Original Article

Bacteria Isolated from Urinary Tract Infection among Patients and Determination of the Antibiotic Susceptibility Patterns of the Gram Negative Bacteria in Iran

Goli Angoti¹, Hossein Goudarzi^{2*}, Maryam Hajizadeh³, Zahra Tabatabaii⁴

¹ Department of Microbiology, Shahid Beheshti University, M.C International Branch, Tehran, Iran

² Department of Microbiology, School of Medicine, Shahid Beheshti University of Medical Science, Tehran, Iran

³ Department of Microbiology, Imam Reza Hospital, Tabriz, Iran

⁴ Department of Microbiology, Faculty of Medical Sciences, Tabriz University of Medical Science, Tabriz, Iran Received: 15 April, 2015; Accepted: 5 October, 2015

Abstract

Background: *Escherichia coli* (*E. coli*) is the most frequent infecting organism in acute infection. So, knowledge about the frequency and distribution of urinary tract infection (UTI) is important to improve infection control measures. The aim of this research was to determine the prevalence of bacteria isolated from urinary tract infection (UTI) in patients and determination of the antibiotic susceptibility patterns of the gram negative bacteria.

Materials and Methods: This descriptive study was performed in Imam Reza hospital, Tabriz (northwest of Iran) during March 2012 to February 2013. We surveyed 8153 patients, who had clinical manifestations of UTI. 5093 (62.47%) of them were female and 3060 (37.53%) of them male. Urine specimens were cultured for isolation of the microbial agents of UTI. The isolated bacteria were identified using biochemical tests. Disk diffusion susceptibility test was used to determine antimicrobial susceptibility.

Results: *E. coli* (55.38%) was the most common isolated pathogen, followed by *Enterobacter spp.* (29.61%), *Pseudomonas spp.* (4.9%), *S. aureus* (3.21%), *Enterococcus spp.* (2.3%), *fungi* (1.5%) and *Klebsiella* (0.48%). The sensitivity rates of isolated gram negative bacteria were for Amikacin (95.7%), Nitrofurantoin (91.5%), Gentamicin (64.1%), Ceftizoxim (56.8%), Ciprofloxacin (37.6%), Cotrimoxazole (31.4%) and Nalidixic acid (23.5%).

Conclusion: This study showed that the frequency of *E. coli* and *Enterobacter spp.* increases the probability of urinary tract infection. Also this survey indicates the emergence of antibiotic resistant infections in the studied hospital. So, there is a need to improve the effectiveness of integrated infection control programs to control and manage nosocomial infections caused by highly resistant organisms.

Keywords: Prevalence of bacteria, Urinary tract infection, antibiotic susceptibility

*Corresponding Author: Hossein Goudarzi, Department of Microbiology, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran. Tel: (+98) 21 23872556. Email: hgod500@yahoo.com

Please cite this article as: Angoti G, Goudarzi H, Hajizadeh M, Tabatabaii Z. Bacteria Isolated from Urinary Tract Infection among Patients and Determination of the Antibiotic Susceptibility Patterns of the Gram Negative Bacteria in Iran. Novel Biomed. 2016;4(1):1-4.

Introduction

Urinary tract infection (UTI) is one of the most

prevalent infections. UTI is classified in to uncomplicated and complicated infections with respect to choices for treatment¹. Among patients referred to physicians, *Escherichia coli* (*E. coli*) is the most common etiological agent, approximately isolated from 75 to 90% of uncomplicated patients, while complicated ones show a broader bacterial spectrum as the cause of infection¹⁻³.

In comparison with men, UTI is reported more in women. It could be due to the proximity of genital tract and urethra/anus⁴ and anatomical predisposition urothelial mucosa adherence or to mucopolysaccharides lining or other host factors⁵. Urinary tract infection may involve only the lower urinary tract or both the upper and the lower tracts. The term cystitis has been used to describe the syndrome involving dysuria, frequency, and occasionally supra pubic tenderness.

More than 95% of urinary tract infections are caused by a single bacterial species. *E. coli* is the most frequent infecting organism in acute infection^{6,7}. *Enterobacter, Staphylococci, Klebsiella, Proteus, Pseudomonas,* and *Enterococci species* are more often isolated from inpatients, whereas there is a greater preponderance of *E. coli* in an outpatient population⁸. Hence, the aim of this study was to isolate pathogenic agents involving UTI and to determine their antibiotic susceptibility pattern among patients referred to Imam Reza hospital, Tabriz, Iran.

Methods

Sampling and identification of isolates: A total, 8153 patients with UTI referred to Imam Reza hospital laboratory located in Tabriz city, East Azerbaijan province, northwest of Iran, during March 2012 to February 2013. There were 5093 (62.47%) females and 3060 (37.53%) males, with an age range of 25-80 years (mean, 53.8 years). Urine samples were obtained from a midstream into standardized, sterile container and delivered to the laboratory within 2 hours. Identification was done based on culture characteristics, gram stain and routine standard biochemical tests⁹⁻¹¹. Guidelines for proper specimen collection were given to all patients on a printed card¹⁰.

Colony count of bacteria in UTI: A measured amount of urine, using calibrated loop method was inoculated on Blood agar medium (Merck, Germany) for colony count. A specimen was considered positive for UTI if a single organism was cultured at a concentration of $\ge 10^5$ cfu/ml, or when a single organism was cultured at a concentration of 10^4 cfu/ml while ≥ 5 leukocytes per high-power field were observed on microscopic examination of the urine¹². Urine specimens were cultured for isolation of the microbial agents of UTI on Blood agar and MacConky agar media (Merck, Germany).

Antimicrobial susceptibility testing: Antibiotic susceptibility test was carried out on Mueller Hinton agar (Merck, Germany) by Kirby-Bauer's disk diffusion method according interpretive criteria recommended by the clinical and laboratory standards institute (CLSI) guidelines to the fallowing agents¹³: Amikacin (AN 30µg), Nitofurantoin (FM 300µg), Gentamicin (GM 10µg), Ceftizoxim (CT 30µg), Ciprofleoxacin (Cp 5 g), Trimethoprimsulfamethoxazole (25µg), Nalidixic acid (NA 30µg), and Ampicillin (AM 10 µg). Antibiotic disks used in this research were supplied by Padtan Teb, Tehran, Iran¹⁰. Appropriate antibiotic discs were tested depending upon whether the organism was gram positive or gram negative. Interpretation of results was done based on the diameter of the zone. E. coli ATCC 25922 was used as quality control for antimicrobial susceptibility test.

Statistical analysis: The results were analyzed using SPSS software vs.16 and presented with 95% confident intervals (CI).

Results

8153 patients with clinical symptoms of UTI in this

Table 1: Frequency of bacterial agents isolated fr	om
urine specimens in this study.	

Organisms	No. of isolates (%)
E. coli	1126 (55.38%)
Entrobacter spp.	602 (29.61%)
Pseudomonas aeruginosa	98 (4.9%)
Staphylococcus aureus	65 (3.21%)
Enterococcus spp.	46 (2.3%)
Coagulase-Negative staphylococci ¹	42 (2.06)
Fungi	30 (1.5%)
Klebsiella spp.	11(0.48%)
Proteus spp.	11 (0.54%)
serratia	4 (0.2%)

Antimicrobial resistance trait of isolates	No. (%)
Amikacin	1949 (95.7)
Nitofurantoin	1862(91.5)
Gentamicin	1304 (64.1)
Ceftizoxim	1155 (56.8)
Ciprofloxacin	765 (37.6)
Trimethoprimsulfamethoxazole	639 (31.4)
Nalidixic acid	478 (23.5)
Tetracycline	407 (20)
Ampicillin	40(2)

Table 2: Antimicrobial sensitivity pattern of bacterial agents isolated from urine specimens^a.

a Shown are the numbers and percentages of isolates exhibiting sensitive to the tested antimicrobials.

retrospectively study were investigated (62.47% of them were female and 37.53% were male). 2035 (24.96%) of cases had positive urine culture and 6118 (75.04%) ones did not have significant bacteriuria or bacterial count of their urine samples were very low. Ten types of microorganisms were isolated from positive urine cultures. The most common isolates were *E. coli* (55.38%), followed by *Entrobacter spp.* (29.61%), *Pseudomonas spp.* (4.9%), *S. aureus* (3.21%), (Table 1). The most common isolated uropathogens in Gram-negative bacilli and Gram- positive cocci were *E. coli* (55.3%) and *S. aureus* (3.21%) respectively.

Analysis of the antimicrobial susceptibility profile of the isolates showed that most of isolate (95.7%) were susceptible to Amikacin. This isolate showed resistance to other tested antibiotics. Of 2035 isolates, 98% of the isolates were resistant to Amoxicillin and Ampicillin (n=1995), 80% were resistant to Tetracycline (n=1628), 76.5% were resistant to Nalidixic acid (n=1557) and 62.4% were resistant to Ciprofloxacin (n=1270) (Table 2).

Discussion

Bacterial infection of the urinary tract is one of the common causes for seeking medical attention in the community¹⁴. Effective management of patients suffering from bacterial UTIs commonly relays on the identification of the type of organisms that caused the disease and the selection of an effective antibiotic agent to the organism in question¹⁵. In this study, the

isolation rate of bacteria from urine was 24.96% which is comparatively lower than reports within the country and other part of the world^{16,17}, this might have been either due to sample size variation or the studies might have been based on retrospective survey. However, this finding is in line with studies done in Addis Ababa¹⁸ and one from Iran which had a rate of $13.2\%^{19}$.

E. coli is the major etiological agent in causing UTI, which accounts for up to 90% of cases²⁰. In this study, the most frequent UTI were Gram negatives which made up 80% of all the isolates. *E. coli* is the predominant etiology of UTI, in both outpatients and inpatients of both sexes, and this finding is in agreement with others finding too^{16,17,19,21}.

Resistance to antimicrobial agents has been noted since the first use of these agents and is an increasing world-wide problem²². This study revealed that a higher prevalence rate of resistance to the commonly prescribed antibiotic agent. The finding that 98% of E. coli and K. pneumonia isolates were resistant to Amoxicillin and Ampicillin is of great importance and implies that these antibiotics cannot be used as empirical therapy for urinary tract infection particularly in the study area. On the other hand, very low levels of resistance were detected to antibiotics such as Nitrofurantoin and Gentamicin and a comparable rate of sensitivity has been reported for these drugs in previous studies done in Ethiopia^{16,18,23,24}, in Kosovo¹⁷, in Iran¹⁹. Nitrofurantoin represented better activity against E. coli isolates, but this drug would not be recommended for serious upper urinary tract infections or for those cases with systemic involvement²⁵. Low resistance was observed for these drugs because they are not easily accessible and relatively expensive in price compared to others. Thus, these drugs could be considered as alternative options in the empirical treatment of UTIs.

Conclusion

In conclusion, the isolation of bacterial Urinary tract infection with a higher resistance rates for commonly used antimicrobials leaves the clinicians with very few options to choose drug used for empirical treatment of UTIs. Therefore, it is important to urge physician and other health worker in the field on the need of reevaluation of empiric treatment of UTI. As drug resistance among pathogens is an evolving process, so there is a need to improve the effectiveness of integrated infection control programs to control and manage nosocomial infections caused by highly resistant organisms.

Acknowledgment

This study was supported by a grant from Imam Reza hospital in Tabriz, Iran and School of Medicine, Shahid Beheshti University of Medical Sciences.

References

1. Sherifa M, Moataz M. Epidemiological and microbiological profile of nosocomial infection in Taif hospitals, KSA (2010-2011). World Journal of Medical Sciences. 2012;7(1):1-9.

2.Gupta K, Hooton TM, Stamm WE. Increasing antimicrobial resistance and the management of uncomplicated community-acquired urinary tract infections. Annals of internal medicine, 2001;135(1):41-50.

3. EN L. Epidemiology of urinary tract infection. Infect Med. 2001;18:153-62.

4.Schaeffer AJ, Rncq BE, Anderson DL, Pruden J, Duncan JL. Host pathogenesis in urinary tract infection. International journal of antimicrobial agents. 2001;17:245-51.

5.Akortha EE. Incidence and antibiotic susceptibility pattern of Staphylococcus aureus amongst patients with urinary tract infection (UTI) in UBTH Benin City, Nigeria. African Journal Biotechnology. 2008;7:1637-40.

6. Ronald A. The etiology of urinary tract infection: Traditional and emerging pathogens. Am J Med. 2002;113:14S-9S.

7. Jellheden B, Norrby RS, Sandberg T. Symptomatic urinary tract infection in women in primary health care: Bacteriological, clinical and diagnostic aspects in relation to host response to infection. Scand J Prim Health Care. 1996;14:122-8.

8. Bronsema DA, Adams JR, Pallares R, et al. Secular trends in rates and etiology of nosocomial urinary tract infections at a university hospital. J Urol. 1993;150:414-6.

9. Mandell GL, Bennett JE Dolin R. Principles and practice of infectious diseases. Churchill Livingstone. 2005;881-2.

10. Forbes BA. Sahm DF, Weissfeld AS. Bailey and Scott's Diagnostic microbiology, 12th edition, Mosby Elsevier, 2007;842-55.

11. MacFaddin JF. Biochemical tests for identification of medical bacteria. 3rd ed. Philadelphia: Lippincott Williams and Wilkins.

2000.

12. Schneider PF, Riley TV. Staphylococcus saprophyticus urinary tract infections: Epidemiological data from Western Australia. Eur J Epidemiol. 1996;12:51-4.

13. Clinical and laboratory standards institute. Performance standards for antimicrobial susceptibility testing. Twenty-Second Informational supplement. CLSI document M100-S22. Pennsylvania, USA. 2012;32:3.

14. Kebira AN, Ochola P, Khamadi SA. Isolation and antimicrobial susceptibility testing of Escherichia coli causing urinary tract infections. J. Appl. Biosci. 2009;1320-5.

15. Water G, Harrison B, and Kunin G. Urinary tract infection. N. Eng Med. J 1996;248-50.

16. Tessema B, Kassu A, Mulu A, Yismaw G. Predominant Isolates of Urinary Tract Pathogens and their susceptibility Patterns in Gonder Univesity Teaching Hospital, Northwest Ethiopia. Ethio Med J. 2007;45:61-7.

17. Rakaa L, Mulliqi-Osmani G, Berisha L, et al. Etiology and susceptibility of urinary tract isolates in Kosova. Int J Antimicrob agents. 2004;23S1S2–S5.

18. Assefa A, Asrat D, Woldeamanuel Y, Hiwot Y, Abdella A, Melesse T. Bacterial profile and drug susceptibility pattern of urinary tract infection in pregnant women at Tikur Anbessa Specialized Hospital Addis Ababa, Ethiopia. Ethiop Med J. 2008;46:227-35.

19. Farajnia S, Alikhani MY, Ghotaslou R, Naghili B, Nakhlband A. Causative agents and antimicrobial susceptibilities of urinary tract infections in the northwest of Iran. Int J Infect Dis. 2009;13:140-4.

20. Ronald A. The etiology of urinary tract infection: Traditional and Emerging Pathogens. Am J Med 2002; 113 Suppl 1A:14S-19S.

21. Dromigny JA, Nabeth P, Perrier Gros Claude JD. Distribution and susceptibility of bacterial urinary tract infections in Dakar, Senegal. Int J Antimicrob Agents. 2002;20:339-47.

22. Sefton AM. The impact of resistance on the management of urinary tract infections. Int J Antimicrob Agents. 2000;16:489–91.

23. Moges F, Genetu A. Antibiotic sensitivity of common bacterial pathogens in urinary tract infections at Gonder Hospital, Ethiopia. East Afr Med J. 2002;79:140-2.

24. Wolday D, Erge W. Increased incidence of resistance to antimicrobial by urinary pathogens isolated at Tikur Anbessa Hospital. Ethiop Med J. 1997;35:127-35.

25. Vasquez Y, HW A. Antibiotic susceptibility patterns of community-acquired urinary tract isolates from female patients on the US (Texas)-Mexico border. The Journal of Applied Research. 2004;4:321-6.

Biadglegne F, Abera B. Antimicrobial resistance of bacterial isolates from urinary tract infections at Felge Hiwot Referral Hospital, Ethiopia. Ethiop. J. Health Dev. 2009;23:236-8.