ANRIL Genetic Variants in Iranian Breast Cancer Patients

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> Received: 31/Jul/2016, Accepted: 26/Sep/2016 Abstract

Objective: The genetic variants of the long non-coding RNA *ANRIL* (an antisense noncoding RNA in the INK4 locus) as well as its expression have been shown to be associated with several human diseases including cancers. The aim of this study was to examine the association of *ANRIL* variants with breast cancer susceptibility in Iranian patients.

Materials and Methods: In this case-control study, we genotyped rs1333045, rs4977574, rs1333048 and rs10757278 single nucleotide polymorphisms (SNPs) in 122 breast cancer patients as well as in 200 normal age-matched subjects by tetra-primer amplification refractory mutation system polymerase chain reaction (T-ARMS-PCR).

Results: The TT genotype at rs1333045 was significantly over-represented among patients (P=0.038) but did not remain significant after multiple-testing correction. In addition, among all observed haplotypes (with SNP order of rs1333045, rs1333048 rs4977574 and rs10757278), four haplotypes were shown to be associated with breast cancer risk. However, after multiple testing corrections, TCGA was the only haplotype which remained significant.

Conclusion: These results suggest that breast cancer risk is significantly associated with *ANRIL* variants. Future work analyzing the expression of different associated *ANRIL* haplotypes would further shed light on the role of *ANRIL* in this disease.

Keywords: ANRIL, Breast Cancer, Polymorphism

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Introduction

Chromosome region 9p21 is a hotspot for disease-associated polymorphisms and encodes three tumor suppressors, namely $p16^{INK4a}$, $p14^{ARF}$ and $p15^{INK4b}$, and the long non-coding RNA *ANRIL* (an antisense noncoding RNA in the INK4 locus) (1). This region has been shown to be altered in about one third of human tumors. *ANRIL* is a 3.8 kb-long non-coding RNA expressed on the reverse strand and has been shown to bind to and recruit PRC2 to repress the expression of $p15^{INK4B}$ (2). Figure 1 shows the genomic location of *ANRIL* and its function in regulation of cell cycle. *ANRIL* expression has been shown to be upregulated in breast cancer tissues with a significantly higher expression in triple-negative highly invasive

(GWAS) have identified *ANRIL* as a risk locus for numerous cancers such as breast cancer (4). This susceptibility may be explained by individual, tightly linked single nucleotide polymorphisms (SNPs) in the *ANRIL* locus; changing expression of *ANRIL* spliced transcripts and consequently influencing cellular proliferation pathways (5). *ANRIL* expression has been shown to be upregulated after DNA damage by the transcription factor E2F1. This suggests that *ANRIL* is involved in an ATM-dependent DNA damage response. In addition, increased levels of *ANRIL* inhibit the expression of *p16^{INK4a}*, *p14^{ARF}* and *p15^{INK4b}* at the late-stage of DNA damage response (6).

cancers (3). Genome-wide association studies



Fig.1: The location of ANRIL and its function in regulation of cell cycle.

Cardiovascular disorders are the most investigated human disorders which have been associated with ANRIL variants. For instance, rs4977574 has been shown to be strongly associated with the risk of coronary artery disease (7). On the other hand, rs11515 has been shown to be over-represented among breast cancer patients and has been associated with aggressive breast tumors, and higher ANRIL and lower $p16^{INK4a}$ expression (1). Among other genetic variants within this gene, rs10757278 has been shown to increase the expression of the ANRIL variant EU741058 which contains exons 1-5 of the long transcript (8). Additionally, rs10757278 has been shown to modulate the ANRIL binding site for the transcription factor STAT1, which in turn regulates ANRIL expression (9). Considering the role of STAT1 in shaping an immunosuppressive tumor microenvironment in breast cancer cells (10), disruption of ANRIL binding site for STAT1 by the rs10757278 allele may be involved in breast cancer pathogenesis. Moreover, rs1333045 is an artery disease susceptibility SNP located in a conserved region in ANRIL which has been shown to have an enhancer activity in a reporter gene experiment (11). The other SNP within this gene, rs1333048, has been associated with the level of high sensitive C-reactive protein (hsCRP) which is a marker for systemic inflammation (12). Since recent studies have suggested an association between pre-diagnostic hsCRP and breast cancer risk as well as overall mortality (13), this variant might be associated with breast cancer susceptibility.

The role of *ANRIL* in breast cancer pathogenesis and risk has been assessed in both expression and GWA studies; however, data regarding the role of specific SNPs within this gene in breast cancer susceptibility are scarce. Consequently, to find the association of *ANRIL* variants with breast cancer susceptibility in Iranian patients, we genotyped and examined the association of rs1333045, rs4977574, rs1333048 and rs10757278 according to their significance in the regulation of *ANRIL* expression and their participation in breast cancer-related pathways.

Materials and Methods

This case-control study was approved by the Ethical Committee of Hamadan University Hospital where 122 unrelated breast cancer patients, as well as 200 normal age-matched females from a routine health survey, were recruited during 2015 (IR.UMSHA.REC.1395.208). Informed consent was obtained from all participants. Clinical and pathological data of patients were collected. Diagnosis of breast cancer was confirmed by a pathologic study.

Single nucleotide polymorphism genotyping

Genomic DNA was extracted from blood samples of the patients and normal subjects using the standard salting out method. SNPs rs1333045, rs4977574, rs1333048 and rs10757278 were genotyped by tetra-primer amplification refractory mutation system PCR (T-ARMS-PCR) (14). PCR was performed in 25 μ L total volume using Taq (2x) red master mix (Ampliqon, Denmark) and 0.5 µL of each forward and reverse primer (10 pmol) in a FlexCycler (Analytik Jena, Germany). The cycling conditions were an initial denaturation at 94°C for 4 minutes, followed by 35 cycles of 94°C for 45 seconds, annealing temperature for 45 seconds and 72°C for 55 seconds, with a final extension of 72°C for 5 minutes. Specific annealing temperatures were 45°C for rs1333048, 53°C for rs4977574, 52°C for rs1333045 and 54°C for rs10757278. The primers were designed by PRIMER1 (http://cedar. genetics.soton.ac.uk/public html/primer1.html.) and are listed in Table 1.

Statistical analysis

The genotype and allele frequencies were calculated by direct counts. Deviation from the Hardy-Weinberg equilibrium was assessed using the Chi-square test. Pearson Chi-square test

was utilized for comparing genotype and allele frequencies between the breast cancer patients and the control group using SPSS 16.0 (SPSS Inc., Chicago, IL, USA). Odds ratio (OR) and 95% confidence intervals (CI) were also calculated. These analyses were implemented. Haplotype frequencies for ANRIL were calculated using the SNPStats (http://bioinfo. iconcologia.net/SNPstats) based on the expectationmaximization algorithm (15). Pairwise linkage disequilibrium (LD) was assessed by calculating D' and squared correlation (r²) in Haploview (https:// www.broadinstitute.org/haploview/haploview) (16). D' was determined as the ratio of the unstandardized D to its maximal/minimal value. To avoid false positive results, permutation testing was performed (n=10,000) for multiple testing correction of the haplotype analysis. Differences were regarded as significant when P < 0.05.

Results

Comparison of age between cases and controls showed no significant difference (mean age of patients: 38.9 ± 2.1 and mean age of healthy controls: 39.1 ± 1.8). The frequencies of all genotypes in both patients and control groups did not significantly deviate from Hardy-Weinberg equilibrium (P>0.05). The allele and genotype frequencies of the SNPs and the association results are shown in Table 2. Among all genotypes, only the TT genotype at rs1333045 was significantly more prevalent among patients (P=0.038), however, it did not remain significant after multiple-testing correction.

Table 1: Sequence of primers					
Primer positionPrimer sequencePCR pro-					
rs1333045					
Forward inner primer (C allele)	CGAAGAGCAATAATATATAGTACACTGGGC	for C allele: 200			
Reverse inner primer (T allele)	TTAATGAATGCTTACTAGATGCCTGA	for T allele: 298			
Forward outer primer (5'-3')	TGAAACTTCTTATTTAGTGGTGCATACC	by outer primers: 442			
Reverse outer primer (5'-3')	GCAGTTCAAAGGAAGTACCATAAAAAG				
rs4977574					
Forward inner primer (G allele)	TTGAGGGTACATCAAAAGCATTCTATATCG	for G allele: 226			
Reverse inner primer (A allele)	TTTATTAGAGTGACTTGAACATCCCGT	for A allele: 166			
Forward outer primer (5'-3')	CACCATTCTTTCTGAAACAACAGGATAT	by outer primers: 335			
Reverse outer primer (5'-3')	AAGGCTCTGACATTTCTAACTCTCTGA				
rs1333048					
Forward inner primer (A allele)	TTAATGCTATTTTGAGGAGATGTCTA	for A allele: 185			
Reverse inner primer (C allele)	TTTTATCAATATTTCAATAATTCGACACTG	for C allele: 253			
Forward outer primer (5'-3')	TTGCCTGATTACCAATTTTATATGTTA	by outer primers: 382			
Reverse outer primer (5'-3')	TCAACTGATGATGATATGGTTAGTATG				
rs10757278					
Forward inner primer (A allele)	AAGTCAGGGTGTGGTCATTACGGGAA	for A allele: 263			
Reverse inner primer (G allele)	CTCAGTCTTGATTCTGCATCGCTTCC	for G allele: 234			
Forward outer primer (5'-3')	GGGCATTAAGAAAtGGATGGGTAGACAAAA	by outer primers: 443			
Reverse outer primer (5'-3')	GCTGTTCTCAATTAGCCAGGACTACCTCT				

PCR; Polymerase chain reaction.

SNP	Mod	el	Numb	er (%)	Cancer vs. cont	trol
			Cancer (%)	Control (%)	OR	P value
rs1333045	Allele	T vs. C	130 (53)	186 (46)	1.31 (0.95-1.80)	0.09
			114 (47)	214 (54)		
	Co-dominant	TT vs. CC	39 (32)	43 (21.5)	0.60 (0.32-1.11)	0.11
		CT vs. CC	52 (42.6)	100 (50)	1.05 (0.60-1.81)	
	Dominant	TT+CT vs. CC	91 (74.6)	143 (71.5)	0.85 (0.51-1.42)	0.54
			31 (25.4)	57 (28.5)		
	Recessive	TT vs. CT+CC	39 (32)	43 (21.5)	0.58 (0.35-0.97)	0.038
			83 (68)	157 (78.5)		
	Over dominant	TT+CC vs. CT	70 (57.4)	100 (50)	1.35 (0.86-2.12)	0.2
			52 (42.6)	100 (50)		
rs1333048	Allele	C vs. A	115 (47)	201 (50)	0.88 (0.64-1.21)	0.44
			129 (53)	199 (50)		
	Co-dominant	CC vs. AA	32 (26.2)	52 (26)	1.24 (0.68-2.28)	0.39
		CA vs. AA	51 (41.8)	97 (48.5)	1.45 (0.85-2.49)	
	Dominant	CC+CA vs. AA	83 (68)	149 (74.5)	1.37 (0.84-2.25)	0.21
			39 (32)	51 (25.5)		
	Recessive	CC vs. CA+AA	32 (26.2)	52 (26)	0.99 (0.59-1.65)	0.96
			90 (73.8)	148 (74)		
	Over dominant	AA+CC vs. CA	71 (58.2)	103 (51.5)	1.31 (0.83-2.06)	0.24
			51 (41.8)	97 (48.5)		
rs4977574	Allele	A vs. G	78 (32)	145 (36)	0.83 (0.59-1.16)	0.27
			166 (68)	255 (64)		
	Co-dominant	AA vs. GG	17 (13.9)	26 (13)	1.15 (0.57-2.31)	0.17
		GA vs. GG	44 (36.1)	93 (46.5)	1.59 (0.98-2.60)	
	Dominant	GA+AA vs. GG	61 (50)	119 (59.5)	1.47 (0.93-2.31)	0.096
			61 (50)	81 (40.5)		
	Recessive	AA vs. GA+GG	17 (13.9)	26 (13)	0.92 (0.48-1.78)	0.81
			105 (86.1)	174 (87)		
	Over dominant	GG+AA vs. GA	78 (63.9)	107 (53.5)	1.54 (0.97-2.45)	0.065
			44 (36.1)	93 (46.5)		
rs10757278	Allele	A vs. G	106 (43)	152 (38)	1.25 (0.91-1.73)	0.17
			138 (57)	248 (62)		
	Co-dominant	AA vs. GG	22 (18)	26 (13)	0.61 (0.30-1.21)	0.36
		GA vs. GG	62 (50.8)	100 (50)	0.83 (0.50-1.37)	
	Dominant	GA+AA vs.GG	84 (68.8)	126 (63)	0.77 (0.48-1.24)	0.28
			38 (31.1)	74 (37)		
	Recessive	AA vs.GA+GG	22 (18)	26 (13)	0.68 (0.37-1.26)	0.22
			100 (82)	174 (87)		
	Over dominant	GG+AA vs.GA	60 (49.2)	100 (50)	0.97 (0.62-1.52)	0.89
NPs; Single nu			62 (50.8)	100 (50)		

Table 2: Allele and genotype frequencies of ANRIL SNPs	in the case and control groups
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ANRIL Polymorphisms in Breast Cancer Patients

Haplotype analysis was undertaken and distribution of haplotype frequencies in both groups was obtained (Table 3). Haplotype frequencies and the LD pattern (based on D') are shown in Table 3 and Figure 2 respectively. No significant LD was observed among the four SNPs (D'<0.6). Among all observed haplotypes (with SNP order of rs1333045, rs1333048 rs4977574

and rs10757278), four haplotypes were shown to be associated with breast cancer risk. However, after multiple testing corrections, TCGA was the only haplotype which remained significant. Interestingly, this haplotype has the derived allele at SNPs rs1333045 and rs10757278, of which the former showed a hint of association with its homozygote form (TT).

rs1333045	rs1333048	rs4977574	rs10757278	Total frequency	Frequency in cancer group	Frequency in control group	OR (95% CI)	P value	Corrected P value
С	С	G	G	0.245	0.159	0.298	1.00	-	-
Т	А	А	А	0.118	0.081	0.135	0.73 (0.35 - 1.53)	0.4	1
Т	А	G	А	0.101	0.113	0.095	0.55 (0.27 - 1.11)	0.097	0.175
Т	С	G	G	0.100	0.109	0.089	0.59 (0.27 - 1.28)	0.18	1
С	А	G	G	0.073	0.106	0.059	0.39 (0.17 - 0.88)	0.024	0.125
С	А	А	А	0.066	0.047	0.075	1.25 (0.42 - 3.77)	0.69	0.812
Т	А	G	G	0.042	0.045	0.038	0.63 (0.24 - 1.64)	0.34	1
С	А	А	G	0.039	0.068	0.019	0.27 (0.09 - 0.84)	0.024	0.687
Т	С	А	G	0.038	0.032	0.043	0.88 (0.28 - 2.73)	0.82	1
Т	А	А	G	0.035	0.032	0.046	0.82 (0.25 - 2.63)	0.74	1
С	А	G	А	0.035	0.037	0.031	0.54 (0.17 - 1.67)	0.29	1
Т	С	G	А	0.032	0.075	0.008	0.07 (0.01 - 0.39)	<10-4	0.002
С	С	А	G	0.026	0.014	0.028	1.31 (0.24 - 7.17)	0.76	0.812
С	С	G	А	0.025	0.036	0.019	0.43 (0.09 - 2.12)	0.3	1
Т	С	А	А	0.023	0.045	0.012	0.23 (0.06 - 0.92)	0.039	0.687

Table 3: Haplotype frequencies of ANRIL SNPs in the case and control groups

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Fig.2: Linkage disequilibrium (LD) plot based on the four SNPs typed. The values in each cell represent D' values.

Discussion

ANRIL identification has accentuated the underrated role of genes encoding long noncoding RNA in pathogenesis of human disorders including cancers (5). Long noncoding RNA have been shown to have a regulatory role in telomere biology, chromatin dynamics, gene modulation and genome structural organization (17, 18). Such a vast range of function suggests that they may also be involved in tumorigenesis processes. The role of ANRIL in regulation of DNA damage response makes it likely for it to have a critical role in breast cancer pathogenesis given that BRCA1, the most well-known breast cancer susceptibility gene mainly participates in DNA damage-induced cell cycle checkpoint activation and DNA repair (19). Other suggested roles for ANRIL include increasing cell proliferation and decreasing apoptosis (20) in addition to participation in inflammatory response; also emphasize its role in tumorigenesis (21). Many disease-associated SNPs identified by GWAS have been shown to be located in noncoding genomic

regions which may contain long noncoding RNA such as ANRIL. Although in the current study, no genotype was strongly associated with breast cancer, one haplotypes was associated with breast cancer susceptibility in this population. Apart from rs11515 which has been assessed in breast cancer patients (1), the other three SNPs within this gene have not been genotyped in breast cancer patients. So the present study is among the first studies which have genotyped multiple SNPs within the ANRIL locus in these patients. The selected Considering the numerous splicing variants of ANRIL and the tissue specificity of some of the splicing variants (22), its physiological significance may be tissuespecific and so would be the effects of each SNP on its splicing variants. Haplotypes may be in closer linkage disequilibrium with a causal variant than any single SNP assessed and is thus more likely to show association. Furthermore, haplotypes may themselves be the causative variants of interest (23). Accordingly, our study also shows that haplotype analysis is more valuable than single SNP analyses.

In addition, since the *ANRIL* genomic region encompasses various risk-associated SNPs (24), expression of *ANRIL* might be influenced by many of these genetic variants in linkage disequilibrium. Such deregulation of *ANRIL* by disease associated polymorphisms may change the expression level of $p15^{INK4B}$ and/ or other target genes (2). Considering the role of $p15^{INK4B}$ as a cyclin-dependent kinase inhibitor which prevents the activation of cyclin dependent kinases by cyclin D and functions as a cell growth regulator that inhibits cell cycle G1 progression (25), any change in its expression may have significant implications in tumorigenesis.

Conclusion

We show that ANRIL is associated with breast cancer susceptibility at the haplotype level. Further work comparing expression of *ANRIL* and its putative target genes based on different *ANRIL* haplotypes is necessary.

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References

- Royds JA, Pilbrow AP, Ahn A, Morrin HR, Frampton C, Russell IA, et al. The rs11515 polymorphism is more frequent and associated with aggressive breast tumors with increased ANRIL and decreased p16INK4a expression. Front Oncol. 2016; 5: 306.
- Kotake Y, Nakagawa T, Kitagawa K, Suzuki S, Liu N, Kitagawa M, et al. Long non-coding RNA ANRIL is required for the PRC2 recruitment to and silencing of p15(INK4B) tumor suppressor gene. Oncogene. 2011; 30(16): 1956-1962.
- Iranpour M, Soudyab M, Geranpayeh L, Mirfakhraie R, Azargashb E, Movafagh A, et al. Expression analysis of four long noncoding RNAs in breast cancer. Tumour Biol. 2016; 37(3): 2933-2940.
- Turnbull C, Ahmed S, Morrison J, Pernet D, Renwick A, Maranian M, et al. Genome-wide association study identifies five new breast cancer susceptibility loci. Nat Genet. 2010; 42(6): 504-507.
- Pasmant É, Sabbagh A, Vidaud M, Bièche I. ANRIL, a long, noncoding RNA, is an unexpected major hotspot in GWAS. FASEB J. 2011; 25(2): 444-448.
- Wan G, Mathur R, Hu X, Liu Y, Zhang X, Peng G, et al. Long non-coding RNA ANRIL (CDKN2B-AS) is induced by the ATM-E2F1 signaling pathway. Cell Signal. 2013; 25(5): 1086-1095.
- Huang Y, Ye H, Hong Q, Xu X, Jiang D, Xu L, et al. Association of CDKN2BAS polymorphism rs4977574 with coronary heart disease: a case-control study and a metaanalysis. Int J Mol Sci. 2014; 15(10): 17478-17492.
- Holdt LM, Beutner F, Scholz M, Gielen S, Gäbel G, Bergert H, et al. ANRIL expression is associated with atherosclerosis risk at chromosome 9p21. Arterioscler Thromb Vasc Biol. 2010; 30(3): 620-627.
- Harismendy O, Notani D, Song X, Rahim NG, Tanasa B, Heintzman N, et al. 9p21 DNA variants associated with coronary artery disease impair interferon-γ signalling response. Nature. 2011; 470(7333): 264-268.
- Hix LM, Karavitis J, Khan MW, Shi YH, Khazaie K, Zhang M. Tumor STAT1 transcription factor activity enhances breast tumor growth and immune suppression mediated by myeloid-derived suppressor cells. J Biol Chem. 2013; 288(17): 11676-11688.
- Jarinova O, Stewart AF, Roberts R, Wells G, Lau P, Naing T, et al. Functional analysis of the chromosome 9p21.3 coronary artery disease risk locus. Arterioscler Thromb Vasc Biol. 2009; 29(10): 1671-1677.

- Teeuw WJ, Laine ML, Bizzarro S, Loos BG. A lead ANRIL polymorphism is associated with elevated CRP levels in periodontitis: a pilot case-control study. PLoS One. 2015; 10(9): e0137335.
- Frydenberg H, Thune I, Lofterød T, Mortensen ES, Eggen AE, Risberg T, et al. Pre-diagnostic high-sensitive Creactive protein and breast cancer risk, recurrence, and survival. Breast Cancer Res Treat. 2016; 155(2): 345-354.
- Ye S, Dhillon S, Ke X, Collins AR, Day IN. An efficient procedure for genotyping single nucleotide polymorphisms. Nucleic Acids Res. 2001; 29(17): E88.
- Solé X, Guinó E, Valls J, Iniesta R, Moreno V. SNPStats: a web tool for the analysis of association studies. Bioinformatics. 2006; 22(15): 1928-1929.
- Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics. 2005; 21(2): 263-265.
- Santosh B, Varshney A, Yadava PK. Non-coding RNAs: biological functions and applications. Cell Biochem Funct. 2015; 33(1): 14-22.
- Soudyab M, Iranpour M, Ghafouri-Fard S. The role of long non-coding RNAs in breast cancer. Arch Iran Med. 2016; 19(7): 508-517.
- Wu J, Lu LY, Yu X. The role of BRCA1 in DNA damage response. Protein Cell. 2010; 1(2): 117-123.
- Sepahvand Hossein Beigi S, Ghaderian SMH, Doosti A. Investigation of the association between rs4977574 A> G polymorphism in ANRIL gene and coronary artery disease in Iranian population. Int Cardiovasc Res J. 2015; 9(3): 139-144.
- Zhou X, Han X, Wittfeldt A, Sun J, Liu C, Wang X, et al. Long non-coding RNA ANRIL regulates inflammatory responses as a novel component of NF-κB pathway. RNA Biol. 2016; 13(1): 98-108.
- Folkersen L, Kyriakou T, Goel A, Peden J, Mälarstig A, Paulsson-Berne G, et al. Relationship between CAD risk genotype in the chromosome 9p21 locus and gene expression. identification of eight new ANRIL splice variants. PLoS One. 2009; 4(11): e7677.
- Stram DO, Seshan VE. Multi-SNP haplotype analysis methods for association analysis. Methods Mol Biol. 2012; 850: 423-452.
- Broadbent HM, Peden JF, Lorkowski S, Goel A, Ongen H, Green F, et al. Susceptibility to coronary artery disease and diabetes is encoded by distinct, tightly linked SNPs in the ANRIL locus on chromosome 9p. Hum Mol Genet. 2008; 17(6): 806-814.
- Hall M, Bates S, Peters G. Evidence for different modes of action of cyclin-dependent kinase inhibitors: p15 and p16 bind to kinases, p21 and p27 bind to cyclins. Oncogene. 1995; 11(8): 1581-1588.