Computed Tomographic Anatomy of the Abdominal Cavity in the Jebeer (Gazella Bennettii)

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ABSTRACT

Introduction: Computed tomography (CT) is an imaging technique with gives us an opportunity to review cross sections of the body in live animals. In veterinary medicine although CT is mostly used for diagnostic purposes in small animals, but in recent years CT also has been used as a non invasive modality in non clinical studies. Jebeer Gazelle (Gazella bennettii) is one of the species of the genus Gazella which lives mostly in the south east of Iran and very little anatomic studies are available on it. The aim of this study was preparing detailed anatomic images of the abdominal cavity of the Jebeer as an endangered species using the non invasive CT technique.

Methods: Spiral CT images were acquired from the abdominal region of four healthy Jebeer, perpendicular to long axis of the body. CT windows were adjusted as necessary to have optimized images of the abdominal organs. The images were studied serially and compared anatomically with two dissected goats.

Results: Liver, spleen, reticulum, omasum, abomasum, rumen, right and left kidneys, transverse colon, ascending colon, descending colon, cecum, pancreas, duodenum, uterine horns, urinary bladder and jejunum were distinguished and addressed according to the thoracic and lumbar vertebrae as landmarks.

Conclusion: According to the present study, we identified the abdominal organs, their precise position and related structures in the Jebeer without any invasive procedure. This is the first study which is addressing abdominal organs of a wild ruminant by using CT modality.

Key Words:

Computed tomography, Anatomy, Deer, Abdomen

1. Introduction

omputed tomography (CT) is an imaging technique with gives us an opportunity to review cross sections of the body in live animals. It is a valuable tool for evaluating diseases and makes it possible to have better diagnosis for presence location and extension of pathology and involvement of structures in comparison to standard radiography [1]. In veterinary medicine although CT is mostly used for diagnostic purposes in small animals [1-7], but in recent years CT also has been used as a non invasive modality in non clinical studies [8-23].

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Jebeer Gazelle (Gazella bennettii) is one of the species of the genus Gazella which lives mostly in the south east of Iran [24, 25]. Although the species is fully protected by the law, only around 1300 heads were estimated for this country [26], and it seems that more consideration must be taken into account. In case of Jebeer very little anatomic studies are available [27, 28] and the aim of this study was preparing detailed anatomic images of the abdominal cavity of the Jebeer as an endangered species using the non invasive CT technique.

2. Materials & Methods

Four healthy adult female Jebeers weighting about 20-25kg and aging 2-2.5 years were used in this study. Each Jebeer was given intramuscular Atropine ¹ (0.4 mg/kg) and anesthetized by intravenous injection of mixed Ketamine ² (2.2 mg/kg) and Xylazine ² (0.11mg/kg). The jebeers were positioned in sternal recumbency and X-ray radiation was adjusted by an angle of 90 degrees to the longitudinal axis of the trunk. Tomograms were acquired at a thickness of 3m1m using a third generation CT scanner with dynamic scanning capabilities, (Toshiba Xvision EX, Japan). The acquisition parameters were as follows: KVp 120, mA 100 and scan–time of 1-S. Window width and level were adjusted as necessary to obtain the optimal image of the abdominal organs (WW: 415 and WL: 28).

Following image acquisition, two goats were euthanized with an overdose of pentobarbital while still under anesthesia. CT image were labeled by comparison with dissected abdomen of the euthanized goats. Thoracic and lumbar

^{2.} Alfasan Holland



Figure 1. 1.8th thoracic vertebra, 2. epaxial muscles, 3. Left Lung, 4. Reticulum, 5. base of heart, 6. internal thoracic vessels, 7. 8th rib, 8. right lung, 9. aorta, 10. esophagus, 11. caudal vena cava, 12. liver, 13. sternum, 14. pectoral muscles.

vertebrae were used as landmarks to describe the location and extension of the abdominal organs.

3. Results

In Figures 1 to 12, CT images, which have been selected from one Jebeer, are viewed from cranial to caudal. Transverse images have been presented how the left and dorsal aspects of the animal are in the left and top sides of the images respectively. The selected images have been achieved from each adjacent vertebra, from 8th thoracic to 6th lumbar vertebrae. Then, the positions of abdominal viscera were evaluated and determined according to the adjacent vertebrae (Table 1).

The esophagus (Figure 1 to 3) passed through diaphragm and immediately attached to the region between rumen and reticulum, this attachment was at the level of the 10th thoracic vertebra (Figure 3). The liver (Figure 1 to 8) was observed in the right side and the most cranial part of the abdominal cavity from 8th thoracic vertebra to the cranial extremity of the 1st lumbar vertebra, the gall bladder was also observed between 13th thoracic and 1st lumbar vertebrae (Figure 6 to 7). The spleen situated in the left side of the abdominal cavity and located over the dorsal sac of rumen from 9th thoracic vertebra to 2nd lumbar vertebra (Figure 2 to 8). The reticulum was seen at the cranial part of the abdominal cavity adjacent to the diaphragm between 8th and 9th thoracic vertebra (Figure 1 & 2).

Atrium ruminis, the cranial sac of rumen (Figure 3 to 5) was recognized between 10th and 12th thoracic vertebrae. It continued caudally with the dorsal sac of rumen which was observed between 13th thoracic to 3rd lumbar vertebrae



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Figure 2. 1. 9th thoracic vertebra, 2. 9th rib, 3. left lung, 4. 8th rib, 5. spleen, 6. reticulum, 7. base of heart, 8. internal thoracic vessels, 9. right lung, 10. aorta, 11. esophagus, 12. caudal vena cava, 13. liver, 14. pectoral muscles.

^{1.} Daroopakhsh, Iran

	T8	т9	T10	T11	T12	T13	L1	L2	L3	L4	L5	L6
Liver	+	+	+	+	+	+	+	+				
Spleen		+	+	+	+	+	+	+				
Esophagus	+	+	+									
Reticulum	+	+										
Omasum			+	+	+	+	+	+				
Abomasum			+	+	+	+	+	+				
Left lung	+	+	+									
Right lung	+	+	+	+	+	+	+	+				
Atrium ruminis			+	+	+							
Recessus Ruminis			+									
Dorsal sac of rumen						+	+	+	+			
Caudodorsal blind sac										+	+	
Caudoventral blind sac										+	+	+
Ventral sac of rumen				+	+	+	+	+	+			
Omasoabomasal Opening				+	+							
Right kidney						+	+	+				
Left kidney								+	+	+		
Urinary bladder												+
Uterine horns												+
Gall bladder						+	+					
Pancreas							+	+				
Duodenum				+	+	+	+	+				
Transverse colon									+			
Descending colon									+	+	+	+
Spiral colon									+	+	+	
Ascending colon					+	+	+	+	+	+	+	
Cecum									+	+	+	+
Jejunum						+	+	+	+	+	+	+

Table 1. Topographical position of major abdominal organs according to the thoracic and lumbar vertebrae.

(Figure 6 to 9). The dorsal sac of rumen also continued with the caudodorsal blind sac between $4^{th}\,and\,5^{th}$ lumbar

vertebrae (Figure 10 to 11). The ventral sac of rumen located between 11^{th} thoracic to 3^{rd} lumbar vertebrae (Figure



Figure 3. 1.10th thoracic vertebra, 2. 10th rib, 3. left lung, 4. 9th rib, 5. spleen, 6. esophagus, 7. 8th rib, 8. atrium ruminis, 9. recessus ruminis, 10. base of abomasum, 11. epaxial muscles, 12. right lung, 13. aorta, 14. caudal vena cava, 15. omasum, 16. liver, 17. pectoral muscles.

4 to 9). The most cranial part of this sac, recessus ruminis, was observed at the level of 10^{th} thoracic vertebra (Figure 3). The ventral sac also replaced with the caudoventral blind sac and continued to the 6th lumbar vertebra (Figure 10 to 12). The omasum was ellipsoidal in form and situated at the right side of the abdominal cavity between 10^{th} thoracic to 2^{nd} lumbar vertebrae (Figure 3 to 8). Fundus, body and pyloric part of the abomasum were seen at the same level of the omasum (Figure 3 to 8). The right kidney began from the 13^{th} thoracic vertebra and extended to the 2^{nd} lumbar vertebra (Figure 6 to 8). On the contrary, the



Figure 5. 1. 12th thoracic vertebra, 2. epaxial muscles, 3. spleen, 4. atrium ruminis, 5. cranial pillar, 6. base of abomasum, 7. 12th rib, 8. right lung, 9. aorta, 10. caudal vena cava, 11. ascending colon, 12. cranial part of duodenum, 13. liver, 14. omasum, 15. omasoabomasal opening, 16. ventral sac of rumen, 17. body of abomasum.



Figure 4.1.11th thoracic vertebra, 2.11thrib, 3. spleen, 4. atrium

rugine 4: 1: 11 utoracte verteora, 2: 11 110, 5: spiceri, 4: athun ruminis, 5: cranial pillar, 6: base of abomasum, 7: epaxial muscles, 8: right lung, 9: aorta, 10: caudal vena cava, 11: liver, 12: cranial part of duodenum, 13: omasum, 14: omasoabomasal opening, 15: ventral sac of rumen, 16: body of abomasum, 17: pectoral muscles.

left one was seen between the 2^{nd} to 4^{th} lumbar vertebrae (Figure 8 to 10).

Pancreas (Figure 7 & 8) was seen in the right side of the abdominal cavity from 1st to 2nd lumbar vertebrae. The pyloric sphincter was observed in the right side of the abdominal cavity between 13th thoracic to 2nd lumbar vertebrae (Figure 6 to 8), it was continued by the cranial part of duodenum (Figure 4 to 8) between 11th thoracic to 2nd



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Figure 6. 1. 13th thoracic vertebra, 2. epaxial muscles, 3. 13th rib, 4. spleen, 5. aorta, 6. 12th rib, 7. 11th rib, 8. dorsal sac of rumen, 9. 10th rib, 10. cranial pillar, 11. 8th rib, 12. costal arch, 13. right lung, 14. cranial pole of right kidney, 15. caudal vena cava, 16. liver, 17. ascending colon, 18. gall bladder, 19. cranial part of duodenum, 20. omasum, 21. ventral sac of rumen, 22. pylorus spinchter, 23. pyloric part of abomasum, 24. base of abomasum, 25. body of abomasum



Figure 7. 1. 1st lumbar vertebra, 2. spleen, 3. aorta, 4. caudal vena cava, 5. dorsal sac of rumen, 6. cranial pillar, 7. ventral sac of rumen, 8. base of abomasum, 9. epaxial muscles, 10. right lung, 11. right kidney, 12. ascending colon, 13. pancreas, 14. gall bladder, 15. liver, 16. cranial part of duodenum, 17.omasum, 18. pylorus spinchter, 19. pyloric part of abomasum, 20. body of abomasum.

lumbar vertebrae. The ascending colon was recognized ventral to the right kidney (Figure 6 to 8) and dorsal to the omasum and the cranial part of duodenum (Figure 5 to 8), it was started at the level of the caudal part of 11th and the cranial part of 12th thoracic vertebrae (Figure 4 to 5) and continued at the right side of abdominal cavity to the level of 5th lumbar vertebra (Figure 11). The cecum was started at the level of 3rd lumbar vertebra (Figure 9) and continued caudally to the pelvic cavity. The transverse colon (Figure



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Figure 9.1.3rd lumbar vertebra, 2. epaxial muscles, 3. sublumbar muscles, 4. caudal vena cava, 5. aorta, 6. dorsal sac of rumen, 7. right and left longitudinal pillars, 8. ventral sac of rumen, 9. spiral part of ascending colon, 10. descending colon, 11. transverse colon, 12. proximal and distal part of ascending colon, 13. left kidney, 14. cecum, 15. jejunum.



Figure 8. 1. 2nd lumbar vertebra, 2. epaxial muscles, 3. spleen, 4. aorta, 5. caudal vena cava, 6. dorsal sac of rumen, 7. cranal pillar, 8. ventral sac of rumen, 9. base of abomasum, 10. right lung, 11. right kidney, 12. ascending colon, 13. liver, 14. pancreas, 15. cranial part of duodenum, 16. omasum, 17. pylorus spinchter, 18. pyloric part of abomasum, 19. body of abomasum, 20. base of abomasum.

9) crossed the abdominal cavity from the right to the left side where the descending colon was started (Figure 9 to 12) and continued caudally to the pelvic cavity. The uterine horns and the urinary bladder were seen at the level of 6th lumbar vertebra (Figure 12).

According to the diaphragmatic convexity, CT images of the intra thoracic part of the abdominal cavity illustrated several thoracic organs beside the intra thoracic organs of the abdominal cavity, such as the right lung (Figure 1 to 8),



Figure 10. 1. 4th lumbar vertebra, 2. epaxial muscles, 3. sublumbar muscles, 4. caudodorsal blind sac, 5. caudal pole of left kidney, 6. caudal pillar, 7. spiral part of ascending colon, 8. caudoventral blind sac, 9. caudal vena cava, 10. aorta, 11. descending colon, 12. ascending colon, 13. cecum, 14. jejunum.



Figure 11. 1. 5th lumbar vertebra, 2. epaxial muscles, 3. sublumbar muscles, 4. aorta, 5. caudodorsal blind sac, 6. caudal pillar, 7. caudoventral blind sac, 8. caudal vena cava, 9. descending colon, 10. ascending colon, 11. spiral part of ascending colon, 12. cecum, 13. jejunum.

the left lung (Figure 1 to 3), the base of heart (Figure 1 to 2) and the thoracic part of esophagus (Figure 1 to 2).

4. Discussion

Although the conventional radiology and ultrasonography are more practical and easily performed procedures, CT scan is superior for the soft tissue differentiation [29, 30]. It has been effectively used to evaluate the abdominal cavity of the domestic animals [16, 20, 31]. CT also enables us to obtain a quick and accurate image of internal organs of those animals which routine invasive anatomic studies were not recommended on them [32, 33]. It is slightly difficult to do anatomic studies on the Jebeer as an endangered wild ruminant. In this study CT modality make us possible to evaluate the abdominal organs of the Jebeer anatomically without euthanasia or any other invasive procedure.

In order to have the reproducibility in the normal CT anatomic studies, consistency of the CT performance parameters are very important. Any deviation in the angle of radiation to the body axis, results in the appearance of various organs in cross sectional CT images, so it is important to use a proper and standard angle of radiation. This angle is 90° to the longitudinal axis of the body and has used in many normal CT anatomic studies which aim to generally demonstrate the body organs [17, 27, 29, 30]. In the present study the radiation was performed vertical to the longitudinal axis of the standard CT images demonstrating the position of the abdominal organs and their vicinities accurately.



Figure 12. 1. 6th lumbar vertebra, 2. wing of Ilium, 3. sublumbar muscles, 4. descending colon, 5. caudoventral blind sac, 6. uterine horn, 7. cecum, 8. urinary bladder, 9. jejunum.

In those CT anatomic studies which no landmark were offered [14, 20], an organ only could be roughly addressed, but in the present study we used the thoracic and lumbar vertebrae as references to describe the topographic position of the abdominal organs. Relative anatomic position of the body organs to the respective vertebrae may be due to the segmental formation of the body during the embryonic period [34] and has been used in other studies for precise description of the location of the internal organs [16, 19, 27, 30].

In this manner we have prepared a sectional imaging anatomic atlas of the abdominal organs in the Jebeer. By this atlas one can find out the situation and extension of these organs according to the vertebral column (Table 1).

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References

- Pather AB, Berry CR, Thrall DE. Use of radiography in combination with computed tomography for the assessment of noncardiac thoracic disease in the dog and cat. Veterinary Radiology & Ultrasound. 2005; 46(2):114-121.
- [2] Diana A, Pivetta M, Cipone M. Imaging evaluation of the small animal mediastinum. Veterinary Research Communication. 2006; 30(1):145-151.

- [3] Hahn KA, Lantz GC, Salisbury SK, Blevins WE, Widmer WR. Comparison of survey radiography with ultrasonography and x-ray computed tomography for clinical staging of subcutaneous neoplasms in dogs. Journal of the American Veterinary Medical Association. 1990; 196(11): 1795-1798.
- [4] Olby NJ, Muntana KR, Sharp NJH, Thrall DE. The computed tomographic appearance of acute thoracolumbar intervertebral disc herniations in dogs. Veterinary Radiology & Ultrasound. 2000; 41(5):396-402.
- [5] Patsikas MN, Rallis T, Kladiks SE, Dessiris AK. Computed tomography diagnosis of isolated splenic torsion in dog. Veterinary Radiology & Ultrasound. 2001; 42(3): 235-237.
- [6] Heninger W. Use of computed tomography in the diseased feline thorax. Journal of Small Animal Practice. 2003; 44(2): 56-64.
- [7] Takahashi A, Yamada K, Kishimoto M, Shimizu J, Maeda R. Computed tomography (CT) observation of pulmonary emboli caused by long-term administration of ivermectin in doges experimentally infected with heartworms. Veterinary Parasitology. 2008; 155(3-4):242-248.
- [8] Regodon S, Franco A, Garin JM, Romina A, Lignereux Y. Computerized tomographic determination of the cranial volume of the dog applied to racial sexual differentiation. Acta Anatomica. 1991; 142(4):347-350.
- [9] Robina A, Regodon S, Guillen MT, Lignereux Y. Utilization of computerized tomography for the determination of the volume of the cranial cavity of the Galgo Hound. Acta Anatomica. 1991; 140(2):108-111.
- [10] Arencibia A, Vazuez JM, Ramirez JA, Sandoval JA, Ramirez G, Sosa C. Anatomy of the craniocephalic sructures of the goat (Capra Hircus L.) by imaging techniques: A computerized tomographic study. Anatomia Histologia Embryologia. 1997; 26(3): 161-164.
- [11] Walker NE, Olszewski ME, Wahle A, Measurement of coronary vasoreactivity in sheep using 64-slice multidetector computed tomography and 3-D segmentation. International Congress Series 1281. The University of Iowa, United States; 2005, pp: 1304.
- [12] Shojaei B, Nazem MN, Vosugh D. Anatomic reference for computed tomography of the paranasal sinuses and their openings in the rayini goat. Iranian Journal of Veterinary Surgery. 2008; 3(2):83-92.
- [13] Onar V, Kahvecioglu KO, Cebi V. Computed tomographic analysis of the cranial cavity and neurocranium in the German shepherd dog (Alsatian) puppies. Veterinarski Arhiv. 2002; 72(2):57-66.
- [14] Cardoso L, Gil F, Ramirez G, Teixeira MA, Agut A, Rivero MA, Arencibia A, et al. Computed tomography (CT) of the lungs of the dog using a helical CT scanner ,intravenous iodine contrast medium and different ct windows. Anatomia Histologia Embryologia. 2007; 36(5):328-331.
- [15] Smallwood JE, George II TF. Anatomic atlas for computed tomography in the mesaticephalic dog: thorax and cranial abdomen. Veterinary Radiology and Ultrasound. 1993; 34(3):65-84.
- [16] Shojaei B, Vajhi AR, Rostami A, Molaei MM, Arashian I, Hashemnia S. Computed tomographic anatomy of the

abdominal region of cat. Iranian Journal of Veterinary Research. 2006; 7(2):45-52.

- [17] Shojaei B, Rostami A, Vajhi A, Shafaei M. Computed tomographic anatomy of the thoracic region of the cat. Veterinarski Arhiv. 2003; 73(5):261-269.
- [18] Vladova D, Stefanov M, Toneva Y. Computed tomography study of thoracic aorta in the cat. Bulgarian Journal of Veterinary Medicine. 2005; 8(3):151-156.
- [19] Vladova D, Toneva Y, Stefanov M. Computed tomography (CT) of the cranial mediastinum in the cat. Trakia Journal of Sciences 2005; 3(1):53-57.
- [20] Samii VF, Biller DS, Koblik PD. Normal cross-sectional anatomy of the feline thorax and abdomen: comparison of computed tomography and cadaver anatomy. Veterinary Radiology and Ultrasound. 1998; 39(6):504-511.
- [21] Braun U, Irmer M, Augsburger H, Jud R, Ohlerth S. Computed tomography of the abdomen in saanen goats. Schweizer Archiv fur Tierheilkunde. 2011; 153(7):314-320.
- [22] Ohlerth S, Becker Birck M, Augsburger H, Jud R, Makara M, Braun U. Computed tomopraphy measurements of thoracic structures in 26 clinically normal goats. Research in Veterinary Science. 2012; 92(1):7-12.
- [23] Alsafy MAM. Computed tomography and cross -sectional anatomy of the thorax of goat. Small Ruminant Research. 2008; 79(2-3):158-166.
- [24] Groves CP. The chinkara (Gazella Bennetti) in Iran, with the description of two new subspecies. Journal of Sciences Islamic Republic of Iran. 1993; 4(3):166-178.
- [25] Ziaie H. A field guide to the mammals of Iran. 2nd ed. Department of the Environment, wildlife center, Tehran; 2008, pp: 357-358.
- [26] Hemami MR, Groves CP. Global survey and regional action plans. In: Mallon Dp, Kingswood SC, editors. Antelopes: part 4: North Africa, the Middle East and Asia. 1st ed. IUCN; 2001, pp: 114-118.
- [27] Sajjadian SM, Shojaei B, Molaei MM. Computed tomographic anatomy of the bronchial tree of the jebeer gazelle. Iranian Journal of Veterinary Surgery. 2008; 3(1):73-80.
- [28] Shojaei B, Sajjadian SM. Computed tomographic anatomy of the nasal cavity and paranasal sinuses of the jebeer. Iranian Journal of Veterinary Surgery. 2008; 3(4):75-84.
- [29] Alsafy MAM, El-Gendy SAA, EL-kammer MH, Ismaiel M. Contrast radiographic, ultrasounographic and computed tomographic imaging studies on the abdominal organs and fatty liver infiltration of zaraibi goat. Journal of Medical Sciences. 2013; 13(5):316-326.
- [30] Shojaei B, Vosough D, Sharifi F. Computed tomographic anatomy of the thoracic cavity vessels in the rayini goat. Iranian Journal of Veterinary Surgery. 2012; 7(1-2):9-22.
- [31] 31-Smallwood JE, George TF. Anatomic atlas for computed tomography in the mesaticephalic dog: caudal abdomen and pelvis. Veterinary Radiology and Ultrasound. 1993; 34(3):143-167.
- [32] Alonso-Farre JM, Gonzalo-Orden M, Barreiro-Vazquez JD, Ajenio JM, Barreiro-Lois A, Llarena-Reino M, Degol-

lada E, et al. Cross-sectional anatomy, computed tomography and magnetic resonance imaging of the thoracic region of common dolphin (Delphinus Delphis) and striped dolphin. Anatomia Histologia Embryologia. 2014; 43(3):221-229.

- [33] Frey R, Gebler A. The highly specialized vocal of the male mongolian gazelle (procapra gutturosa pallas, 1777-mammalia, Bovidae). Journal of Anatomy. 2003; 203(5):451-471.
- [34] McGeady TA, Quinn PJ, Fitzpatrick ES, Ryan MT, Cahalan S. Veterinary Embryology. 1st ed. Blackwell Pub; 2006, pp: 220-221.