diet diversification, supplementation, or fortifica-tion of foods. The best long-term approach in reduction of the incidence of iron deficiency is food fortification ³⁸.

Oral submucous fibrosis is also considered as an "Asian version of Sideropenic dysphagia". Chronic iron deficien-cy leads to mucosal exposure to irritants such as arecanut ¹⁸. Peptidyl lysine hydroxylase requires molecular

oxygen, ferous iron, alpha-ketoglutr ate and ascorbic acid to form collagen type I coupled with loss of more soluble collagen type III and type IV^{14,18} In eastern countries, Oral submucous fibrosis (OSMF) may be the manifestation of chronic iron (Fe) deficiency and a counterpart of Plummer-Vinson syndrome. The reasons of iron deficiency in OSMF patients could be multifactorial. It may be due to its utilization in collagen formation by the process of hydroxylation of proline and lysine, altered epithelial cell turnover rate or depletion of nutrients due to pronounced difficulty in mastication ¹⁹.

Zinc

Zinc (Zn) is an important part of biomembranes that manages membrane stability and lipid peroxidation-relat-ed injury. It has a role in RNA and DNA polymerase, inhibitory effect on phosphodiesterase, activation of membrane-bound adenylcyclase thus suggesting a role of zinc in carcinogenesis. . Zinc deficiency also contributes to cancer initiation by activation of NF-kB expression and the consequent induction of tumorigenic signaling ²⁰.

Selenium

Selenium (antioxidant nutrient) has been considered as an integral part of the glutathione peroxidase enzyme, type I iodothyronine deiodinase, metalloprotein, fatty acid binding protein and selenoprotein P ⁹, ²⁴. Low serum, plasma or blood levels of selenium have been found to be associated with the incidence of malignant lesions of the oral cavity^{24,25}, breast^{26,27}, ovary^{25,28,29}, oesophagus²⁴, ²⁵, colon²⁸, ²⁹ and prostate ²⁸, ²⁹. It is responsible for immune modulation and cells growth inhibitory properties that effects immune response by making immune cells more resistant to oxidative stress ²⁴, ³⁰. According to the various epidemiological studies, selenium is considered as a protective agent and its dietary intake is of great benefit against cancer ³⁰.

Cadmium

Rajendran et al ²¹ noticed elevation of cadmium (Cd) levels in OSMF where as cadmium values were decreased in Oral Cancer and oral leukoplakia. Cadmium accumulates in the body hence cadmiumn burden increases with age. Increase intestinal absorbtion of Cd (a component of ghutka) can be linked to low iron status in OSMF patients. The Cadmium burden in the body will replace zinc (anti cariogenic agent) and will show its cariogenic effect by reducing zinc anticancer activity ²¹. Cadmium may be one of the cause for malignant transformation of OSMF and its estimation may be a helpful tool in differential diagnosis of premalignant and malignant lesions of the oral mucosa.

Antioxidants

Reactive oxygen species (ROS) generation initiates lipid peroxidation (LPO) which highly promotes the carcino-genesis process. Antioxidants especially enzymatic antioxi-dant like Superoxide Dimutase (SOD), beta carotene and Vitamin A, Vitamin E, Vitamin C and Vitamin E play an important role in this process ⁹. Stahelin et al in his 12-year research on vitamins, plasma antioxidants and subse-quent cancer mortality proved that decrease levels of antioxidants such as beta carotene, Vitamin C and Vitamin E is linked with the increased mortality rate due to cancer ³¹.

Haya Mohammad
M.Phil Candidate of Oral Pathology, Ziauddin University and Hospitals, Karachi.
Naila Irum Hadi
Pathology Department, Ziauddin University and Hospitals, Karachi.
Shumaila Younus
M.Phil Candidate of Oral Pathology, Ziauddin University and Hospitals, Karachi.
Farah Ahmed
Community Health Sciences Department, Ziauddin University and Hospitals, Karachi.
Naila Younus
Resident Radiology, Ziauddin University and Hospitals, Karachi.
Corresponding Author
Haya Mohammad

Superoxide Dismutase

Betel quid generates free radicals in the oral cavity. It is initiated by lipid peroxidation while enzymatic antioxidant superoxide dismutase detoxifies the effect of these harmful radicals (hydrogen peroxide and hydroxyl). These radicals transfer their unpaired electron to oxygen to form superoxide in order to prevent oxidative stress^{14,34}. Beta carotene and Vitamin A

Beta carotene (red-orange coloured pigment) is abundantly present in plants and animals. It is the inactive precursor of Vitamin A⁹. Beta carotene ingestion quickly increases helper T lymphocytes. It plays an important role in OSMF and its level decreases with disease progression³². An irreversibly oxidised form of Vitamin A is retinoic acid which is the principal hormone-like growth factor for maintenance of epithelial and other cells⁽⁹⁾. It has immunoregulatory properties and an excellent radical trap for hydroxyl and peroxy radicals, therefore it should be maintained in adequate levels in the blood³².

Vitamin C (Ascorbic acid)

Vitamin C is an antioxidant scavenging free radical,

reduces vitamin E degradation, inhibits nitrosamine formation, enhances detoxification via cytochrome P450 and iron absorption by reducing dietary iron from ferric form to the ferrous form. Vitamin C is utilized in conversion of proline into hydroxyproline. This hydroxylation reaction requires ferrous iron and Vitamin C. Lysyl oxidase upregulates the collagen cross linkages in the presence of Vitamin C that results in the advancement of the condition from the stage I to stage II. Rajendran et al proved that deficiency of vitamins and iron will result in abnormal repair of the lamina propria. This will result in defective healing and scar formation, which ultimately led to OSMF. Singh et al concluded that the therapeutic supplementation of Vitamin C reduces the oedema between the collagen bundles and regenerates new collagen bundles with good approximation in OSMF patients³³.

Vitamin E

Vitamin E is the fat soluble antioxidant that include both tocopherols and tocotrienols. It ceases the production of reactive oxygen radicals (ROS) formed after the oxidation of fat⁹. Gupta et al evaluated antioxidant parameters and found decreased Vit E level in stage II and III but not in stage I OSMF patients. Studies show that OSMF or the products associated (areca nut and additives) induce oxidative stress on the tissues^{9,35,36}.

Immunoglobulins

The role of active immune response in OSMF is to accelerate body protection and detection of the foreign antigen. This process will cause abnormal lymphocyte function and hyperactivity of B cells. High levels of IgG were observed among OSMF patients by Gupta et al, which is one of the earliest-recorded studies in India¹⁴. In few recent studies increase in IgG and IgA has been noted in OSMF patients.^{14,37}

Alterations in Oncosuppressor Genes and Other Genes

It was evident from earlier studies, that some oncosuppressor genes play an important role in areca related carcinogenesis^{39,40,41}. A progressive reduction of PTEN

expression (tumor suppressor gene) was noticed in Oral submucous fibrosis (OSMF) and Oral Squamous Cell Carcinoma (OSCC). Hence, PTEN alteration is considered as a specific molecular event in carcinogenesis^{41,42,43}. FHIT (Fragile Histidine Triad) is expressed in the epithelium of normal oral mucosa and a decrease in the expression of FHIT was noticed in OSMF and more significantly in carcinoma arising from OSMF. There were 716 genes upregulated and 149 genes downregulated among OSMF patients identified through oligonucleotide microarray that are responsible for pathogenesis and malignant transformation of OSMF⁴¹. Genomic instability denotes early genetic events during the malignant transformation of the disease. This may be due to the Loss of Heterozygosity (LOH) and chromosomal copy number abnormality^{41,42}.

The cytochrome P450 (CYP) gene family functions actively in oxidative metabolism of active endogenous and xenobiotic substrates. Cytochrome P450 has been identified as a genetic biomarker for susceptibility to OSMF and authors have further suggested that individuals with high genetic risk for OSMF could be investigated according to the genetic polymorphisms in some exclusive regions of the Cytochrome P450 3A genes⁴⁶. Genes like CYP2B6, CYP2C18, CYP2F1, CYP3A5, microsomal glutathione S-transferase 2 (MGST2), alcohol dehydrogenase (ADH), UDP glucuronosyl transferase 2B15 (UGT2B15), ADH1C) related to the pathway of CYP metabolism were found to be down regulated in all stages of OSMF⁴⁷. It is suggested that these polymorphisms can be the cause of high risk of OSMF among men if they use arecanut or smokeless tobacco in abundance^{41,48}.

CONCLUSION

Oral submucous fibrosis has a high incidence and carries a significant morbidity rate due to its progression to oral cancer. Cessation of areca nut and ghutka products should be the first step among such patients. Intervention studies and public health awareness programmes linked with hazards of carcinogenic products (areca nut and ghutka that has become the common trend in the Asian society), OSMF conditions and habits may prove the best way to control the disease process at community level. The evaluation of trace elements put the clinician in a better position to determine the stage of precancerous condition and also highlights the importance of iron supplementation and healthy diet as a part of overall treatment of this disease.

REFERENCES

1. Feller L, Altini M, Lemmer J. Inflammation in context to oral cancer. *Oral Oncol* 2013; 49: 887-92.
2. Namboodiripad P. Cystatin C: its role in pathogenesis of OSMF. *J Oral Biol Craniofac Res*. 2014; 4: 42-6.
3. Kalbande A, Khakse G, Priya D, Tamgadage P. Epidemiological Study of oral submucous fibrosis in Yavatmal district. *Int. J Recent Trends Sci Tech*. 2013; 6: 38-40.
4. Arakeri G, Hunasgi S, Colbert S, Merx M, Brennan P. Role of drinking water copper in pathogenesis of oral submucous fibrosis: a prospective case control study. *BJOMS* 2014; 52: 507-12.
5. Sarode S, Mahuli A, Sarode G, Mahuli S. Why only areca nut chewing cannot cause oral submucous fibrosis? *Med Hypothesis* 2013; 81: 47-9.

Fig 1: Molecular Changes Caused By Arecanut – Salivary Arecoline concentration during betel quid chewing may lead to 5.7µg/ml- 97.4 7µg/ml. Studies show arecoline causes upregulation of cytokines (IL-6, TNF, INF alpha), Cystatin C (2), Nuclear factor-kappa B (NF-Kb), Tissue Inhibitor Metalloproteinase (TIMP) which are one of the factors responsible for increase collagen formation and MMP-2 and MMP-9 were found in minimal amounts that ultimately leads to fibrosis. Epithelial Mesenchyme Transition (EMT) a process that contributes to tumour cell invasion is a newly forwarded concept that has gained substantial attention recently.

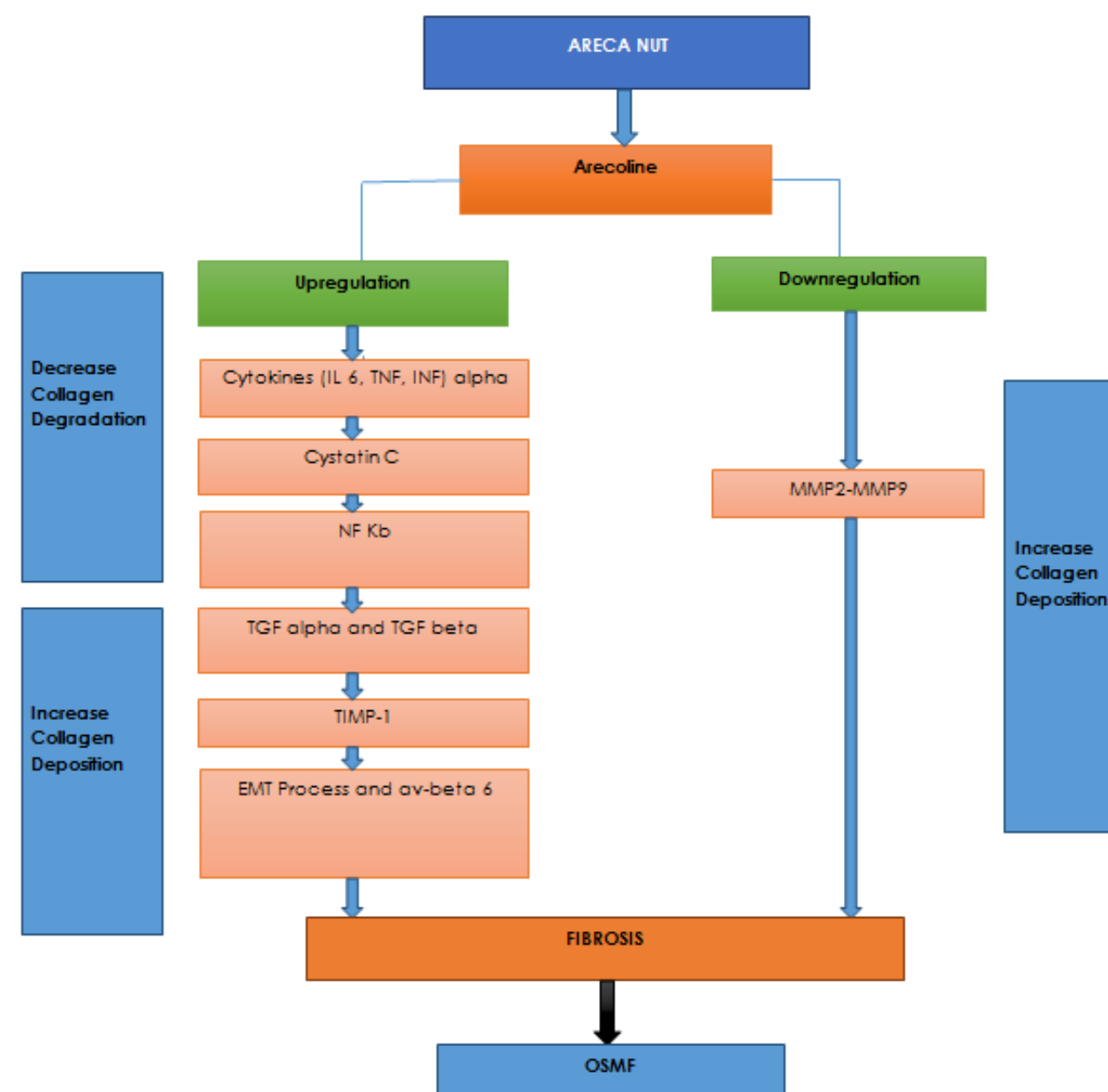


Table 1. Classification of Potential Biomarkers in OSMF

Potential Biomarkers	Role In Oral submucous fibrosis (OSMF)
1) Trace Elements	
Copper (Cu)	Forms hydroxyl radicals and increase fibroblast proliferation.
Iron (Fe)	Peptidyl lysine requires ferrous iron to form collagen type I that promotes fibrosis.
Zinc (Zn)	Active component of antioxidant enzymes (superoxide dimutase, etc), interfere in Cu absorption and acts as antifibrotic agent.
Selenium (Se)	Immune modulation and consists of growth inhibitory properties.
Cadmium (Cd)	Cd burden replace Zn (anticarcinogenic agent) and linked to low iron status in OSMF patients.
2) Antioxidants	
Superoxide Dimutase	Detoxifies free radical (hydrogen peroxide and hydroxyl effect.
Beta carotene and Vitamin A	Excellent radical trap and decrease disease progression.
Vitamin C (Ascorbic acid)	Upregulates collagen cross linkages, enhances detoxification and iron absorption.
Vitamin E	Ceases oxidative stress.
3) Immunoglobins	
IgG and IgA	Accelerates body protection.
4) Oncosuppressor gene and other genes alterations	
PTEN	Decrease expression in areca related carcinogenesis
FHIT	Decrease expression specifically in carcinoma arising from OSMF.
CYP gene family	Function in oxidative metabolism of active xenobiotic and endogenous substrates.

⁶ Kerr AR, Warnakulasuriya S, Mighell AJ, Dietrich T, Nasser M, Rimal J et al. A systemic review of medical interventions for oral submucous fibrosis and future research opportunities. *Oral Diseases* 2010; 17: 42-57.

⁷ Chang M, Lin D, Wu H, Ho Y, Hsein H, Wang T et al. Areca nut-induced buccal mucosa fibroblast contraction and its signaling: potential role in oral submucous fibrosis-a precancer condition. *Carcin* 2013; 34: 1096-1104.

⁸ Kapoor S, Manpreet K, Dangi C, Singh M, Wig P, Singh H et al. Quantitative analysis of trace elements and haemoglobin as biological markers in patients with oral submucous fibrosis in Central India. *The J Pub Health* 2013; 115: 147-58.

⁹ Kammath V, Satelur K, Komali Y. Biochemical markers in oral submucous fibrosis: a review and update. *Dent Res J* 2013; 10: 576-83.

¹⁰ Ni W, Tsai C, Yang S, Chang Yu. Elevated expression of NF-kB in oral submucous fibrosis-evidence for NF-kB induction by saffrole in human buccal mucosal fibroblasts. *Oral Oncol* 2007; 43: 557-562.

¹¹ Tilakaratne W, Klinikowski M, Saku T, Peters T, Warnakulasuriya S. Oral submucous fibrosis: review on aetiology and pathogenesis. *Oral Oncol* 2006; 42: 561-8.

¹² Arakeri P, Arakeri G. Dietary copper: A novel predisposing factor for oral sub mucous fibrosis? *Med Hypothesis* 2013; 80: 241-3.

¹³ Sabharwal R, Gupta S, Kapoor K, Puri A, Rajpal K, Oral Submucous Fibrosis- a review. *J Adv Med Dent Scie* 2013; 1: 29-37.

¹⁴ Khanna S and Karjodkar F. Circulating immune complexes and trace elements (copper, iron and selenium) as markers in oral precancer and cancer: a randomised, controlled clinical trial. *Head Face Med* 2006; 2: 1-10.

¹⁵ OSMF. *J Adv Med Dent Scie* 2013; 1: 101-5.

¹⁶ Bansal S, Leekha S, Puri D. Biochemical changes in

¹⁶ Maher R, Aga P, Johnson N, Sankaranarayanan R and Warnakulasuriya S. Evaluation of multiple micronutrients supplementation in the management of oral submucous fibrosis in Karachi, Pakistan. *Nutr Cancer* 1997; 27(1): 41-47.

¹⁷ Kode M, Karjodkar F. Estimation of the serum and the salivary trace elements in osmf patients. *J Clinc Diag Res* 2013; 7(6): 1215-18.

¹⁸ Rupak S, Baby G, Padiyath S, Kumar K. Oral submucous fibrosis and iron deficiency relationship revisited- results from indian study. *E-J Dent* 2012; 2(2): 159-165.

¹⁹ Shetty S, Babu S, Kumari S, Shetty P, Hegde S, Catelino R. Status of salivary iron in oral cancer and oral potentially malignant disorders. *J Cranio Maxil Disease* 2014; 3: 17-20.

²⁰ Hosthor S, Mahesh P, Priya S, Sharada P, Jyotsna M, Chitra S. Quantitative analysis of serum levels of trace elements in patients with oral submucous fibrosis and oral squamous cell carcinoma: a randomized cross-sectional study. 2014; 18: 46-51.

²¹ Desai V, Kumar M, Bathi R. Gaurav I, Sharma R. Molecular analysis of trace elements in oral submucous fibrosis and future perspectives. *Universal Res J Dent* 2014; 4: 26-35.

²² Ray G, Ghosh R, Mallick D, Swain N, Gandhi P, Ram S et al. Correlation of trace elemental profiles in blood samples of Indian patients with leukoplakia and oral submucous fibrosis. *Biol Trace Elem Res* 2011; 144: 295-305.

²³ Neethi H, Patil S, Rao R. Estimation of serum copper and zinc levels in oral submucous fibrosis: an atomic absorption spectroscopic study. *JCDP* 2013; 14: 801-5.

²⁴ Khanna S, Udas A, Kumar G, Suvarna S, Karjodkar F. Trace elements (copper, zinc, selenium and molybdenum) as markers in oral sub mucous fibrosis and oral squamous cell carcinoma. *J Trac Elem Med Biol* 2013; 27: 307-11.

²⁵ Nayak A, Chatra L, Shenai P. Analysis of Cu and Zn levels in the mucosal tissue and serum of oral submucous fibrosis patients. *World J Dent* 2010; 1: 75-8

²⁶ Charalabopoulos K, Kotsalos A, Batistatou A, Charalabopoulos A, Vezyraki P, Peschos D et al. Serum and neoplastic tissue in breast cancer: correlation with CEA. *Br J Cancer* 2006; 95: 674-6.

²⁷ Pourmand G, Salem S, Moradi K, Nikoobakht M, Tajik P, Mehraei A. Serum Se level and prostate cancer: a case-control study. *Nutr Cancer* 2008; 60: 171-6.

²⁸ Charalabopoulos K, Kotsalos A, Karkabounas S, Vezyraki P, Kalfakakou V, Metsios A, et al. Low Se levels in serum and increased concentration in neoplastic tissues in patients with colorectal cancer: correlation with serum carcinoembryonic antigen. *Scand J Gastroenterol*, 2006; 41:359-60.

²⁹ Charalabopoulos K, Kotsalos A, Batistatou A, Charalabopoulos A., Peschos D, Vezyraki P et al. Serum and tissue Se levels in gastric patients and correlation with CEA.. *Anticancer Res* 2009; 29: 3465-7.

³⁰ Mates J, Segura J, Alonso J, Marquez J. Oxidative stress in apoptosis and cancer: an update. *Arch Toxicol* 2012; 11: 1649-65.

³¹ Aravindh L, Jagathesh P, Shanmugam S, Sarkar S, Kumar P, Ramasubramanian. Estimation of plasma antioxidants beta carotene, vitamin C and vitamin E levels in patients with OSMF and Oral Cancer - Indian population. *Int J Biol Med Res.* 2012; 3: 1655-7.

³² Aggarwal A, Shetti A, Keluskar V and Bagewadi A. Estimation of serum beta carotene levels in patients with oral submucous fibrosis in India. *J Oral Scie* 2011; 53: 427-31.

³³ Guruprasad R, Preeti P, Nair P, Manika Singh M, Singh M, Jain A. Serum vitamin C and iron levels in oral submucous fibrosis. *Indian J Dent.* 2013: 1-5.

³⁴ Gurudath S, Ganapathy KS, Sujatha D, Pai A, Ballal S, Asha ML. Estimation of superoxide dismutase and glutathione peroxidase in oral submucous fibrosis, oral leukoplakia and oral cancer-a comparative study. *Asian Pacific J Cancer Prev* 2012; 13.

³⁵ Gupta S, Reddy MV, Harinath BC. Role of oxidative stress and antioxidants in aetiopathogenesis and management of oral submucous fibrosis. *Indian J Clin Biochem* 2004; 19: 138-41.

³⁶ Metkari SB, Tupkari JV, Baroande SR. An estimation of serum malondialdehyde, superoxide dismutase and vitamin A in oral submucous fibrosis and its clinicopathologic correlation. *J Oral Maxillofac Pathol* 2007; 11:23-7.

³⁷ Ptidar KA, Parwani RN, Wanjari SP. Correlation of salivary

and serum IgG, IgA levels with total protein in oral submucous fibrosis. *J Oral Sci* 2011; 53: 97-102.

³⁸ Trinidad T, Kurilich A, Mallillin A, Walczyk T, Sagum R, Singh R.. Iron absorption from NaFeEDTA-fortified oat beverages with or without added vitamin C. *Int J Food Sci Nutr* 2013: 1-5.

³⁹ Lee PH, Chang MC, Chang WH, Wang TM, Wang YJ, et al. Prolonged exposure to arecoline arrested human KB epithelial cell growth: regulatory mechanisms of cell cycle and apoptosis. *Toxicology* 2006; 220: 81-89.

⁴⁰ Ji WT, Yang SR, Chen JY, Cheng YP, Lee YR, et al. Arecoline downregulates levels of p21 and p27 through the reactive oxygen species / mTOR complex 1 pathway and may contribute to oral squamous cell carcinoma. *Cancer Sci* 2012; 103: 1221-29.

⁴¹ Ekanayaka R, Tilakaratne W. Oral submucous fibrosis: review on mechanisms of pathogenesis and malignant transformation. *J Carcinogene Mutagene* 2013; 42: 1-11.

⁴² Teh MT, Tilakaratne WM, Chaplin T, Young BD, Ariyawardana A, et al. Fingerprinting genomic instability in oral submucous fibrosis. *J Oral Pathol Med* 2008; 37: 430-6.

⁴³ Angadi PV, Krishnapillai R. Evaluation of PTEN immunoeexpression in oral submucous fibrosis: role in pathogenesis and malignant transformation. *Head Neck Pathol* 2012; 6: 314-321.

⁴⁴ Karwan M, Veronika J, Karen S, Daniel M, Paul W et al. Betel-derived alkaloid up-regulates keratinocyte alpha v beta 6 integrin expression and promotes oral submucous fibrosis. *J Pathol* 2011; 223: 366-77.

⁴⁵ Guo F, Jian XC, Zhou SH, Li N, Hu YJ et al. A retrospective study of oral squamous cell carcinomas originated from oral submucous fibrosis. *Zhonghua Kou Qiang Yi Xue Za Zhi* 2011; 46: 494-97.

⁴⁶ Li N, Hu Q, Jiang C, Hu Y, Yuan Y, et al. Novel genetic biomarkers for susceptibility to oral submucous fibrosis: cytochrome P450 3A. *Med Hypotheses* 2011; 77: 834-6.

⁴⁷ Xie H, Liu J, Ling TY. Expression of cytochrome P450 related genes in oral submucous fibrosis tissue. *Zhonghua Kou Qiang Yi Xue Za Zhi* 2012; 47: 743-7.

⁴⁸ Bhowmik M, Roychoudhury P, Mukhopadhyay K, Ray JG et al. Association of XRCC1, XRCC3, and NAT2 polymorphisms with the risk of oral submucous fibrosis among eastern Indian population. *J Oral Pathol Med* 2012; 41: 292-302.