Evaluation of specific biochemical indicators of Helicobacter pylori-associated gastric cancer in Egypt

M.M. Anwar, A.I. Youssef, M.I. Sheta, A. Zaki, N.R. Bernaba and M.A. El-Toukhi

ABSTRACT The aim of the study was to assess the accuracy of some specific biochemical indicators in discriminating between Helicobacter pylori-associated gastritis and H. pylori-associated stomach cancer (serum gastrin level, serum soluble E-cadherin and tissue COX-2 activity, as well as serodiagnostic markers for H. pylori infection) in order to find a simple diagnostic test that can reasonably predict the development of gastric cancer. The study participants comprised 20 patients with gastric carcinoma, 20 patients with positive H. pylori-associated gastritis and 20 individuals as the control group. Standard procedures and quality control measures were followed. Using cut-off values and ROC analysis to assess the diagnostic abilities of the biochemical indicators, E-cadherin showed the highest sensitivity (100%). We suggest that close follow-up together with periodic endoscopic examination for all patients with persistent H. pylori infection and serum soluble E-cadherin level above 5 µg/mL is essential.

RÉSUMÉ La présente étude visait à évaluer l'exactitude de certains indicateurs biologiques spécifiques (gastrinémie, concentration sérique de la E-cadhérine soluble, activité tissulaire de la cyclo-oxygénase 2, et marqueurs sérologiques d'une infection à Helicobacter pylori) permettant de différencier une gastrite associée à H. pylori d'un cancer de l'estomac associé à cette bactérie, afin de déterminer un test diagnostique simple capable de prédire raisonnablement l'apparition d'un cancer de l'estomac. L'étude portait sur 20 patients atteints d'un carcinome gastrique, 20 patients souffrant d'une gastrite à H. pylori et 20 personnes en tant que groupe témoin. Des procédures normalisées ont été suivies et des mesures de contrôle de la qualité ont été effectuées. Après l'utilisation de valeurs seuils et d'une analyse de la fonction d'efficacité du récepteur pour évaluer les qualités diagnostiques des indicateurs biologiques, la E-cadhérine a montré la sensibilité la plus élevée (100 %). Nous suggérons qu'un suivi attentif et un examen endoscopique régulier sont essentiels pour tous les patients atteints d'une infection à H. pylori persistante et présentant une concentration sérique de la E-cadhérine soluble supérieure à 5 µg/ml.

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Introduction

Gastric cancer is the fourth commonest malignant disorder and the second commonest cause of cancer-related death worldwide [1]. It represents 1.64% of all cancers in Egypt, according to the National Cancer Institute registry and the median age is 55 years with male predominance [2]. Several studies have linked Helicobacter pylori and gastric cancer. The etiopathogenetic cascade of the infection leads to various errors in the genome of dividing gastric epithelial cells, which leads to abnormal differentiation of cells [3]. In addition, chronic inflammation associated with the infection leads to damage in proteins, the production of reactive oxygen species and deficient repair of replication errors of DNA. All increase the risk of gastric cancer [3,4].

Among the pathogenic virulence factors in H. pylori infection is CagA protein, which is produced by most strains into the gastric cells and phosphorylated by the host kinases, resulting in morphological changes in the epithelial cells [5]. CagA positive H. pylori infection up-regulates the expression of the enzyme cyclo-oxygenase-2 (COX-2), which converts arachidonic acid to prostaglandin E2 [6]. The angiogenic-enhancing role of COX-2 facilitates the process of invasion and metastasis through decreasing the expression of the adhesion molecule E-cadherin. E-cadherin is expressed by epithelial cells and is responsible for homotypic cell–cell adhesion. Degradation of tissue E-cadherin produces serum soluble E-cadherin, which is greatly elevated in inflammatory conditions by mediators, or in cancerous disease due to the over-expression of proteases [7]. As the initial stage of H. pylori infection is an acute gastritis, so the acid response of the stomach is affected, and gastrinemia is induced; this is associated with the occurrence of gastric adenocarcinoma [8].

Researchers have documented that H. pylori eradication therapy attenuates the endoscopic and histological lesions [9]. Besides, many studies have consistently reported that H. pylori eradication can lead to a regression of acute gastritis and intestinal metaplasia, and that such intervention could reduce the risk for adenocarcinoma of the distal stomach [10,11].

When clinically manifested, gastric cancer has an extremely poor prognosis since the 5-year survival rate using currently available treatments, surgery and radio-chemotherapy, is less than 20% [12]. Therefore a potential benefit could be expected if we had a feasible diagnostic test for follow-up of patients with H. pylori gastritis and were able to predict the risk of development of stomach cancer.

In this study, we aimed to assess the accuracy of specific biochemical indicators to discriminate between H. pylori-associated gastritis and H. pylori-associated stomach cancer.

Methods

Patients and samples

Twenty consecutive adult patients attending the Endoscopy Unit, Department of Surgery, Medical Research Institute in the University of Alexandria during the period January 2008–June 2009 and proven by endoscopic, pathological and radiological examinations to have an operable stomach cancer were involved in the study. Sample size was restricted owing to limited time resources. None of the patients had received chemotherapy before surgical operation. For each patient with stomach cancer, the next adult patient with endoscopic diagnosis of H. pylori-associated gastritis, free of any malignant changes, was recruited in the study. In addition, for each patient with stomach cancer, the next age-matched (within 5 years) individual presenting with epigastric pain which proved to have normal endoscopic findings and who was negative for H. pylori was included in the study as part of the control group. All individuals in the study sample were subjected to routine history and clinical examination, followed by an upper gastrointestinal endoscopic examination using a long forward viewing instrument (Fuji, EG 250D video). The stomach was completely examined for the presence or absence of any gross pathological sign.

All 60 contributors were subjected to multiple endoscopic gastric mucosal tissue biopsies. These were examined pathologically to confirm the presence or absence of gastritis and/or malignancy and for biochemical determination of tissue COX-2 activity using COX Activity Assay Kit (Cayman Chemical Company, Cat. No. 760151). The COX activity assay utilizes the peroxidase component of cyclo-oxygenases. Peroxidase activity was assayed colorimetrically by monitoring the appearance of oxidized N,N,N′,N′-tetramethyl-p-phenylenediamine at 590 nm [13]. The assay included COX-1 and COX-2 specific inhibitors in order to distinguish between the 2 isozymes.

Serum gastrin level was measured using Gastrin-17 ELISA kit (Biohit Plc, Cat. No.601035). The G-17 ELISA is based on a sandwich enzyme immunoassay technique with a G-17 specific capture antibody adsorbed to a micro-well plate and a detection antibody labelled with horseradish peroxidase [14]. Serum soluble E-cadherin was measured using the Human E-cadherin EIA kit (Takara Bio Inc., Code No. MK117). The human E-cadherin EIA kit is a solid phase EIA based on a sandwich method that utilizes 2 mouse monoclonal anti-human E-cadherin antibodies to detect soluble E-cadherin in a 2-step procedure [15].
H. pylori Ag ELISA (ABC Diagnostics, New Damietta) was used to determine serum H. pylori antigen level; determination of serum H. pylori CagA antibodies was done using ELISA H. pylori CagA IgG Quantitative (RightChoice Diagnostics, code REP1004), while H. pylori IgG antibody was measured using H. pylori IgG Quantitative ELISA Kit (MP Biomedicals, LLC, Cat. No. 07BC1052). All steps were carried out according to the manufacturer’s instructions [16–18].

Quality control
All the steps for the blood and tissue sampling were done under the authors’ supervision and followed standard procedures. All tests were performed in duplicate. All individuals agreed to participate after the aims of the study were explained to them and gave informed consent.

The study was approved by the institutional ethical committee of the Medical Research Institute, Alexandria University.

Histopathological examination
After the surgical management for patients with stomach cancer was done, the gastric specimens were fixed into 10% formalin and routinely processed for pathological examination. Paraffin wax-embedded sections were cut into 3–5 µ thick serial sections and stained with haematoxylin and eosin. Gastric cancer was classified according to Laurén’s classification [19], which divides gastric adenocarcinoma into intestinal, diffuse and mixed types.

Statistical analysis
Data analysis was conducted using SPSS, version 17.0. Descriptive statistics were presented as mean [standard deviation (SD)] for quantitative data and frequency for qualitative data. Analysis of variance (ANOVA) was used to test the significance of the mean differences in serum gastrin level, serum soluble E-cadherin, CagA antibody, and tissue COX-2 activity between the 3 different groups. Unpaired t-test with Bonferroni correction for multiple comparisons was used to test the significance of differences in mean H. pylori antigen, H. pylori IgG antibody and H. pylori CagA antibody. Due to the small sample size in the subgroups of stomach cancer patients, a non-parametric test (Mann–Whitney) was used. P-value < 0.05 was considered statistically significant.

The ROC analysis was used to assess the diagnostic abilities of the biochemical indicators in the discrimination between patients with H. pylori gastritis and those with stomach cancer. The cut-off point which optimizes the accuracy of the diagnostic test was defined as the point which maximizes the value of the Youden index (sensitivity + specificity – 1).

Results
Patients were divided into 3 groups, each with 20 patients, group I (control group), group II (patients with H. pylori-associated gastritis), and group III (patients with stomach cancer). The distribution of sex and age of all the participants is shown in Table 1.

Endoscopic, surgical and histopathological findings
In group II patients, pathological examination of the gastric tissue biopsies showed 5 cases with superficial gastritis, 4 cases with atrophic gastritis, 2 cases with gastrooduodenitis, and 9 cases with benign gastric ulcer.

In group III patients, all endoscopic malignant changes were localized in the antrum and prepyloric region except in 2 patients where cancer was localized in the body of the stomach as malignant ulcers and polyoidal mass. After confirmation of the malignant nature of the disease by pathological examination of the endoscopic biopsies, 14/20 (70%) underwent subtotal gastrectomy and 6/20 (30%) underwent total gastrectomy. After surgery, the pathological findings were as follows: 10 cases (50%) were of intestinal type adenocarcinoma, 7 (35%) were diffuse type adenocarcinoma and 3 (15%) were of the mixed type. Table 2 shows the histopathological type of operated gastric cancer patients. None of the gastric cancer patients included in this study was free from H. pylori infection.

Biochemical indicators
All the differences in mean serum gastrin level, tissue COX-2 activity, serum E-cadherin and CagA antibody between the study groups were highly significant (P < 0.01). Patients with stomach cancer had the highest in serum gastrin levels, tissue COX-2 activity and serum E-cadherin followed by the group with positive H. pylori-associated gastritis (Table 3). For CagA antibody,
the group with \textit{H. pylori} gastritis showed the highest level.

Considering the levels of the studied biochemical indicators in the 3 pathologic subtypes of gastric cancer patients, serum gastrin and tissue COX-2 activity were significantly higher in the intestinal subtype compared with the diffuse type ($P < 0.05$), while serum \textit{E-cadherin} was significantly higher in the diffuse type (Table 4). Intermediate levels were seen in the mixed type.

ROC analysis was adopted to identify the cut off points for serum gastrin level, tissue COX-2 activity, and serum \textit{E-cadherin}, which maximizes the overall accuracy of these biochemical indicators in the discrimination between individuals with \textit{H. pylori} infection with or without stomach cancer. The corresponding sensitivity and specificity of each cut-off point was relatively high, ranging from 80\% to 100\%. \textit{E-cadherin} had the greatest sensitivity (100\%) in detecting stomach cancer (Table 5).

Discussion

In our study all patients with stomach cancer were positive for \textit{H. pylori}. The significantly higher levels in \textit{H. pylori} antigens and \textit{H. pylori} IgG and \textit{CagA} antibodies when comparing each of the infected groups with the control group explain the evidence that \textit{CagA} positive strains of \textit{H. pylori} are considered virulent and have a highly pathogenic effect on gastric mucosa and that they have been related to ulcers and greatly increase the risk of gastric cancer [20,21]. In our work, the levels of \textit{H. pylori} \textit{CagA} antibodies in the gastric cancer group were about 19\% lower than those in

### Table 2 Histopathology and type of surgery for the 20 patients with stomach cancer

<table>
<thead>
<tr>
<th>Type of gastrectomy</th>
<th>Intestinal ($n = 10$)</th>
<th>Histopathological adenocarcinoma type</th>
</tr>
</thead>
<tbody>
<tr>
<td>No.</td>
<td></td>
<td>No.</td>
</tr>
<tr>
<td>Subtotal</td>
<td>9</td>
<td>2</td>
</tr>
<tr>
<td>Total</td>
<td>1</td>
<td>5</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Type of gastric cancer</th>
<th>Diffuse ($n = 7$)</th>
<th>Intestinal ($n = 10$)</th>
<th>Mixed ($n = 3$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
</tr>
<tr>
<td>Serum gastrin (pmol/L)</td>
<td>24.0 (1.3)</td>
<td>33.8 (1.2)*</td>
<td>33.7 (1.5)</td>
</tr>
<tr>
<td>Tissue COX-2 activity (U/mL)</td>
<td>880 (87)</td>
<td>1297 (440)*</td>
<td>999 (271)</td>
</tr>
<tr>
<td>Serum \textit{E-cadherin} (µg/mL)</td>
<td>7.80 (0.30)</td>
<td>5.50 (0.11)*</td>
<td>6.45 (0.01)</td>
</tr>
</tbody>
</table>

*All between-group comparisons were statistically significant ($P < 0.05$).

ANOVA test.

Group I = control group; group II = patients with \textit{H. pylori}-associated gastritis; group III = patients with stomach cancer.

SD = standard deviation.

### Table 3 Levels of serum gastrin, tissue COX-2 activity, serum \textit{E-cadherin}, and \textit{CagA} antibodies (Ab) in the 3 study groups

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Group I* ($n = 20$)</th>
<th>Group II* ($n = 20$)</th>
<th>Group III* ($n = 20$)</th>
<th>$P$-value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum gastrin (pmol/L)</td>
<td>5.3 (1.4)</td>
<td>16.6 (3.8)</td>
<td>30.3 (5.9)</td>
<td>&lt; 0.01</td>
</tr>
<tr>
<td>COX-2 activity (U/mL)</td>
<td>414.9 (87.4)</td>
<td>632.8 (122.7)</td>
<td>1106.4 (298.5)</td>
<td>&lt; 0.01</td>
</tr>
<tr>
<td>Serum \textit{E-cadherin} (µg/mL)</td>
<td>1.9 (0.3)</td>
<td>4.3 (0.7)</td>
<td>6.4 (1.2)</td>
<td>&lt; 0.01</td>
</tr>
<tr>
<td>Serum \textit{CagA}-Ab (Uarb/mL)</td>
<td>2.2 (1.0)</td>
<td>64.1 (14.3)</td>
<td>52.4 (11.0)</td>
<td>&lt; 0.01</td>
</tr>
</tbody>
</table>

*All between-group comparisons were statistically significant ($P < 0.05$).

ANOVA test.

Group I = control group; group II = patients with \textit{H. pylori}-associated gastritis; group III = patients with stomach cancer.

SD = standard deviation.

### Table 4 Mean levels of the studied parameters in different pathological types of gastric cancer

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Diffuse ($n = 7$)</th>
<th>Intestinal ($n = 10$)</th>
<th>Mixed ($n = 3$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
</tr>
<tr>
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<td>24.0 (1.3)</td>
<td>33.8 (1.2)*</td>
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<tr>
<td>Serum \textit{E-cadherin} (µg/mL)</td>
<td>7.80 (0.30)</td>
<td>5.50 (0.11)*</td>
<td>6.45 (0.01)</td>
</tr>
</tbody>
</table>

*Significant at $P < 0.05$.

Mann–Whitney test was used for comparison between diffuse and intestinal type. No statistical tests were applied to the mixed type owing to the very small sample size.

SD = standard deviation.
the *H. pylori*-associated gastritis group, while, *H. pylori* IgG antibodies were about 26% lower. The significant difference in these parameters was similar to the results obtained by Meimarakis et al., who found that there was a spontaneous decline in *H. pylori* antibody as gastric carcinogenesis proceeds, and that CagA antibody persists longer in serum that *H. pylori* IgA antibody [22]. This also may be explained by the idea that in patients with severe atrophic gastritis, *H. pylori* infection may be difficult to demonstrate because the bacteria disappear spontaneously and even *H. pylori* antibody titres eventually decline to normal [23].

The availability of a simple diagnostic test which can reasonably predict the development of stomach cancer could be of high practical importance. The most impressive finding in this study was the striking differences in the levels of serum gastrin, COX-2 activity, serum soluble E-cadherin and CagA antibody in patients with *H. pylori* gastritis compared with healthy control individuals. A further striking increase in these biochemical parameters is found in individuals with *H. pylori*-associated gastric cancer. The reasons for that need some explanation.

Gastrin is involved in the tumorgenesis in the gastrointestinal tract [24], and plays a key role in the initiation and the progression of cancer through the adenoma-carcinoma sequence in the stomach [25]. Recent studies showed a significantly higher level in *H. pylori*-infected patients than the seronegative ones [26], which is secondary to *H. pylori* colonization of the gastric body and fundus, resulting in decreased acid secretion and thus lowering the inhibitory feedback on gastrin release [27]. Also, *H. pylori* infection plays a significant role in the stimulation of G cells [28]. This explains the progressively significantly higher serum gastrin levels in groups II and III.

The statistically significant elevation of serum gastrin level in group III, especially in the intestinal and mixed forms, compared with patients in group II led us to believe like other researchers that gastrin could originate from extra-G-cell sources, including cancer cells themselves [2]. Also, a strong positive correlation between serum gastrin and each of *H. pylori* IgG and CagA antibodies confirms that hypergastrinaemia and COX-2 activity may be indicators for the development of stomach cancer [29].

The roles of *H. pylori* infection and gastrin have been reported to stimulate and induce overexpression of COX-2 [30]; this explains the increase in tissue COX-2 activity in the cancer group being *H. pylori* positive compared to group II. This increase may inhibit apoptosis and increase the invasiveness of malignant cells [31]. The highest activity for COX-2 was found in patients with the intestinal subtype of stomach cancer and this finding is in agreement with previous studies which revealed that COX-2 expression may be associated with the carcinogenesis of the intestinal type of gastric cancer and, speculatively, inhibition of COX-2 might have preventative effects on the intestinal type of gastric cancer. Also, in tumours of mixed type, COX-2 is increased in the intestinal component compared to the diffuse one [32].

Conditions with rapid cell turnover or inflammation lead to an increase in serum level of E-cadherin [7]. Perturbation in the expression or function of the transmembrane glycoprotein leading to an increase in permeability mediated by the reduction in cell adhesion might allow *H. pylori* antigens to reach the gastric lamina propria with resultant

### Table 5

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Cut-off</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>E-cadherin (µg/mL)</td>
<td>5</td>
<td>100</td>
<td>80</td>
</tr>
<tr>
<td>COX-2 (U/mL)</td>
<td>755</td>
<td>85</td>
<td>90</td>
</tr>
<tr>
<td>Gastrin (pmol/L)</td>
<td>26</td>
<td>80</td>
<td>100</td>
</tr>
</tbody>
</table>

*All between-group comparisons were significant (\(P < 0.05\)) using Bonferroni correction.*

**Table 6 Mean levels of Helicobacter pylori antigen, CagA and IgG antibodies (Ab) in the 3 groups of participants**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Group I* ((n = 20))</th>
<th>Group II* ((n = 20))</th>
<th>Group III* ((n = 20))</th>
<th>(P)-value*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
<td>Mean (SD)</td>
<td></td>
</tr>
<tr>
<td><em>H. pylori</em> antigen</td>
<td>0.20 (0.01)</td>
<td>1.10 (0.04)</td>
<td>0.71 (0.03)</td>
<td>&lt; 0.05</td>
</tr>
<tr>
<td><em>H. pylori</em> CagA-Ab</td>
<td>2.2 (0.2)</td>
<td>64.4 (3.2)</td>
<td>52.3 (2.4)</td>
<td>&lt; 0.05</td>
</tr>
<tr>
<td><em>H. pylori</em> IgG-Ab</td>
<td>8.8 (0.5)</td>
<td>58.9 (1.97)</td>
<td>43.6 (1.4)</td>
<td>&lt; 0.05</td>
</tr>
</tbody>
</table>

*ANOVA test:
Group I = control group; group II = patients with *H. pylori*-associated gastritis; group III = patients with stomach cancer.
SD = standard deviation.
tissue damage [33]. In our work, the significant elevation of serum soluble E-cadherin in the cancer group is in agreement with previous work which showed that serum E-cadherin is found in the circulation of normal individuals, but is particularly elevated in patients with gastric carcinoma [34]. Generally, tissue E-cadherin is strongly expressed in well differentiated cancers, but is absent, or markedly reduced in undifferentiated cancers [35], this decrease was mainly observed in diffuse type of gastric cancer and less in intestinal type [36]. These results are supported by our own since we found a significant elevation in serum soluble E-cadherin in the diffuse type of group III compared with the intestinal type (5.5 µg/mL). The elevated level could be an indicator for poor prognosis, metastases and decreased survival [7].

The ROC curve analysis demonstrated that tissue COX-2 activity, serum gastrin and serum soluble E-cadherin have a very important diagnostic possibility for discrimination between people with *H. pylori*-associated gastritis and those with *H. pylori*-associated stomach cancer. The high accuracy of these parameters in prediction of stomach cancer, as shown by a greater area under ROC curve, raises the possibility of their potential benefit as a follow-up test in individuals with *H. pylori* positive gastritis. As E-cadherin showed the highest sensitivity (100%) in discriminating between *H. pylori* gastritis and stomach cancer, we suggest close follow-up together with endoscopic examination for all patients with previous history of *H. pylori*-associated gastritis and serum E-cadherin levels above 5 µg/mL.

Although our findings suggest a potential benefit for measuring E-cadherin in the serum of patients with gastritis, we would need to conduct a larger clinical trial that would extend to include early stomach cancer as well as advanced cancer together with proper staging of the disease in order to obtain a level of confidence that would lead to a change in current clinical practices and outcomes. With a larger trial, we would be able to make more definitive statements regarding the importance of these biochemical parameters and the proper timing for measurement, especially in those patients positive for *H. pylori*. Further research is needed to help planning the follow-up strategy using these biochemical indicators.

**Acknowledgements**

The authors thank Professor El-Sayed Ibrahim Awad, Professor of Surgery, Medical Research Institute, Alexandria University, for his advice and valuable suggestion throughout the work.

**References**


Infectious agents and cancer

Infectious agents are responsible for almost 22% of cancer deaths in the developing world and 6% in industrialized countries. Viral hepatitis B and C cause cancer of the liver; human papilloma virus infection causes cervical cancer; the bacterium *Helicobacter pylori* increases the risk of stomach cancer. In some countries the parasitic infection schistosomiasis increases the risk of bladder cancer and in other countries the liver fluke increases the risk of cholangiocarcinoma of the bile ducts. Preventive measures include vaccination and prevention of infection and infestation.

Further information about WHO’s response to cancer can be found at: http://www.who.int/cancer/en/