The Relationship between -2548 G/A Leptin Gene Polymorphism and Risk of Breast Cancer and Serum Leptin Levels in Ahvazian Women

Ghorban Mohammadzadeh¹, Mohammad-Ali Ghaffari², Ahmmad Bafandeh³, Seyed-Mohammad Hosseini⁴, Behnaz Ahmadi³

Abstract

Background: Potential association of leptin (LEP) gene polymorphisms has been suggested in the processes leading to breast cancer initiation and progression. We investigated whether genetic variations in the LEP -2548G/A gene are associated with risk of breast cancer.

Methods: This case-control study consisted of 100 breast cancer cases and 100 control subjects without breast cancer that matched for age and body mass index (BMI). Genotyping of LEP -2548G/A polymorphism was performed by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) assay. Serum leptin level was determined by ELISA in all study subjects.

Results: The genotype distributions (AA, AG, and GG) were 36, 55, and 9% in breast cancer cases and 52, 45, and 3% in control group, respectively. The frequency of LEP -2548 GG genotype was significantly elevated in breast cancer cases as compared to controls ($\chi 2$ =6.90, p=0.032). Similar difference was also found in allele frequencies between two groups ($\chi 2$ =5.65, p=0.017). A markedly increase risk of breast cancer was associated with the LEP -2548GG genotype when compared to the LEP -2548 AA genotype (OR=4.33, 95% CI=1.09-17.22). In addition, postmenopausal women who bear at least one LEP -2548 G allele were at a markedly increased risk of breast cancer after adjusting for age and BMI confounders (OR=12.24, 95% CI=1.13-131.73).

Conclusion: The LEP -2548 G/A polymorphism is associated with markedly increased risk of breast cancer especially in postmenopausal Ahvazian women and supported the hypothesis that leptin is involved in breast cancer.

Keywords: Brest Cancer; Leptin; polymorphism; PCR-RFLP

Please cite this article as: Mohammadzadeh Gh, Ghaffari MA, Bafandeh A, Hosseini SM, Ahmadi B. The Relationship between -2548 G/A Leptin Gene Polymorphism and Risk of Breast Cancer and Serum Leptin Levels in Ahvazian Women. Iran J Cancer Prev. 2015;8(2):100-8.

Introduction

Breast cancer is one of the commonest causes of cancer death among women worldwide, but its etiology is still not fully determined [1]. In The recent years, the incidence of breast cancer has increase dramatically in Iran, and consider as of the most frequent malignancies among Iranian women [2]. Although, it has been determined that presence less than 10% of breast cancer is due to dominant mutations in highly penetrate susceptibility genes, including BRCA1 and BRCA₂. However, lowpenetrate genes, that also increase susceptibility to breast cancer, have been suggested too [3].

1. Hyperlipidemia Research Center, Dept. of Clinical Biochemistry, Faculty of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran 2. Cellular and Molecular Research Center, Dept. of Clinical Biochemistry, Faculty of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran 3. Dept. of Clinical Biochemistry, Faculty of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran 4. Dept. of Radiation and oncology of Gholestan University Hospital, Faculty of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

Corresponding Author:

Ghorban Mohammadzadeh, Ph.D; Assistant professor of Biochemistry Tel: (+98) 06133367543-50 Email: mohammadzadeh@ajums.ac.ir Received: 11 Aug. 2014 Accepted: 22 Feb. 2015 Iran J Cancer Prev. 2015; 2:100-8.

Leptin (LEP), an adipocyte-derived satiety hormone, is thought to plays a crucial role in the regulation of energy expenditure and body weight through acting on its receptor expressed mainly in the hypothalamus [4, 5]. Moreover, studies have indicated that the leptin is involved in carcinogenesis of breast tissue and acts to favor the proliferation and angiogenesis of breast cancer cells [6-8]. Interestingly, over expression of leptin in breast cancer appears to be associated with higher tumor grade and size [9-13]. The leptin gene, the human homologue of the rat obese gene (OB), that had been cloned and sequenced by Zhang et al. [14], is located on chromosome 7q31.3 [15, 16] and the length of its mRNA in adipose tissue is 4.5 kb [14, 17].

humans, In several single nucleotide polymorphisms (SNPs) have been identified in the LEP gene. Among them, G to A substitution at nucleotide (nt) -2548 upstream of the ATG start site in the LEP gene promoter, LEP -2548G/A, has been with adipocytes increased associated leptin production secretion [18-20]. and Increased circulating levels of leptin and over gene expression have been reported in subjects carrying the LEP -2548 A allele [21]. The LEP -2548 A allele has also been associated with a two-fold increase of leptin secretion from adipocytes when compared to secretion by adipocytes bearing only the LEP -2548 G allele [21]. Herein, an association between leptin and breast cancer risk could be proposed. However, only a few studies with inconsistent results have addressed the relationship between leptin and breast cancer [10, 22-26], and to date, no study has addressed the association between polymorphisms within the LEP gene and breast cancer risk in Iranian population. Thus, in this study we investigate the associations of LEP -2548 G/A polymorphism and risk of breast cancer in Ahvazian women, a sample of Iranian population.

Materials and Methods

Study Population

A total of 200 cases were investigated in two groups. One group was comprised of One-hundred Iranian unrelated women with confirmed breast cancer recruited from the department of radiation and oncology of Golestan University Hospital, Ahwaz, Iran as previously described elsewhere (27). The diagnosis of cancer was confirmed by histopathological analyses. Clinical information such as stage of the breast cancer, menopausal status at the time of onset, hormonal receptor status (ER. PR), tumor size, and BMI was obtained from the hospital records. Menopausal status was determined using information provided by the subjects about the date of her last menstrual period. Postmenopausal status was defined as having a last menstrual period more than 6 months before the reference date. The second group was composed of 100 unrelated age and BMI matched women without any personal or family history of cancer and other serious disease to serve as controls. All the breast cancer cases and controls enrolled in the study were Iranians and informed about the study and consent was taken. This study was approved by the Clinical Research Ethics Committee. Women with suspected breast cancer without histological confirmation and those that refused sample donation were excluded from the study.

Anthropometric and other Measurements

Anthropometric indices, consisted of height, weight, body mass index (BMI), as well as a full-fasted lipid profile including total cholesterol, high density lipoprotein cholesterol (HDL-C), low density lipoprotein cholesterol (LDL-C), and triglycerides (TG) were measured as previously described [27].

Polymorphism analysis of the -2548 G/A LEP gene

For DNA extraction, blood samples were collected into K3-EDTA-treated tube from both patients and healthy controls and were stored at -20°C. Genomic DNA was isolated from peripheral blood leukocytes using the salting-out method. The LEP -2548 G/A variants were genotyped by using PCR-RFLP analysis with the forward primer 5'-TTTCCTGTAATTTTCCCATGAG-3' and reverse primer 5'-AAAGCAAAGACAGGCATAAAA-3'. Conditions for amplification were 12.5 µl master mix, 2.0 μ l (10 pmol/ μ l) forward and reverse primers, 2.0 µl (50 ng/µl) template DNA, and 6.5 µl sterile nuclease free water (18 µl). The PCR conditions were as follows: initial denaturation at 94°C for 5 minutes, 35 cycles of denaturation at 94°C for 60 seconds, annealing at 53°C for 45 seconds, and extension at 72°C for 60 seconds, with a final extension of 5 minutes at 72°C. The PCR amplified products were scored in 242 bp in a mixture reaction consisting of: PCR products (10 μ l), 10× buffer (2 μ l), 10 units *Hhal* restriction enzyme, and sterile nuclease free water (18 µl). The reaction mixture was kept overnight at 37°C for 1-16 The fragments were separated by hours. electrophoresis on 3% agarose gel, stained with ethidium bromide and results were recorded with photographs of gels in UV light. The polymorphism was defined by presence (G) or absence (A) of the Hhal restriction site.

Statistical analysis

SPSS software for Windows version 15.0 (SPSS, Inc., Chicago IL, USA) was used for the all statistical analyses. All frequencies were estimated by gene counting. The observed genotype frequencies in the breast cancer cases and controls were tested for the Hardy-Weinberg equilibrium

Genotype	Breast cancer Cases	controls	χ2	p value
All Women	(n = 100)	(n = 100)		
Genotype frequencies				
AA	36 (36.0)	52 (52.0)		
AG	55 (55.0)	45 (45.0)		
GG	9 (9.0)	3 (3.0)	6.90	0.032
AA	36 (36.0)	52 (52.0)		
AG+GG	64 (64.0)	48 (48.0)	5.19	0.023
Allele frequencies				
A	63.5	74.5		
G	36.5	25.5	5.65	0.017
Pre-menopausal				
Genotype frequencies	(n = 55)	(n = 60)		
AA	21 (38.2)	30 (50.0)		
AG	32 (58.2)	28 (46. 7)		
GG	2 (3.6)	2 (3.3)	1.64	0.440
AA	21 (38.2)	30 (50.0)		
AG+GG	34 (61.8)	30 (50.0)	1.62	0.203
Allele frequencies				
A	67.3	73.3		
G	32.7	26.7		
Post-menopausal				
Genotype frequencies	(n =45)	(n=40)		
AA	15 (33.3)	22 (55.0)		
AG	23 (51.1)	17 (42.5)		
GG	7 (15.6)	1 (2.5)	6.45	0.040
AA	15 (33.3)	22 (55.0)		
AG+GG	30 (66.7)	18 (45.0)	4.044	0.044
Allele frequencies				
A	58.9	76.3		
G	41.1	23.8		

Table 1. Genotype and allele frequencies of LEP -2548G/A polymorphism in breast cancer cases and controls

Data are presented as number of cases with frequency in parentheses. Allele frequencies and genotype distribution of the LEP -2548G/A in breast cancer cases were compared to those in controls by chi-square (χ^2) test.

(HWE). Chi-square (χ^2) statistics were used to evaluate breast cancer cases and controls differences in the distribution of -2548A/G genotypes. The relationship between genotypes and breast cancer was also examined by odds ratios (ORs) with 95% confidence intervals (CIs) in logistic regression models. Stratified analyses according to menopausal status were then carried out. The comparisons of the case-control for serum leptin levels were performed with a non-parametric Mann-Whitney U test. A p value of <0.05 was considered statistically significant.

Results

Genotype and Allele Frequencies in All Women

The genotypes distributions and allele frequencies for the -2548G/A LEP gene variants in breast cancer cases and controls, and their corresponding ORs are presented in Table 1. The Genotypes distributions were in Hardy-Weinberg equilibrium in both breast cancer cases and controls at this locus. In breast cancer cases the genotype

Genotype	Cases	Controls		p value OR (95%CI)	OR (95%CI)*	p value
All Women	(n=100)	(n=100)		///		
Genotype frequencies						
AA	36 (36.0)	52 (52.0)	1.0			
AG	55 (55.0)	45 (45.0)	1.76 (0.98-3.15)	0.055	1.72 (0.95-3.09)	0.07
GG	9 (9.0)	3 (3.0)	4.33 (1.09- 17.22)	0.036	4.23 (1.06- 18.83)	0.041
AA	36 (36.0)	52 (52.0)	1.0			
AG+GG	64 (64.0)	48 (48.0)	1.92(1.09-3.39)	0.023	1.87 (1.05-3.33)	0.031
Pre-menopausal Genotype frequencies	(n=55)	(n=60)				
AA	21 (38.2)	30 (50.0)	1.0			
AG	32 (58.2)	28 (46. 7)	1.63 (0.76-3.46)	0.202	1.50 (0.68-3.28)	0.307
GG	2 (3.6)	2 (3.3)	1.42 (0.18- 10.96)	0.732	1.31 (0.16- 10.28)	0.796
AA	21 (38.2)	30 (50.0)	1.0		· · · · ·	
AG+GG	34 (61.8)	30 (50.0)	1.61 (0.77-3.40)	0.204	1.49 (0.69-3.21)	0.310
Post-menopausal Genotype frequencies	(n=45)	(n=40)				
AA	15 (33.3)	22 (55.0)	1.0			
AG	23 (51.1)	17 (42.5)	1.98 (0.80-4.91)	0.139	1.96 (0.78-4.95)	0.151
GG	7 (15.6)	1 (2.5)	10.26 (1.14- 92.25)	0.038	12.24 (1.13- 131.73)	0.039
AA	15 (33.3)	22 (55.0)	1.0		,	
AG+GG	30 (66.7)	18 (45.0)	2.44 (1.01-5.88)	0.046	2.34 (0.95-5.77)	0.064

Table 2. Distribution of LEP	-2548G/A	genotypes	in re	elation	breast	cancer	risk	(unconditional	logistic
regression).									

Data are presented as number of cases with frequency in parentheses. Results were presented for the dominant genetic model (homozygous for LEP -2548 AA vs. Carriers of G allele); OR=odds ratio. *Adjusted for age and BMI.

frequencies were 36% for AA, 55% for AG and 9% for GG, and were 52% for AA, 45% for AG and 3% for GG in controls (Table 2). The frequency of GG genotype was significantly elevated in breast cancer cases as compared to controls (χ^2 =6.90, p=0.032). Similar significance differences were also found in allele frequencies for A and G of the breast cancer cases, which were 63.5% and 36.5% compared with 74.5% and 25.5% of the controls (χ^2 =5.65, p=0.017) (Table 1).

Genotype and Allele Frequencies in Premenopausal Women

The LEP -2548 G allele was less frequent in premenopausal controls compared to those with breast cancer cases with allele frequencies of 26.7 and 32.7, respectively. The LEP -2548 AA genotype was less frequent in the women with breast cancer

cases compared to controls, with genotype frequencies of 38.2 and 50, respectively (Table 1).

Genotype and Allele Frequencies in Postmenopausal Women

There was a significant difference in the distribution of the LEP -2548G/A genotype in postmenopausal breast cancer cases compared to the controls (χ^2 =6.45, p=0.04) (Table 1).

LEP -2548G/A Genotypes and Breast Cancer Risk in All Women

Compared to women with the LEP -2548 AA genotype, those bearing the heterozygous LEP -2548 AG genotype (OR=1.765, 95% CI=0.98-3.15, p=0.055) had no significant increased risk of breast

variables	Mean leptin level ±SD (ng/ml)	p value*
Genotypes		
Breast Cancer Cases		
AA	68.20±45.90	
AG+GG	69.33±41.92	0.957
Controls		
AA	34.35±25.28	
AG+GG	30.97±28.80	0.364
Pre-menopausal		
Breast cancer cases	67.39±43.26	
Controls	35.13±28.23	0.001
Post-menopausal		
Breast cancer cases	71.56±43.08	
Controls	28.88±24.80	< 0.001
Pre-menopausal		
in breast cancer cases		
ER positive	68.45±42.68	
ER negative	59.10±45.39	0.258
PR positive	73.43±40.83	
PR negative	36.00±40.65	0.008

Table 3. Mean leptin levels according to LEP -2548 G/A genotypes, menopausal status, estrogen and progesterone receptor status at disease onset

*p values derived by non-parametric test

cancer. However, Compared to women with the LEP -2548 AA genotype, those bearing the homozygous - 2548 GG genotype (OR=4.33, 95% CI=0.09-17.22, p=0.036) have significant increased risk of breast cancer. Thus, the presence of at least one LEP -2548 G allele (LEP -2548 AG + LEP -2548 GG genotypes) was associated a significant risk of breast cancer in these women (OR=1.926, 95% CI=1.09-3.39, p=0.023) (Table 2).

LEP -2548G/A Genotypes and Breast Cancer Risk in Premenopausal Women

Among premenopausal women, there was no significant association between the LEP -2548G/A polymorphism and breast cancer risk (Table 2).

LEP -2548G/A Genotypes and Breast Cancer Risk in Postmenopausal Women

Among premenopausal women, compared to women with LEP -2548 AA genotype, those bearing the homozygous LEP -2548 GG (OR=10.26, 95% CI=1.14-92.25, p=0.038) had significant increased risk of breast cancer. Moreover, Adjusting for age and BMI confounders, did markedly and significantly increased risk of breast cancer in these women (OR=12.24, 95% CI=1.13-131.73, p=0.039) as shown in table 3. Therefore, the presence of at least one LEP -2548G allele (LEP -2548AG +LEP-2548GG genotypes) was associated with a significant increased risk of breast cancer in these women (OR=2.44, 95% CI=1.01-5.88, p=0.046) (Table 2).

Relationships between serum leptin level and breast cancer

Because of the opposite effect of obesity on breast cancer risk among pre- and post-menopausal women, we compared the serum levels of leptin between breast cancer cases and controls according to the menopausal status. The mean leptin levels was significantly different between breast cancer cases

and controls according to the pre-menopausal (67.39±43.26 vs. 35.13± 28.23 ng/ml, p=0.001) and post-menopausal (71.56±43.08 vs. 28.88±24.80 ng/ml, p<0.001), respectively. Pre-menopausal breast cancer cases with positive progesterone receptors (PR) on breast cancer tissue indicated significantly higher serum leptin levels than those with negative PR (73.43±40.83 vs. 36.00±40.65 ng/ml, p=0.008). There was no significant difference in leptin levels in pre-menopausal breast cancer cases with positive estrogen receptor (ER) and negative ER (68.45±42.68 vs. 59.10±45.39 ng/ml, p=0.258). Additionally, to determine whether LEP -2548G/A polymorphism is associated with differences in serum leptin levels in breast cancer cases and controls, we compared the mean levels of serum leptin in each group according to the LEP -2548G/A genotypes. Results indicated that there were no significant difference in mean levels of serum leptin in both breast cancer cases and controls according to the LEP -2548G/A genotypes (Table 3).

Discussion

This study showed significant differences in genotypic distribution and allele frequencies of the LEP -2548G/A polymorphism between breast cancer cases and controls. Additionally, women with the LEP -2548G allele have markedly increased risk of breast cancer, an association that was modified by menopausal status. In this way, our results supported the hypothesis that breast cancer might be associated with the serum levels of leptin.

Our results indicated that there was no significant difference regarding the mean serum levels of leptin between case and control groups according to the LEP -2548G/A observed genotypes. Data regarding to the potential action of the LEP -2548G/A genotypes on the serum levels of leptin is still controversial. A study conducted in the Taiwanese population reported that no association was found between this LEP polymorphism and serum levels of the leptin [28]. Conversely, reports from a cohort study in men French showed that there was association between AA genotype and increase serum levels of leptin [29]. This finding was confirmed by a Sweden study, which revealed that the -2548 A allele of this polymorphism was associated with increased leptin mRNA levels and increased rate of leptin secretion from adipose tissue [20]. Inversely, data from a study conducted in a French population, revealed that the -2548G/A LEP polymorphism could potentially alter leptin expression, and female subjects with the AA homozygous had lower mean levels of leptin than girls with other genotypes [19]. The association between the AA genotype of LEP -254G/A and lower serum levels of leptin was also described in healthy subjects from Greece, in French morbidly obese patients and in obese Brazilian women [30-32]. The reason of these different results is not completely understood but suggested due to interaction of LEP -2548G/A polymorphism with other polymorphisms in leptin and/or leptin receptor genes, sample size of population, or from to the model used in statistical analysis.

However, we found a significant difference of serum lptin levels between case and control according to the both premenopausal and postmenopausal status. These results conflict with the reports of several studies [2, 33]. According to another study in northern Sweden, no significant association between leptin levels and risk of postmenopausal breast cancer was observed [24]. The issue regarding leptin levels and breast cancer was controversial and an additional study with a larger number of cases would be helpful to clarify this relationship. Compared with the leptin levels used in several experiments that claimed that leptin caused significant effects on cellular proliferation [12, 21, 25, 34] there was a middle gap between the mean serum leptin levels in experiments, above 100 ng/ml, and in that of our post-menopausal patient group 71.56 ng/ml. Further studies are needed to elucidate to what degree the serum leptin levels reflect tissue levels in the breast and whether or not there would be a threshold of leptin which could stimulate the development of cancer.

Reports in premenopausal breast cancer cases were showed some association between leptin level and increased tissue levels of PR from cancer tissue. In a study conducted by Tessitire et al. in patients with breast cancer, found that, increased tissue levels of PR were associated with increased leptin levels [10]. In consideration of a finding that leptin is a hormone in adipose tissues that are regulated by transcriptional control with sex steroid hormones including progesterone [35], it thought to be there was some relationship between leptin and progesterone. Thus, leptin could stimulate the production of progesterone, or amplified signals through PR could increase the production of leptin.

Leptin, seems to be a growth factor, has several effects on the growth of normal and tumor cell, migration and invasion of tumor cell, development of angiogenesis, and proposed that can promote an aggressive of breast cancer phenotype [36]. Previous studies regarding to the association between LEP -2548 polymorphism and breast cancer risk have been controversial. Data reported in a case-control study conducted in Tunisian population showed that subjects who carried the LEP -2548AA genotype having a threefold increase in risk of breast carcinoma, those carried the heterozygous GA genotype having an intermediate risk compared to women who were homozygous for the LEP -2548 G allele [37]. In a large population-based study of mostly European - American women reported that the LEP -2548AA genotype was associated with a modest (30%) increase in the risk of breast cancer when compared to those with the LEP -2548GG genotype, an association that was not modified by menopausal status. Moreover, they did not observe an increased association with breast cancer risk for subjects bearing a single A allele [38]. Conversely, we observed 4-fold increased association with breast cancer risk for carrying a single G allele. Whether the differences in the magnitude of the association according to the LEP genotype between two studies is due to differences in ethnicity, sample size, or statistical analysis, is not clear. Although, the functional status of the LEP -2548G/A polymorphism is not completely understood, however, previous reports have shown that the A allele is associated with increased mRNA expression and higher circulating levels of leptin [20]. Additionally, the A allele of the LEP -2548 polymorphism has also been linked to other cancers, such as those of the lung and prostate [39, 40]. In spite of evidence that leptin induces proliferation of breast cancer cell in vitro [41] however, epidemiological reports regard to the effects of circulating levels of leptin on the breast cancer development and progression has been limited. A study conducted by Hu et al. showed an inverse association between circulating levels of leptin and breast cancer development in premenopausal women only [21], though in two other studies no such association was found [24, 26]. With regard to breast cancer prognosis, one study found no association between leptin levels and either disease-free or overall survival [42]. Why these studies failed to show evidence that leptin levels are associated with breast cancer is unknown. One probability may be

that the leptin levels in these studies were measured in circulation which could vary significantly from tissue concentrations found in the breast.

Conclusion

The results of this study have demonstrated a markedly increased risk of breast cancer in women bearing the LEP -2548G allele, and the association may be markedly profound in the postmenopausal women. According to our knowledge, this is the first study to provide information on the role of LEP -2548G/A polymorphism in breast cancer risk in a sample of Iranian women. These results should be confirmed with additional studies to further evaluate the potential action and interaction between this polymorphism, obesity and breast cancer development both in-vivo and in-vitro.

Acknowledgement

This study is part of M.Sc thesis for Ahmmad Bafandeh which has been kindly supported by Hyperlipidemia Research Center, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran, whom we thanks a lot (Grant No. HLRC-9101). We would like to thank the staffs of Radiation and Oncology Department of Golestan University Hospital for their friendly cooperation. We have extended our thanks to all the participants and their relatives for their kind cooperation with the researchers as well.

Conflicts of Interest

There is no conflict of interest in this article.

Authors' Contribution

All authors participated in the study design, data analysis, interpretation and manuscript writing.

References

1. Ray A. Adipokine leptin in obesity-related pathology of breast cancer. J Biosci. 2012;37:289-94.

2. Goya M. Iranian annual cancer registration report. Ministry of health and medical education, health deputy, center for disease control and prevention. March 2005.

3. Goldgar DE, Healey S, Dowty JG, Silva LD, Chen X, Spurdle AB, et al. Rare Variants in the ATM Gene and Risk of Breast Cancer. Breast Cancer Res. 2011;13(4):R73

4. Ahima RS, Lazar MA. Adipokines and the peripheral and neural control of energy balance. Mol Endocrinol. 2008;22:1023–31.

5. Bingham NC, Anderson KK, Reuter AL, Stallings NR, Parker KL. Selective loss of leptin receptors in the ventromedial hypothalamic nucleus results in increased adiposity and a metabolic syndrome. Endocrinology. 2008;149:2138–48.

6. Miyoshi Y, Funahashi T, Tanaka S, Taguchi T, Tamaki Y, Shimomura I, Noguchi S. High expression of leptin receptor mRNA in breast cancer tissue predicts poor prognosis for patients with high, but not low, serum leptin levels. Int J Cancer. 2006;118:1414–19.

7. Ishikawa M, Kitayama J, Nagawa H. Enhanced expression of leptin and leptin receptor (OB-R) in human breast cancer. Clin Cancer Res. 2004; 10:4325–4331.

8. Catalano S, Marsico S, Giordano C, Mauro L, Rizza P, Panno ML, Ando S. Leptin enhances, via AP-1, expression of aromatase in the MCF-7 cell line. J Biol Chem. 2003;278:28668–76.

9. Garofalo C, Koda M, Cascio S, Sulkowska M, Kanczuga-Koda L, Golaszewska J, et al. Increased expression of leptin and the leptin receptor as a marker of breast cancer progression: possible role of obesity-related stimuli. Clin Cancer Res. 2006;12:1447–53.

10. Tessitore L, Visio B, Pesola D, Cecchini f, Mussa A, Argiles JM, Benedetto C: Adipocytes expression and circulating levels of leptin increase in both gynecological and breast cancer patients. Int J Cancer. 2004;24:1529-35.

11. Han C, Zhang HT, Du L, Liu X, Jing J, Zhao X, Tian B. Serum levels of leptin, insulin, and lipids in relation to breast cancer in china. Endocrine. 2005;26:19-24.

12. Laud K, Gourdou I, Pessemesse L, Peyrat JP, Djiane J. Identification of leptin receptors in human breast cancer: functional activity in the T47-D breast cancer cell line. Mol Cell Endocrinol. 2002;188:219–26.

13. Koda M, Sulkowska M, Wincewicz A, Kanczuga-Koda L, Musiatowicz B, Szymanska M, Sulkowski S. Expression of leptin, leptin receptor, and hypoxiainducible factor 1 alpha in human endometrial cancer. Ann N Y Acad Sci. 2007;1095:90–8.

14. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM: Positional cloning of the mouse obese gene and its human homologue. Nature. 1994;372:425-32.

15. Green ED, Maffei M, Braden VV, Proenca R, DeSilva U, Zhang Y, Chua SC, Leibel RL, Weissenbach J, Friedman JM: The human obese (OB) gene: RNA expression pattern and mapping on the physical, cytogenetic, and genetic maps of chromosome 7. Genome Res. 1995;5:5-12.

16. Isse N, Ogawa Y, Tamura N, Masuzaki H, Mori K, Okazaki T, Satoh N, et al. Structural organization and chromosomal assignment of the human obese gene. J Biol Chem. 1995;270:27728-33.

17. Masuzaki H, Ogawa Y, Isse N, Satoh N, Okazaki T, Shigemoto M, et al. Human obese gene expression: Adipocyte-specific expression and regional differences in the adipose tissue. Diabetes. 1995;44:855-8.

18. Mammès O. Betoulle D. Aubert R, Giraud S, Tuzet A, Petiet N, et al. Novel polymorphisms in the 50 region of the LEP gene: association with leptin levels and response to low-calorie diet in human obesity. Diabetes. 1998;47:487–9.

19. Le Stunff C, Le Bihan C, Schork NJ, Bougneres P. A common promoter variant of the leptin gene is associated with changes in the relationship between serum leptin and fat mass in obese girls. Diabetes. 2000;49:2196–200.

20. Hoffstedt J, Eriksson P, Mottagui-Tabar S, Arner P. A polymorphism in the leptin promoter region (-2548 G/A) influences gene expression and adipose tissue secretion of leptin. Horm Metab Res. 2002;34:355–9.

21. Hu X, Juneja SC, Maihle NJ, Cleary MP. Leptin–a growth factor in normal and malignant breast cells and for normal mammary gland development. J Natl Cancer Inst. 2002;94:1704–11.

22. Sauter ER, Garofalo C, Hewett ER, Morelli C, Surmacz E. Leptin expression in breast nipple aspirate fluid (NAF) and serum is influenced by body mass index (BMI) but not by the presence of breast cancer. Horm Metab Res. 2004;36:336-40.

23. Ozet A, Arpaci F, Yilmaz M, Ayta H, Ozturk B, Komurcu S, et al. Effects of tamoxifen on the serum leptin level in patients with breast cancer, Jpn. J clin Oncol. 2001;31:424-7.

24. Stattin P, Soderberg S, Biessy C, Lenner P, Hallmans G, Kaaks R, et al., Plasma leptin and breast cancer risk: a prospective study in northern Sweden. Breast Cancer Res Treat. 2004;86:191–6.

25. Dieudonne MN, Machinal-Quelin F, Serazin-Leroy V, Leneveu MC, Pecquery R, Giudicelli Y. Leptin mediates a proliferative response in human MCF7 breast cancer cells. Biochem Biophys Res Commun. 2002;293:622–8.

26. Mantzoros CS, Bolhke K, Moschos S, Cramer DW. Leptin in relation to carcinoma in situ of the breast: a study of premenopausal cases and controls. Int J Cancer. 1999;80:523–6.

27. Mohammadzadeh G, Ghaffari M, Bafandeh A, Hosseini S. Association of serum soluble leptin receptor and leptin levels with breast cancer. J Res Med Sci. 2014;19:433-8.

28. Wang TN, Hung MC, Chang WT, et al. G-2548A polymorphism of the leptin gene is correlated with extreme obesity in Taiwanses aborigines. Obesity. 2006;14:183-7.

29. Mammes O, Betoulle D, Aubert R, Herbeth B, Siest G, Fumeron F. Association of the G-2548A polymorphism in the 50 region of the LEP gene with overweight. Ann Hum Genet. 2000;64:391–4.

30. Yiannakouris N, Relistas L, Yannakoulia M, Mungal K, Mantzoros CS. The-2548G/A polymorphism in the human leptin gene promoter region is associated with plasma free leptin levels: interaction with adiposity

and gender in healthy subjects. Hormones. 2003;2:229-36.

31. Poitou C, Lacorte JM, Coupaye M, et al. relationship between single nucleotide polymorphisms in leptin, IL6 and adiponectin gene and their circulating product in morbidly obese subjects before and after gastric banding surgery. Obese Surg. 2005;15:11-23.

32. Hinuy HM, Hirata MH, Forti N, et al. leptin G-2548A promoter polymorphism is associated with increased plasma leptin and BMI in Brazilian women. Arq Bras Endocrinol metab. 2008;52:611-6.

33. O'Brien SN, Welter BH, Price TM, Clemson U, Glemson SG. Presence of leptin in breast cell lines and breast tumors. Biochem Biophys Res Commun. 1999;259:695–8.

34. Okumura M, Yamamoto M, Sakuma H, Kojima T, Maruyama T, Jamali M, et al. Leptin and high glucose stimulate cell proliferation in MCF-7 human breast cancer cells: reciprocal involvement of PKC-alpha and PPAR expression. Biochim. Biophys. Acta. 2002;1592:107–16.

35. Mayes JS, Watson JS. Direct effects of sex steroid hormones on adipose tissues and obesity. Obes Rev. 2004;5:197–216.

36. Rose DP, Gilhooly EM, Nixon DW. Adverse effects of obesity on breast cancer prognosis and the biological actions of leptin (review). Int J Oncol. 2002;21:1285–92

37. Snoussi K, Strosberg AD, Bouaouina N, Ben Ahmed S, Helal AN, Chouchane L. Leptin and leptin receptor polymorphisms are associated with increased risk and poor prognosis of breast carcinoma. BMC Cancer. 2006;6:38.

38. Cleveland RJ, Gammon MD, Long CM, Gaudet MM, Eng SM, Teitelbaum SL, et al. Common genetic variations in the LEP and LEPR genes, obesity and breast cancer incidence and survival. Breast Cancer Res Treat. 2010;120(3):745–52.

39. Ribeiro R, Araujo AP, Coelho A, Catarino R, Pinto D, Araujo A, Calcada C, Lopes C, Medeiros R. A functional polymorphism in the promoter region of leptin gene increases susceptibility for non-small cell lung cancer. Eur J Cancer. 2006;42:1188–93.

40. Ribeiro R, Vasconcelos A, Costa S, Pinto D, Morais A, Oliveira J, Lobo F, Lopes C, Medeiros R. Over expressing leptin genetic polymorphism (-2548 G/A) is associated with susceptibility to prostate cancer and risk of advanced disease. Prostate. 2004;59:268–74.

41. Petridou E, Papadiamantis Y, Markopoulos C, Spanos E, Dessypris N, Trichopoulos D Leptin and insulin growth factor I in relation to breast cancer. Cancer Causes Control. 2000;11:383–8.

42. Goodwin PJ, Ennis M, Fantus IG, Pritchard KI, Trudeau ME, Koo J, Hood N. Is leptin a mediator of adverse prognostic effects of obesity in breast cancer? J Clin Oncol. 2005;23:6037.