Scientific Report

**Isolation and identification of Helicobacter pullorum from caecal content of broiler chickens in Mashhad, Iran**

Jamshidi, A.¹; Bassami, M. R.²; Salami, H.³ and Mohammadi, S.³

¹Department of Food Hygiene and Public Health, Faculty of Veterinary Medicine, Ferdowsi University of Mashhad, Mashhad, Iran; ²Department of Clinical Sciences, Faculty of Veterinary Medicine, Ferdowsi University of Mashhad, Mashhad, Iran, and Institute of Biotechnology, Ferdowsi University of Mashhad, Mashhad, Iran; ³Graduated from Faculty of Veterinary Medicine, Ferdowsi University of Mashhad, Mashhad, Iran

*Correspondence: A. Jamshidi, Department of Food Hygiene and Public Health, Faculty of Veterinary Medicine, Ferdowsi University of Mashhad, Mashhad, Iran. E-mail: ajamshidi@ferdowsi.um.ac.ir

(Received 4 Nov 2012; revised version 30 Jan 2014; accepted 19 Feb 2014)

**Summary**

The presence of *Helicobacter pullorum* in intestinal tract of broiler chickens may be a potential risk for human health. In this study, a total of 100 caecal samples of broilers carcasses from 20 flocks at a poultry abattoir in Mashhad suburb were tested for the presence of *H. pullorum* using modified conventional culture method by combination of culturing on Brucella sheep blood agar and a filtering technique. Suspected colonies were determined as *H. pullorum* using polymerase chain reaction (PCR) by amplifying a 447 bp fragment of the 16S rRNA gene of this bacteria. 41% of caecal content samples and samples from 12 broiler flocks (60%) were determined as positive for the presence of *H. pullorum*. This is the first report of *H. pullorum* in Iranian poultry flocks. The results showed high prevalence of this bacterium in broiler chickens in this area of Iran. It seems using combination of conventional culture method and PCR assay based on amplification from conserved genes allows reliable detection and identification of *H. pullorum*.

**Key words:** Helicobacter pullorum, Broiler, PCR

**Introduction**

Since the identification of *Helicobacter pylori* in 1984, some more *Helicobacter* species have been described (Solnick and Schauer, 2001). Most of these species appear to be hosted by one or more animals and enzootic infection is evident in most geographic regions (Solnick, 2003).

Whereas the *H. pylori* is well characterized as a severe gastric pathogen, the pathogenic potential of the other species within the genus is not known (Tee et al., 2001).

*Helicobacter pullorum* was classified as a new species of *Helicobacter* by Stanley et al. (1994) on the basis of 16S rRNA sequence data. This organism has been isolated from the livers and intestinal contents of laying hens with vibrionic hepatitis and also from the caeca of broiler chickens (Stanley et al., 1994; Atabay et al., 1998). In human, the organism has been isolated from gastroenteritis, diarrhoea, and liver and gall bladder diseases (Stanley et al., 1994). It has been suggested that *H. pullorum* may play a role in Crohn’s disease (Bohr et al., 2002). The genomic DNA has also been detected in the livers from patients with primary sclerosing cholangitis, cirrhosis and hepatocellular carcinoma (Rocha et al., 2005), although it was unclear if the organism had a causal role in these infections (Gibson et al., 1999).

It appears that the number of *H. pullorum* infections in human has most probably been underestimated because of the phenotypic similarities between the genera *Helicobacter* and *Campylobacter* on one hand and the specific isolation requirements of *H. pullorum* on the other hand (Atabay et al., 1998; Gibson et al., 1999). Therefore, a significant number of patients with diarrhoea may have been misdiagnosed in the past (Atabay et al., 1998).

Poultry carcasses can be contaminated by *H. pullorum* during slaughtering (Atabay et al., 1998). The preferred colonization site is the caecum wherein the bacterium shows close association with the surface epithelium (Ceelen et al., 2006).

There is no superior method for isolation of all *Helicobacter* species, therefore a combination of phenotypic and genotypic methodologies has been recommended for identifying the organism (On et al., 1996; On et al., 2005).

The aim of this study was to investigate the occurrence of *H. pullorum* in caecal contents of broiler chickens, collected from a slaughterhouse in Mashhad, Iran, employing combination of a modified conventional culture method and a PCR assay.

**Materials and Methods**

**Sampling**

During the summer of 2011, a total of 100 samples were randomly collected from gastrointestinal tract of...
broiler chickens, belonging to 20 broiler flocks (5 samples from each flock), after the evisceration stage in the processing line at a commercial broiler slaughtering facility in Mashhad. The complete intestinal tract from each bird was obtained and packed into a separate sterile plastic bag. The samples were brought to the laboratory on ice and examined within 4 h after sampling.

Conventional culture method
In the laboratory, the caeca were aseptically severed and their surfaces were decontaminated using ethyl alcohol. Approximately 5 g of caecal contents were squeezed into 5 ml of sterile saline and vortexed to obtain a homogenous suspension. An aliquot of 200 μl of each sample was diluted in 400 μl of a sterile mixture containing 7.5 g glucose, 25 mL brain heart infusion broth (Merck, Germany), and 75 mL sterile inactivated horse serum, and then homogenized (Ceeelen et al., 2006).

Samples were then inoculated on Brucella agar (Merck, Germany) supplemented with 5% sheep blood using the modified filter technique of Steele and McDermott (1984). Briefly, a sterile cellulose acetate membrane filter (0.45 μm) was applied with a sterile pair of tweezers directly onto the surface of the agar.

When the filter was totally absorbed on the agar, 300 μl of the mixture was placed in the middle of the filter. The inoculated plates were incubated overnight at 37°C for 1 h in a microaerobic atmosphere. After at least 1 h of incubation the filter was removed with a sterile pair of tweezers and the filtrate was streaked on the agar with a loop. Incubation was conducted in microaerobic conditions (10% CO₂, 5% O₂, and 85% N₂) at 37°C for a week and examined daily for growth. Approximately five small, grayish-white colonies of Gram-negative, gently curved, slender rod bacteria were pooled and subcultured from each plate on sheep blood agar.

DNA extraction
Suspected colonies which were subcultured on sheep blood agar were collected and suspended in 500 μl of sterile, deionized distilled water and heated in a boiling water bath for 10 min. The samples were cooled immediately on ice for 5-10 min and centrifuged at 13,000 × g for 5 min. The supernatants were used as DNA template for PCR assay.

The PCR assay
A PCR assay amplifying a 447 bp fragment of the 16S rRNA of H. pullorum was used (Stanley et al., 1994). To confirm the isolates identity, the reaction mixture consisted of 2.5 μl of bacterial lysate, 2.5 μl of 10 × BSA buffer (1 ml of 10 × contained 500 μl of 1 M Tris-HCl, pH = 8.5, 200 μl of 1 M KCl, 30 μl of 1 M MgCl₂, 5 mg of BSA and 270 μl of deionized water), 2.4 μl of 10 × dNTP mixture (2.5 mM of each dNTP), 0.7 μl of each primer (100 pmoles/μl), 0.2 μl of Taq DNA polymerase (5 U/μl) and deionized water to a final volume of 25 μl. According to Stanley et al. (1994), the sequences of used primers were as follow: 5’ ATG AAT GCT AGT TGT TGT CAG 3’ and 5’ GAT TGG CTC CAC TTC ACA 3’. The conditions used for the amplifications were as follow: an initial denaturation at 94°C for 5 min, followed by 35 cycles of denaturation at 94°C for 1 min, annealing at 60°C for 90 s, elongation at 72°C for 90 s, and a final elongation at 72°C for 5 min. The PCR products were separated by electrophoresis in 1.5% agarose gel at 100 V for 40 min in Tris-acetate buffer, visualized by ethidium bromide staining, illuminated by UV-transilluminator and documented by a gel documentation apparatus. A 100 bp DNA ladder (fermentas) was used as a size reference for PCR assay.

For positive control, the first positive sample in PCR assay with a molecular weight of 447 bp was purified and DNA sequence analysis of the amplicon was performed. By the use of BLAST software, the sequence was compared to published data for H. pullorum 16S rRNA sequences in GenBank (accession No. AY631956, L36143, and L36144). Confirmed isolate was used as positive control through the experiment and sterile distilled water was used as negative control.

Individual and herd level prevalence and 95% confidence interval (CI) for H. pullorum was calculated.

Results
In total 100 samples from caecal contents of poultry carcasses, representing 20 broiler flocks were analysed by conventional culture method using a modified filter technique. In 85 out of 100 samples suspected colonies appeared on sheep blood agar.

In PCR assay using specific primers of the 16SrRNA gene of H. pullorum, the expected amplicon of 447 bp was produced in 41 suspected colonies, representing 41% of all 100 samples examined (Fig. 1). In total, 12 flocks out of 20 flocks, representing 60% were confirmed to be carrier of (or contaminated by) H. pullorum.

![Fig. 1: Detection of H. pullorum in samples of caecal content of broilers carcasses by PCR assay, amplifying 447 bp segment of 16SrRNA gene. Lane 1: Negative control, Lane 2: 100 plus bp markers, Lane 3: Positive control and Lanes 4-9: Positive samples](image-url)

Individual level prevalence of H. pullorum was 41%
was detected in experimentally, P-in broiler farms of the 100% and France (100%) has been reported (Pilon et al., 2005; Zanoni et al., 2007). Atabay et al. (1998) reported the occurrence of H. pullorum in 60% of poultry carcasses in the UK and more recently Ceelen et al. (2006), reported the presence of H. pullorum in 33.6% of cecal content of broiler carcasses and in 64% of broiler flocks from Belgium. In Egypt H. pullorum was identified in 39.33% of chickens caecal droppings tested (Moemen 1997,). To our 2006.Stanley 2002 tion with (Ceelen et al., 2005), but it is not clear whether H. pullorum in humans is acquired by consumption of undercooked poultry meat, as is the case with Campylobacter jejuni (Humphrey et al., 2007; Verhoef-Bakkenes et al., 2008). By the way, retail raw poultry meats and other poultry products may constitute vehicles for human H. pullorum infections through consumption of the contaminated carcasses (Antolin et al., 2001). In this study we used PCR method for final confirmation of suspected colonies, using primers, designed by Stanley et al. (1994) that target a 447 bp fragment of the H. pullorum 16SrDNA gene. Although in general, a concordance between different detection techniques for H. pullorum has been reported (Ceelen et al., 2007), a combination of conventional culture and PCR assay seems to increase the chance of H. pullorum identification. Concerning health monitoring, PCR may be helpful in detecting this pathogen not only in intestinal tissue but also in broiler chicken cecal droppings (Ceelen et al., 2006).

Discussion

In this study, H. pullorum was detected in 60% of broiler flocks and 41% of broiler carcasses. Our results point out a high prevalence, although it may not represent a true prevalence of H. pullorum in broiler farms of the region under investigation. Higher prevalence of H. pullorum in broiler farms in Italy (100%) and France (100%) has been reported (Pilon et al., 2005; Zanoni et al., 2007). Atabay et al. (1998) reported the occurrence of H. pullorum in 60% of poultry carcasses in the UK and more recently Ceelen et al. (2006), reported the presence of H. pullorum in 33.6% of cecal content of broiler carcasses and in 64% of broiler flocks from Belgium. In Egypt H. pullorum was identified in 39.33% of chickens caecal droppings tested (Moemen 1997,). To our 2006. Stanley 2002. tion with (Ceelen et al., 2005), but it is not clear whether H. pullorum in humans is acquired by consumption of undercooked poultry meat, as is the case with Campylobacter jejuni (Humphrey et al., 2007; Verhoef-Bakkenes et al., 2008). By the way, retail raw poultry meats and other poultry products may constitute vehicles for human H. pullorum infections through consumption of the contaminated carcasses (Antolin et al., 2001). In this study we used PCR method for final confirmation of suspected colonies, using primers, designed by Stanley et al. (1994) that target a 447 bp fragment of the H. pullorum 16SrDNA gene. Although in general, a concordance between different detection techniques for H. pullorum has been reported (Ceelen et al., 2007), a combination of conventional culture and PCR assay seems to increase the chance of H. pullorum identification. Concerning health monitoring, PCR may be helpful in detecting this pathogen not only in intestinal tissue but also in broiler chicken cecal droppings (Ceelen et al., 2006).

In conclusion, the results of this experiment demonstrate the presence of H. pullorum in caecum content of broiler chicken at a high prevalence, possessing a potential risk for human population. To our knowledge, this is the first report of H. pullorum in Iranian poultry flocks. The results may be considered as a base in larger scale studies for estimating the true prevalence.

Acknowledgements

The authors thank the abattoir Tiuran (Mashhad, Iran) for providing intestinal tracts, and Dr. H. HeidariFard for his generous assistance in sampling procedure.

References


Bohr, URM; Primus, A; Zagoura, A; Glasbrenner, B; Wex, T and Malfertheiner, P (2002). A group-specific PCR assay for the detection of Helicobacter acese in human gut.
Helicobacter. 7: 378-383.


Rocha, M; Avenaud, P; Menard, A; Le Bail, B; Balabaud, C; Bioulaec-Sage, P; de Magalhaes Queiroz, DM and Megraud, F (2005). Association of Helicobacter species with hepatitis C cirrhosis with or without hepatocellular carcinoma. Gut., 54: 396-401.


Stanley, J; Linton, D; Burnens, AP; Dewhirst, FE; On, SLW; Porter, A; Owen, RJ and Costas, M (1994). Helicobacter pullorum sp. nov. genotype and phenotype of new species isolated from poultry and from human patients with gastroenteritis. Microbiology, 140: 3441-3449.


