

Epidemiology of malaria and predictions of retransmission in Babylon governorate, Iraq

A.A. Al-Ghoury,¹ W.K. El-Hashimi² and J. Abul-Hab³

وبائيات الملاريا واحتمالات معاودة سريانها في محافظة بابل في العراق

عبد الباسط أحمد الغوري، وليد خالد الهاشمي، جليل أبو الحَب

الخلاصة: نجحت جهود وقف سراية الملاريا بعد وباء 1997 – 1998 في محافظة بابل، بالعراق. ومع أن المسح الطفيلي، الذي أُجري في عام 2002، لم يكشف أي حالة ملاريا، إلا أن المسح الحشري أثبت وجود الأنوفيلة الاصطفانية *Anopheles stephensi*، والأنوفيلة البديعة *A. pulcherrimus* بكثافة عالية. وقد سُجِّلَت أعلى معدلات الكثافة في أيلول/سبتمبر، بينما سُجِّلَت أدنى معدلات الكثافة في كانون الأول/ديسمبر وكانون الثاني/يناير. ولكن على الرغم من الكثافة العالية للأنوفيلة، فلم تكتشف أي حيوانات بوغية أو بيضات متكيسة من الطفيلي في البعوض الذي تم تشريحه. ومهما يكن من أمر، فإن سراية الملاريا قد تنبعث من مرقدها، إذا تجاوزت كثافة الأنوفيلة الاصطفانية داخل المنازل الحدود الحرجة، ولم يتم رصد حالات الملاريا الوافدة.

ABSTRACT After the 1997–98 malaria epidemic in Babylon governorate, Iraq, malaria transmission in this area was successfully interrupted. A parasitological survey in 2002 identified no malaria cases but an entomological survey found both *Anopheles stephensi* and *A. pulcherrimus* in high densities. The highest density was recorded in September and the lowest in December and January. Despite the high density of *Anopheles*, no parasite sporozoites or oocysts were found in dissected mosquitoes. Nevertheless, malaria transmission could recur if *A. stephensi* indoor resting density exceeds the critical threshold and imported malaria cases are not monitored.

Épidémiologie du paludisme et prévisions concernant la reprise de la transmission dans le gouvernorat de Babylone (Iraq)

RÉSUMÉ Après l'épidémie de paludisme de 1997-1998 dans le gouvernorat de Babylone (Iraq), la transmission du paludisme dans cette région a pu être interrompue. Lors d'une enquête parasitologique en 2002, aucun cas de paludisme n'a été identifié mais une enquête entomologique a trouvé de fortes densités d'*Anopheles stephensi* et d'*A. pulcherrimus*. La densité la plus forte a été enregistrée en septembre et la plus faible en décembre et janvier. Malgré la forte densité d'anophèles, aucun sporozoïte ou oocyste n'a été trouvé dans les moustiques disséqués. Néanmoins, la transmission du paludisme pourrait reprendre si la densité de *A. stephensi* au repos à l'intérieur des habitations dépassait le seuil critique et si les cas de paludisme importés n'étaient pas suivis.

¹Department of Parasitology, College of Medicine, University of Sana'a, Sana'a, Yemen (Correspondence to A.A. Al-Ghoury: basit_alghoury@yahoo.com).

²Department of Microbiology; ³Department of Public Health, College of Medicine, Al-Mustansiriya University, Yarmouk, Baghdad, Iraq.

Received: 28/03/04; accepted: 06/07/04

Introduction

Malaria is considered one of the greatest challenges of all the health problems in tropical countries. The *Plasmodium* parasite undergoes a complete cycle of sporogony leading to the formation of the infective stages that propagate the infection during the feeding process of the mosquito. The efficiency of transmission of the disease depends mainly on the presence of favourable environmental conditions for the anopheline vectors [1]. Iraq has achieved a dramatic reduction in the number of malaria cases from an estimated 1 million per year in the early 1950s to less than 4000 cases per year by the 1990s.

Recently, the situation has been deteriorating in the 3 north-eastern governorates and malaria has spread outside this area [2]. Babylon governorate has been free of indigenous malaria cases since 1977, but a sudden epidemic occurred during the period 1997–98 in Hilla city, the capital of Babylon governorate [3]. The epidemiological features contributing to the epidemic included: increased rainfall and temperature in Babylon province after 1995 that favoured malaria transmission during that period; agricultural development projects around the city of Hilla that created new mosquito breeding habitats; and population movements when migrant workers returned from endemic areas in the north of Iraq. Control measures at the time included anti-plasmodial measures (treatment of cases, chemoprophylaxis and anti-relapse treatment, i.e. mass treatment); vector control (chemical and biological larvicides/insecticides and spraying with residual insecticides); individual protection aimed at reduction of man–vector contact; establishment of a malaria surveillance programme; and public health education.

The present study in 2002 aimed to evaluate the success of the interruption of

malaria transmission after control measures at the focus of the 1997–98 epidemic. Parasitological and entomological criteria were used to predict the possible recurrence of transmission.

Methods

The study period was 1 year from 1 January to 31 December 2002.

A longitudinal parasitological and entomological survey was carried out in 2 districts: Centre district, which covers Hilla city, the capital of Babylon province and the focus of the 1997–98 epidemic, and the villages of Al-Hashimiyah district. Centre district includes 3 fixed stations, Al-Askery, Al-Zuweer and Offee, whereas Al-Hashimiyah district includes 3 fixed stations, Al-Owidine, Kisher and Al-Jebssah.

Parasitological survey

According to the World Health Organization there are 2 parasitological criteria indicating that transmission has been interrupted at a certain date. First, no resident child born after that date should ever be found positive. Second, careful surveillance with adequate coverage should detect no new cases after that date.

The sample was selected by multi-stage methods and the population of the study was then stratified into 5 groups as follows: malaria cases reported to the Primary Health Care Directorate during 1997–98 ($n = 455$), contacts of malaria cases ($n = 795$), children below 5 years of age from the same area ($n = 314$), foreigners arriving in the province ($n = 24$) and subjects from surrounding areas which had a high mosquito density ($n = 97$). The total for all groups was 1685 individuals. Table 1 indicates the age groups included. As both sexes are usually equally infected, sex was not included as a category.

Table 1 Methods of diagnosis and number of tests used in the malaria parasitological survey in Babylon governorate according to patient age group

Diagnostic method	Age (years)					Total
	0–5	6–10	11–15	16–20	> 20	
Thick & thin blood films	314	246	238	220	667	1685
Haematocrit concentration method	82	57	55	59	99	352
Improved thick blood films	45	26	14	2	24	111
Rapid diagnostic tests	12	16	19	16	39	102
Total	453	345	326	297	829	2250

Blood samples were collected from study subjects and all 1685 individuals were tested by ordinary thick and thin blood smears as this is regarded as the gold standard in malaria diagnosis. Three other diagnostic methods were included to corroborate the smear method: haematocrit concentration ($n = 352$ individuals) [4]; improved thick blood films ($n = 111$) [5]; and rapid diagnostic (dipstick) test ($n = 102$) [6]. Financial constraints did not allow the same tests to be used on all samples. All thick and thin blood films were double checked by the general health laboratory of Primary Health Care Directorate in Babylon governorate.

Entomological survey

For the entomological survey, 6 fixed stations in the study areas were visited every month to carry out malaria vector surveillance in these areas. The vectors were monitored at both larval and adult stages from various habitats. The sucking tube collection method was used for outdoor and semi-indoor resting mosquitoes, but pyrethrum space spray was used for indoor resting mosquitoes.

In each station, 5 rooms were searched by 1 person who spent half an hour using an aspirator and a torch light. Also 5 rooms were searched by the pyrethrum space spray

method in the morning [7]. In each station 10 suspected breeding places were searched and 10 dips were made in each place.

The samples of adult mosquitoes and larvae were examined and identified according to the keys described by Abul-Hab [8]. In addition, the salivary glands and stomach of the captured female mosquitoes were dissected for the detection of malaria sporozoites and oocysts [7].

Prediction of malaria epidemics

Entomological survey and indoor resting density were used for this purpose as well as entomological records before, during and after the 1997–98 epidemic. *A. stephensi* is the major malaria vector in central and southern Iraq [18], so that the indoor resting density of this species was used in a theoretical trial for predicting a malaria epidemic. Two methods were used to determine whether the household indoor resting density of *A. stephensi* has exceeded critical levels associated with epidemic transmission: the direct approach and the minimum sample size approach [9].

Statistical analysis

The results were presented in simple tables and graphs, using simple statistical measures as mean and variance as well as regres-

sion. $P < 0.05$ was considered statistically significant.

The following measures were calculated for the purpose of prediction of malaria epidemics:

1. Taylor's power law, to measure the relationship between variance (s) and mean (x) as a power function [10].

$$s^2 = ax^{-b}$$

parameters a and b are Taylor's power law coefficients and provide information on insect aggregation.

2. An equation for calculating the number of houses required to estimate the mean *Anopheles* density for selected allowable errors using the direct approach [11].

$$n = \left(\frac{Z_{a/2}}{AE} \right)^2 ax^{-(b-2)}$$

where $Z_{a/2} = 1.96$ at $P = 0.05$

n = number of houses required to calculate mean *Anopheles* density.

AE = allowable error, typically set between 10% and 50% of the mean.

3. An equation for calculating the number of houses required to estimate the mean *Anopheles* density using the minimum sample size approach [9].

$$n = \frac{\ln(0.05)}{\ln(P_{(o)})}$$

where 0.05 = error rate

$$P_{(o)} = \left(1 + \frac{ax^{-b} - \bar{x}}{\bar{x}} \right)^{\left(\frac{\bar{x}^2}{ax^b - \bar{x}} \right)}$$

Results

Parasitological survey

A total of 2250 samples were tested from the 1685 individuals (Table 1). Although more than 1 diagnostic method was used in this study, no malaria cases were found using any of the 4 diagnostic tests. It is important to note that no cases were found among children below 5 years of age who were born after the epidemic of 1997–98.

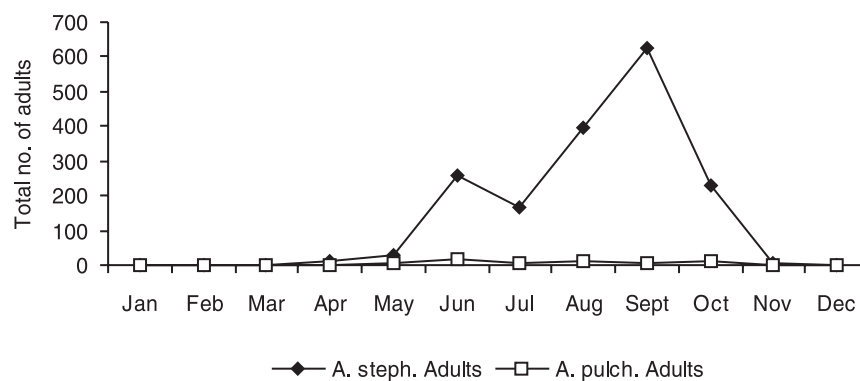
Entomological survey

The entomological survey found that the 2 main malaria vectors in central and southern Iraq (*A. stephensi* and *A. pulcherrimus*) were present at the adult and larval stages.

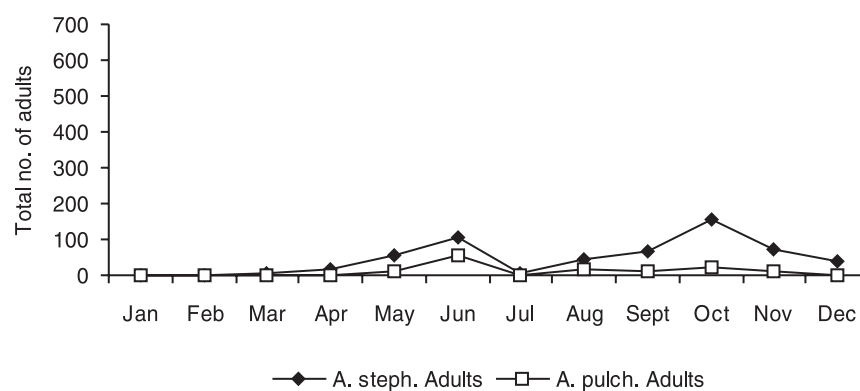
The findings from all 6 fixed stations were grouped together according to the method of collection of the adults and larvae by month of collection (Figure 1). *A. stephensi* adults were present during all months of the year except January, whereas *A. pulcherrimus* adults were present only during May to November. The highest number of both *A. stephensi* and *A. pulcherrimus* adults were found in June and September/October respectively. The larval stages of *A. stephensi* disappeared in December and January when the mean temperature was below 14 °C. *A. pulcherrimus* larvae appeared in May and continued to be present until November. Two peaks of the *Anopheles* larvae were seen: the first in June and the second in October.

A small number ($n = 85$) of captured *Anopheles* adults were dissected for malaria oocysts and sporozoites. Neither oocysts nor sporozoites were found in either *A. stephensi* or *A. pulcherrimus*.

a) Total adults found by sucking tube method



b) Total adults found by pyrethrum spray method



c) Total larvae found

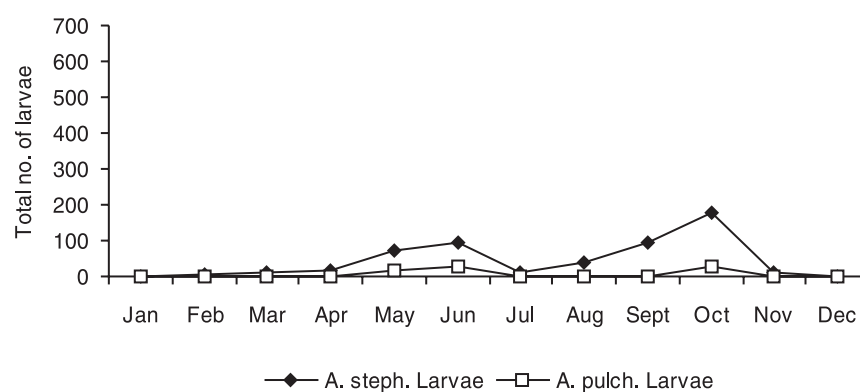


Figure 1 Annual numbers of *Anopheles stephensi* and *Anopheles pulcherrimus* species in Babylon governorate (Al-Hashimiyah and centre districts) according to collection method and month

Prediction of malaria epidemics

The total number of *A. stephensi* caught in 6 stations from January to December 2002 was 1731 mosquitoes by pyrethrum spray collection method. This number was pooled into 1 sample, giving a mean 9.2 of mosquitoes per room (Table 2). The malaria annual reports in 1997–98 indicated that the epidemic began in October 1997 and peaked in January 1998 [3]. During the pre-epidemic months of August and September, the mean *Anopheles* density ranged from 0 to 11.2 mosquitoes per room. After the incidence of malaria began to decline in February, the mean *Anopheles* density ranged from 0 to 1.6 mosquitoes per room. An indoor resting density of 6 mosquitoes per room was chosen as the critical density indicating epidemic risk, because this value appeared to provide good separation between pre-

and post-epidemic periods. Conversely, 1 mosquito per room was chosen as a normal density expected during non-epidemic periods. Because we were interested in periods of elevated *Anopheles* density, only those sampling periods with a mean *Anopheles* density > 1 mosquito per room were used to determine Taylor's power law coefficients. The number of *A. stephensi* caught in 6 stations over 7 sampling periods (April to October 2002) were pooled into 1 sample with mean of 16.5 mosquitoes per room (95% confidence interval, 14.4–18.6). This was significantly greater than the mean *Anopheles* density in 1997–98, 8.1 mosquitoes per room.

Using the 7 sampling periods in which the mean *A. stephensi* density was > 1 mosquito per room, regression of [log (sampling period variance)] on [log (sampling period mean)] indicated a strong positive correlation ($r = 0.98$, $P = 0.05$). Estimates for the parameters a and b were 0.16 and 2.36 respectively, indicating aggregation in the data.

Table 2 Indoor resting density of *Anopheles stephensi* during the study period in Babylon governorate (Al-Hashimiyah and centre district)

Month	<i>A. steph.</i> (No.)	Sampled rooms (No.)	Indoor resting density (No./room)
Jan	0	15	0
Feb	1	15	0.1
Mar	2	10	0.2
Apr	14	10	1.4
May	30	12	2.5
Jun	260	15	17.3
Jul	164	13	12.6
Aug	397	18	22.1
Sept	625	18	34.7
Oct	230	18	12.8
Nov	8	21	0.4
Dec	0	24	0
Total	1731	189	9.2

$r = 0.98$.

Direct approach

The number of houses required to calculate mean *Anopheles* density with allowable errors ranging from 10% to 50% of the mean was calculated using equation 2 with estimates of a and b as determined above. Samples of 115, 29, 13, 7 and 5 houses would be needed to calculate mean mosquito densities with allowable errors of 10%, 20%, 30%, 40%, and 50% respectively (Figure 2).

Minimum sample size approach

The number of houses required to find at least 1 *A. stephensi* according to the mean *Anopheles* density was calculated using equation 3 and the parameters a and b as determined above. Samples of only 1 house would be needed to calculate mean mosquito density with an error rate of 0.05%.

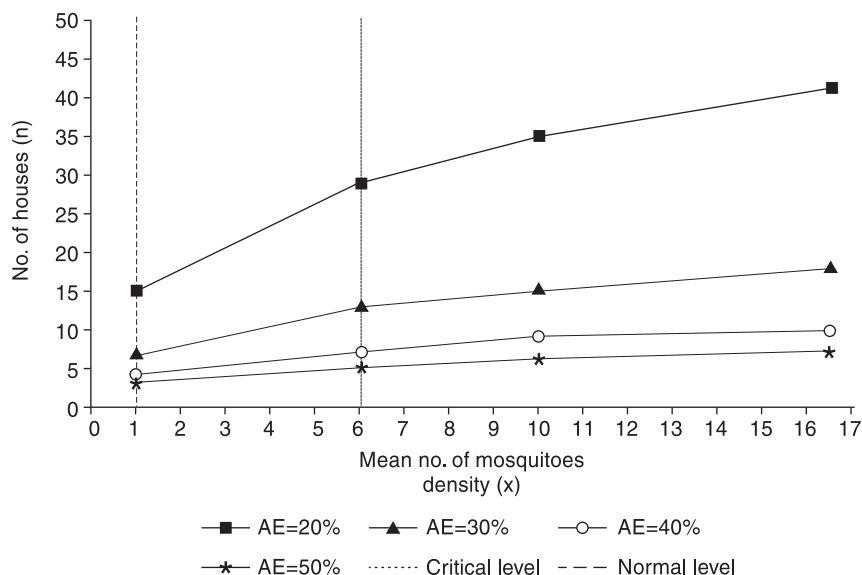


Figure 2 Direct approach operating curve (AE = allowable error)

Failure to find any mosquitoes in a sample of 1 house would indicate that *Anopheles* spp. density was not yet above the critical threshold.

Using the above 2 methods revealed that *A. stephensi* density had exceeded the critical threshold and indicated there was a risk of an epidemic (Figure 2).

Discussion

In the parasitological survey the total number of blood films examined was higher than other diagnostic methods, because this method is considered the gold standard in malaria diagnosis in Iraq and other countries [12,13]. Blood smears were collected from all age groups, but were carefully focused on children below 5 years of age who were born after the epidemic of 1997–98. All of their results were negative. The World

Health Organization has 2 parasitological criteria indicating that transmission has been interrupted at a certain date. First, no resident child born after that date should ever be found positive. Second, careful surveillance with adequate coverage should detect no new cases after that date [14,15]. Similar studies in central and southern areas of Iraq [16] and in Nineveh governorate in 1998–2000 (G. Al-Mukhtar, in press) found that malaria transmission had been interrupted and was under control.

Northern Iraq belongs to the palaearctic region of the zoogeographical zones while its central and southern regions belong to the oriental zone and the annual occurrence of mosquito fauna differs accordingly between these regions. The annual occurrence of *A. stephensi* and *A. pulcherrimus* recorded by the entomological survey in the present study agree closely with the find-

ings of Abul-Hab and Kassal [17] as well as Macan [18]. Our survey was extended to neighbouring districts from the original epidemic to check for the presence of the vectors. This is necessary now because Iraq is experiencing a re-emergence of the *A. stephensi* population after it had been well controlled since the 1960s [19].

It was found that the *A. stephensi* population was more abundant than *A. pulcherrimus*. *A. stephensi* is regarded as the only important vector of malaria in the central and southern regions [18]. There were 2 peaks of *Anopheles* species, larvae and adults, the first in June and the second in October. These findings agreed with the findings of Macan [18] and Abul-Hab and Kassal [17]. It is important to note that while both malaria oocysts and sporozoites were detected in these other studies, no such stages were found in the dissected specimens of either *A. stephensi* or *A. pulcherrimus* in this study.

Malaria transmission in Iraq is regarded as unstable in the whole country [20]. Therefore, it is vital to develop the tools to predict epidemics. In Iraq, increased *Anopheles* densities are not always associated with an epidemic but could be used as an indicator of epidemic risk. *A. stephensi* is the major malaria vector in the central and southern regions of Iraq. Indoor resting *A. stephensi* density was used as an indicator of epidemic risk when its density exceeded the critical level. Both the critical and normal thresholds were determined from the entomological data before, during and after the epidemic of 1997–98 [3]. We chose 6 mosquitoes per room as the critical threshold indicating the risk of an epidemic because it appeared to adequately separate mean *A. stephensi* density associated with epidemic transmission from the

normal threshold during the post-epidemic period of 1997–98. Similarly, 1 mosquito per room was chosen as a normal threshold of *A. stephensi* density that was not associated with the risk of an epidemic. It was observed that closely similar thresholds of *A. stephensi* densities were found and recorded by the Communicable Disease Centre Baghdad, i.e. 5 mosquitoes per room was regarded as a critical level and below this value was regarded as a normal level [3]. In our study, both the direct approach and the minimum sample size approach were used for determining whether *A. stephensi* density had exceeded a critical level [9]. In the direct approach, it was demonstrated that calculation of mean *Anopheles* density with a high degree of precision requires a large number of houses, because of the variance in *Anopheles* density among houses. The number of houses required to sample would range from 5 to 115 houses for allowable errors of 50%–10% of the mean respectively. The mean *A. stephensi* density of 16.5 mosquitoes per room exceeded the critical level and indicated epidemic risk. In the minimum sample size approach, a mean density of 1 mosquito/house was required to determine whether *A. stephensi* density had exceeded the critical level. Less than this would indicate that the critical level had not been reached. However, it could be used as an initial screening tool during periods when *Anopheles* density was expected to be increasing. If a mean *Anopheles* density of 1 mosquito/house was found, monitoring could continue using the direct approach. A similar study in the African Highlands [9] found that it is feasible and probably appropriate to include monitoring of *Anopheles* spp. density in the prediction of malaria epidemics.

This study concludes that malaria transmission in the Babylon governorate was successfully interrupted after the recent epidemic of 1997–98. However, malaria transmission could recur if *A. stephensi* indoor resting density exceeds the critical threshold and imported malaria cases are not monitored. We propose that malaria epidemics should be predicted by measuring the indoor resting mosquito density.

Acknowledgements

We thank Dr Jalil Al-Zudaïdi, Director of Endemic Diseases Institute, Baghdad, for his assistance and all the staff of the malaria unit Primary Health Care Directorate, Babylon Governorate for their continuous help during most stages of the fieldwork of this study.

References

1. Radford A J, Van Leeuwen H, Christian SH. Social aspects in the changing epidemiology of malaria in the highlands of New Guinea. *Annals of tropical medicine and parasitology*, 1976, 70:11–23.
2. Roll back malaria. *Epidemiological situation. Iraq*. Online fact sheet. Cairo, World Health Organization Eastern Mediterranean Regional Office (<http://www.emro.who.int/rbm/EpidemiologicalSituation-CountryProfiles.htm>, accessed 30 October 2005).
3. *Annual reports of malaria in Iraq*. Baghdad, Iraq, Communicable Disease Centre, Ministry of Health, 1997–98.
4. Worth MR. The heparinized capillary tube as an epidemiologic tool. II. concentration of blood parasites by centrifugation. *American journal of hygiene*, 1964, 80:70–4.
5. Al-Khairi KS. An improved method of preparing thick blood films for the examination of malaria parasites. *Saudi medical journal*, 1992, 13:542–5.
6. *Malaria diagnosis—new perspectives. Report of a joint WHO/USAID informal consultation, 25–27 October 1999*. Geneva, World Health Organization, 2000 (WHO/CDS/RBM/2000).
7. *Manual on practical entomology in malaria prepared by the WHO division of malaria and other parasitic diseases*. Geneva, World Health Organization, 1975.
8. Abul-Hab J. *Medical and veterinary entomology in Iraq*. Baghdad, Iraq, Baghdad University, 1979.
9. Lindblade KA, Walker ED, Wilson ML. Early warning of malaria epidemics in African highlands using *Anopheles* (Diptera: Culicidae) indoor resting density. *Journal of medical entomology*, 2000, 37: 664–74.
10. Taylor LR. Aggregation, variance and the mean. *Nature*, 1961, 189:732–5.
11. Wilson LT, Room PM. Clumping patterns of fruit and arthropods in cotton, with implications for binomial sampling. *Environmental entomology*, 1983, 12:50–4.
12. Ossi GT. Malaria in Iraq from 1980 to 1983. *Bulletin of endemic diseases*, 1984, 24:5–23.
13. Gilles HM, Warrell DA, eds. *Bruce-Chewatt's essential malariology*, 3rd ed. London, Edward Arnold, 1993.
14. *WHO Expert Committee on Malaria [meeting held in Geneva from 21 to 27 September 1965]: twelfth report*. World Health Organization, Geneva, 1966 (Technical Report Series, No. 324).
15. *WHO Expert Committee on Malaria [meeting held in Geneva from 19 to 30 October 1965]: thirteenth report*. World Health Organization, Geneva, 1966 (Technical Report Series, No. 344).

- 1970]: *fifteenth report*. Geneva World Health Organization, 1971 (Technical Report Series, No. 467).
16. Shihab KI et al. Immunological and parasitological survey in areas of Iraq where malaria transmission has been interrupted since several years. *Bulletin of endemic diseases*, 1987, 28:17–28.
 17. Abul-Hab J, Kassal, S. Impact of anti-malaria spraying on the occurrence of Anopheles (Diptera: Culicidae) in Iraq. *Bulletin of endemic diseases*, 1986, 27:37–51.
 18. Macan TT. The anopheline mosquitoes of Iraq and North Persia. In: Leeson HS et al., eds. *Anopheles and malaria in the Near East (Memoirs of the London School of Hygiene and Tropical Medicine, No. 7)*. London, HK Lewis, 1950.
 19. *Annual reports of malaria in Iraq*. Baghdad, Iraq, Communicable Disease Centre, Ministry of Health, 1960–2001.
 20. Al-Kafajei A, Ahmed KJ. Effectiveness of malaria control programs in Nineveh 1992. *Journal of community medicine, Baghdad*, 1993, 6:135–42.

Malaria control in complex emergencies

The areas of the world whose populations are most affected by complex emergencies are often those with the greatest malaria burden. Consequently, malaria is a significant cause of death and illness in complex emergency situations. This inter-agency handbook focuses on effective malaria control responses to complex emergencies, particularly during the acute phase when reliance on international humanitarian assistance is greatest. It provides policy-makers, planners, field programme managers and medical coordinators with practical guidance on designing and implementing measures to reduce malaria morbidity and mortality. Such measures must address the needs of both the displaced and the host populations and must accommodate the changes in those needs as an acute emergency evolves into a more stable situation. A glossary is provided at the beginning of the handbook and suggestions for further reading are included at the end of several chapters.

Malaria control in complex emergencies. An inter-agency field handbook (ISBN 92 4 159389) is available at the WHO bookshop at (<http://www.who.int/publications/en/>)